

R資料視覺化 基礎統計圖形

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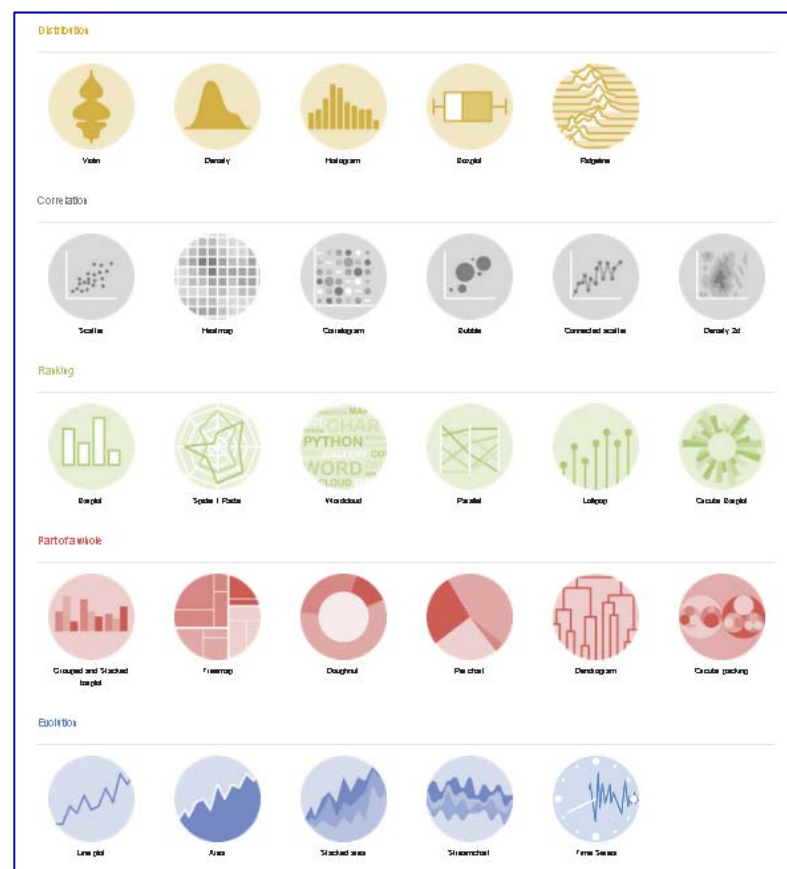
<http://www.hmwu.idv.tw>

本章大綱與學習目標

- 簡介: Why Data Visualization? 圖形輸出、向量圖與非向量圖格式
- 客製化圖形: R圖形座標系統、圖標題和範圍、abline、lines、arrows、segments 一頁多張圖形、顏色、文字標號、Data Symbols、圖例說明、座標軸、讀取外部影像檔案
- 基本統計圖形: 索引圖、直方圖、Density Plots、Quantile Plots、圓餅圖、散佈圖、curve、盒形圖、長條圖、散佈圖矩陣、時間序列、多邊形、Radar Plot、3D透視圖、3D散佈圖、rgl、影像。
- 其它: pheatmap、networkD3、Sankey Diagram、Word Cloud

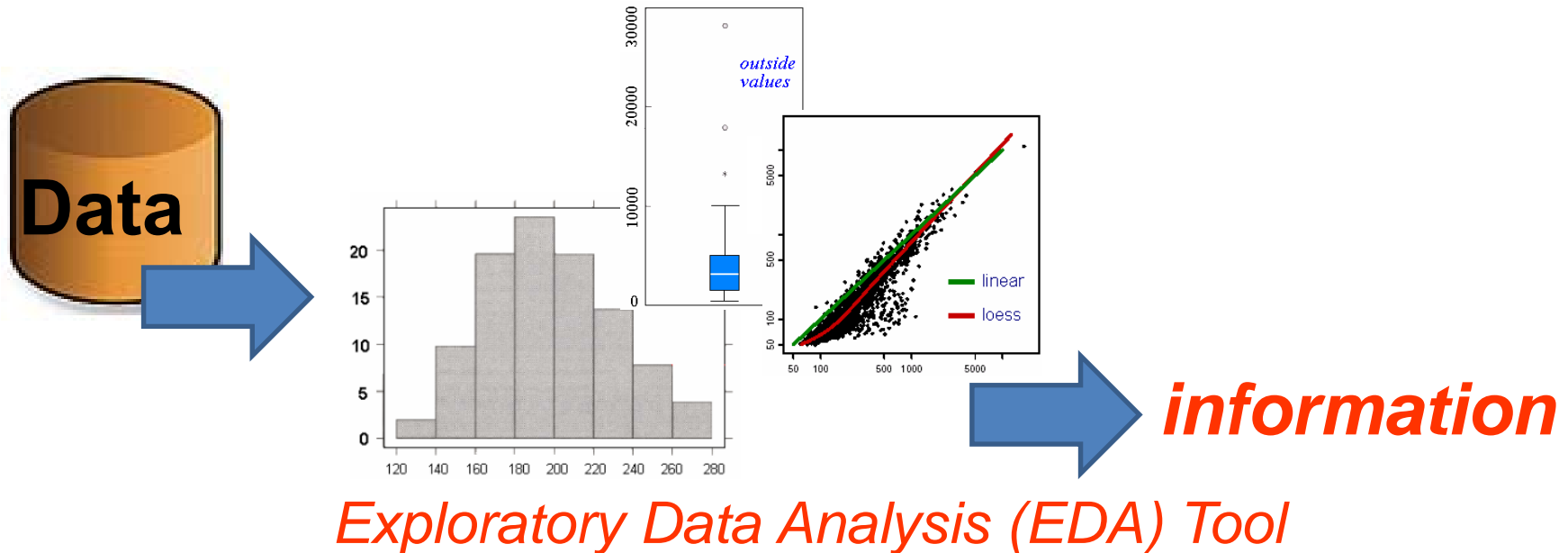
The R Graph Gallery

<https://r-graph-gallery.com/>



Graphical Methods

The purpose of statistical graphics is to provide **visual** representations of **quantitative** information.



Statistical graphics comprise

a set of **strategies and techniques** that provide the research with important **insights** about the data under examination and help guide the subsequent steps of the research process.

Why Data Visualization?

- It is not about "**infographics**", the beautiful, heavily customized products of expert graphic designers.
- Data visualization can provide clear understanding of patterns in data, detect hidden structures in data, condense information.
- Anscombe's quartet** comprises four datasets. They were constructed in 1973 by the statistician Francis Anscombe to demonstrate both the importance of graphing data before analyzing it and the effect of outliers on statistical properties.
- Four datasets have nearly identical simple statistical properties, yet appear very different when graphed.

	I		II		III		IV	
	<i>x</i>	<i>y</i>	<i>x</i>	<i>y</i>	<i>x</i>	<i>y</i>	<i>x</i>	<i>y</i>
1	10	8.04	10	9.14	10	7.46	8	6.58
2	8	6.95	8	8.14	8	6.77	8	5.76
3	13	7.58	13	8.74	13	12.74	8	7.71
4	9	8.81	9	8.77	9	7.11	8	8.84
5	11	8.33	11	9.26	11	7.81	8	8.47
6	14	9.96	14	8.1	14	8.84	8	7.04
7	6	7.24	6	6.13	6	6.08	8	5.25
8	4	4.26	4	3.1	4	5.39	19	12.5
9	12	10.84	12	9.13	12	8.15	8	5.56
10	7	4.82	7	7.26	7	6.42	8	7.91
11	5	5.68	5	4.74	5	5.73	8	6.89

Mean of x in each case: **9** (exact)

Sample variance of x in each case: **11** (exact)

Mean of y in each case: **7.50** (to 2 decimal places)

Sample variance of y in each case: **4.122** or **4.127** (to 3 decimal places)

Correlation between x and y in each case: **0.816** (to 3 decimal places)

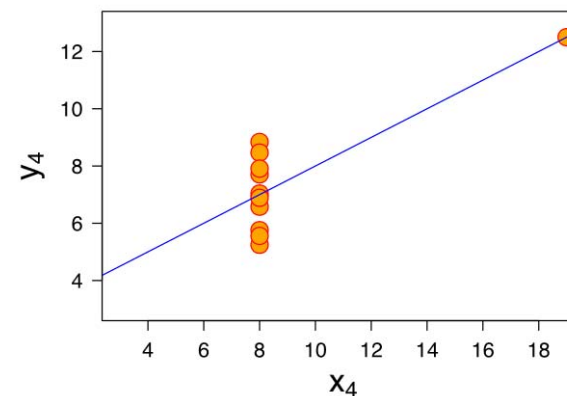
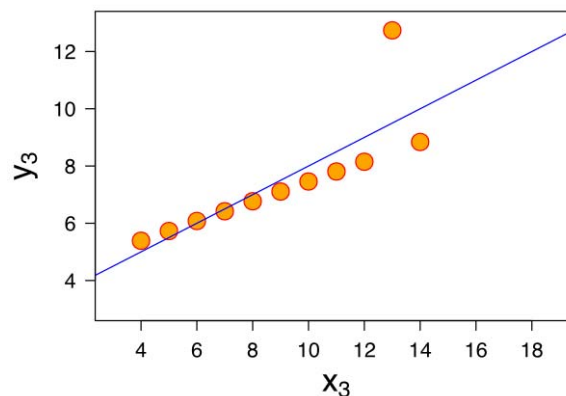
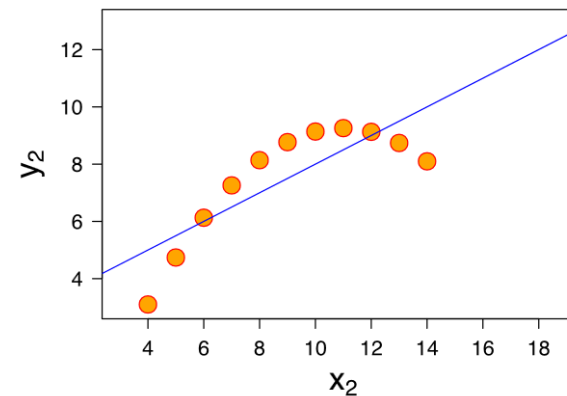
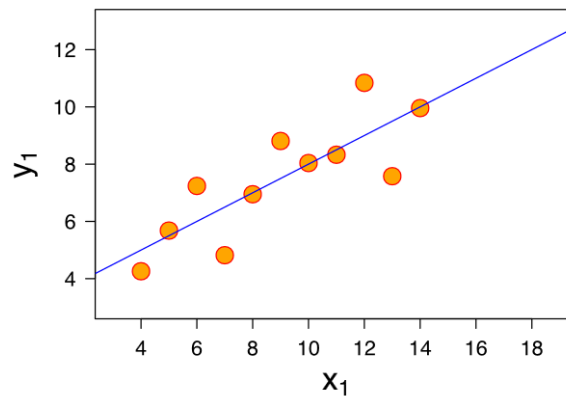
Linear regression line in each case: **$y = 3.00 + 0.500x$** (to 2 and 3 decimal places, respectively)

https://en.wikipedia.org/wiki/Anscombe's_quartet

<http://ryanwomack.com/IASSIST/DataViz/>

Anscombe's Quartet

- Mean of x in each case: 9 (exact)
- Sample variance of x in each case: 11 (exact)
- Mean of y in each case: 7.50 (to 2 decimal places)
- Sample variance of y in each case: 4.122 or 4.127 (to 3 decimal places)
- Correlation between x and y in each case: 0.816 (to 3 decimal places)
- Linear regression line in each case: $y = 3.00 + 0.500x$ (to 2 and 3 decimal places, respectively)





Anscombe's Quartet of 'Identical' Simple Linear Regressions

```

> head(anscombe, 3)
  x1 x2 x3 x4  y1  y2  y3  y4
1 10 10 10  8  8.04 9.14  7.46  6.58
2  8  8  8  8  6.95 8.14  6.77  5.76
3 13 13 13  8  7.58 8.74 12.74  7.71
> apply(anscombe, 2, mean)
  x1      x2      x3      x4      y1      y2      y3      y4
9.000000 9.000000 9.000000 9.000000 7.500909 7.500909 7.500000 7.500909
> apply(anscombe, 2, sd)
  x1      x2      x3      x4      y1      y2      y3      y4
3.316625 3.316625 3.316625 3.316625 2.031568 2.031657 2.030424 2.030579
> mapply(cor, anscombe[,1:4], anscombe[,5:8])
  x1      x2      x3      x4
0.8164205 0.8162365 0.8162867 0.8165214
> mapply(function(x, y) lm(y~x)$coefficients, anscombe[, 1:4], anscombe[, 5:8])
      x1      x2      x3      x4
(Intercept) 3.0000909 3.000909 3.0024545 3.0017273
x          0.5000909 0.500000 0.4997273 0.4999091

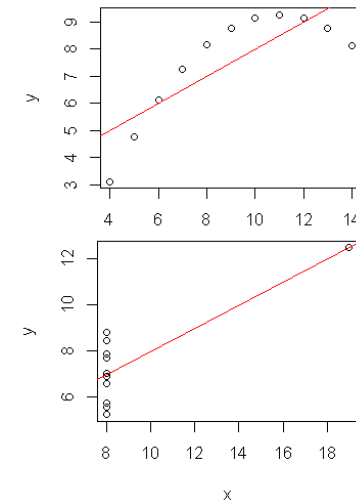
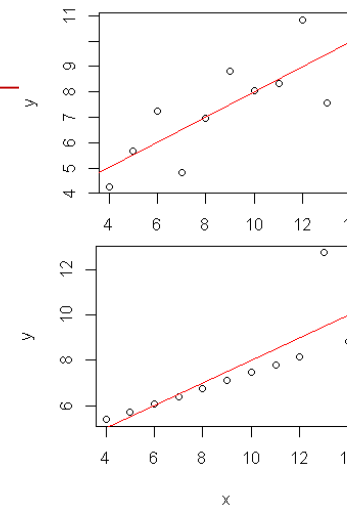
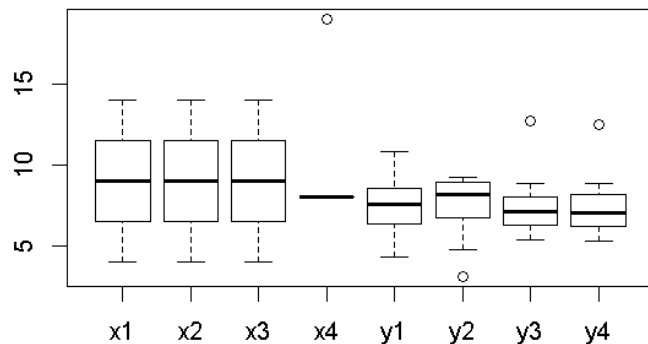
```

```

par(mfrow=c(2, 2))
regplot <- function(x, y){
  plot(y~x)
  abline(lm(y~x), col="red")
}
mapply(regplot, anscombe[, 1:4], anscombe[, 5:8])

```

boxplot(anscombe)





Example

RGui

方法1: Save as ...

```
> plot(iris[,1])
```

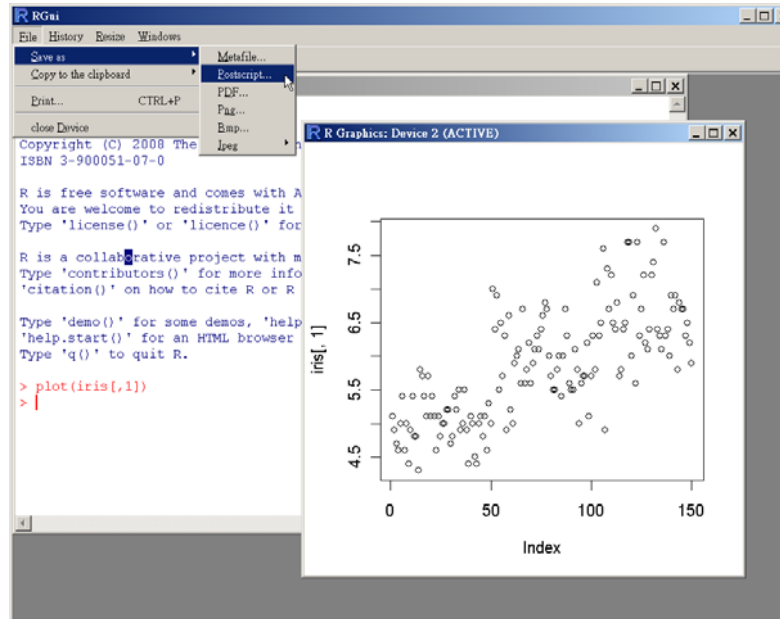
方法2: Activate device

```
> postscript(file="iris2.ps")
> plot(iris[,2])
> dev.off()
```

or

```
> jpeg(file="iris3.jpg")
> plot(iris[,3])
> dev.off()
```

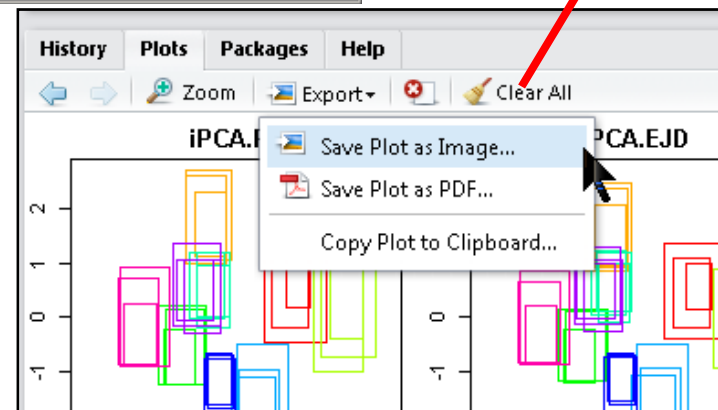
```
# Graphics Device:
bmp, jpeg, png, tiff, pdf
```



注意圖形的比例及大小

```
> dev.off()
```

RStudio



向量圖與非向量圖格式

存檔格式及目的

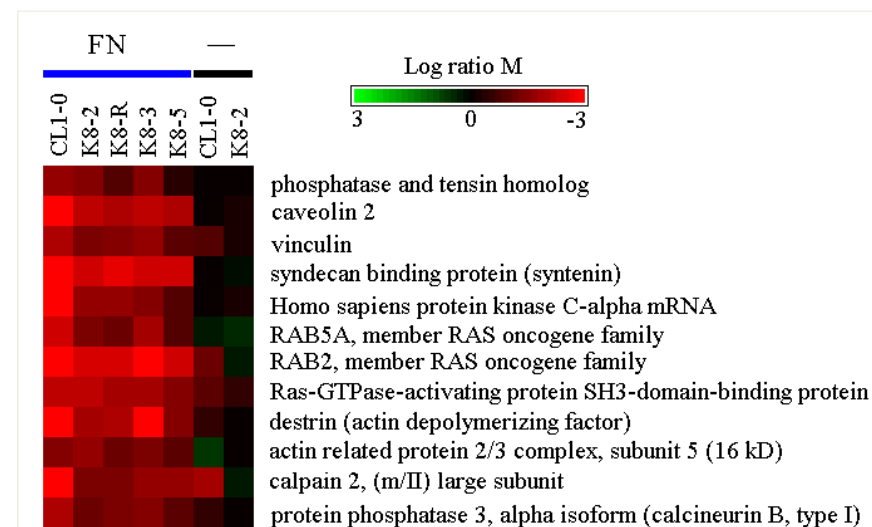
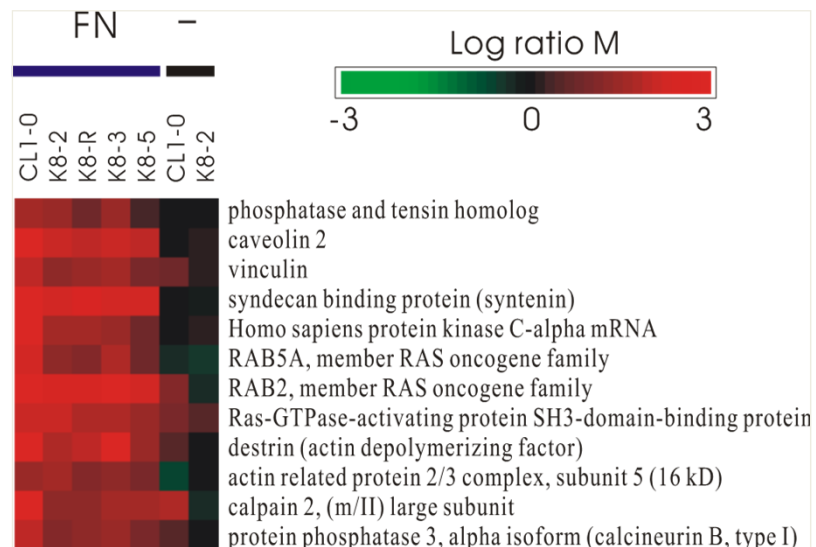
- 點陣圖檔 (壓縮失真: PNG, JPEG, TIFF, BMP) : web page, powerpoint, word,...
- 向量圖檔 (物件不失真: Metafile, SVG, EPS) : Publication, LaTeX.

向量圖(在此為wmf格式):

可拉大或縮小不失真。
此時在螢幕看起來可能會有點糊(平滑化結果)，其實列印出來會很清楚的。

點陣圖(在此為bmp格式):

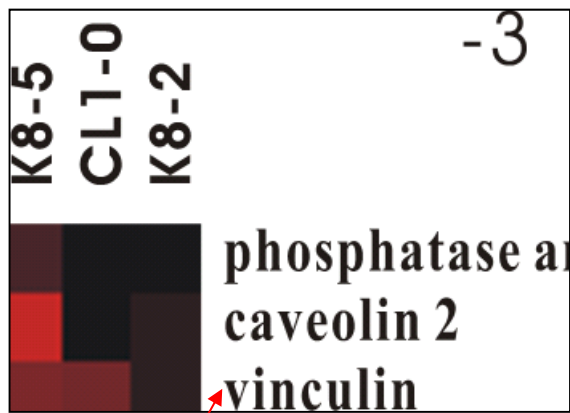
與向量圖(在螢幕上看的)最大差別是拉大縮小有明顯的鋸齒狀。此圖插入powerpoint後會被平滑化，所以看起來會霧霧的。



如何放大圖片:

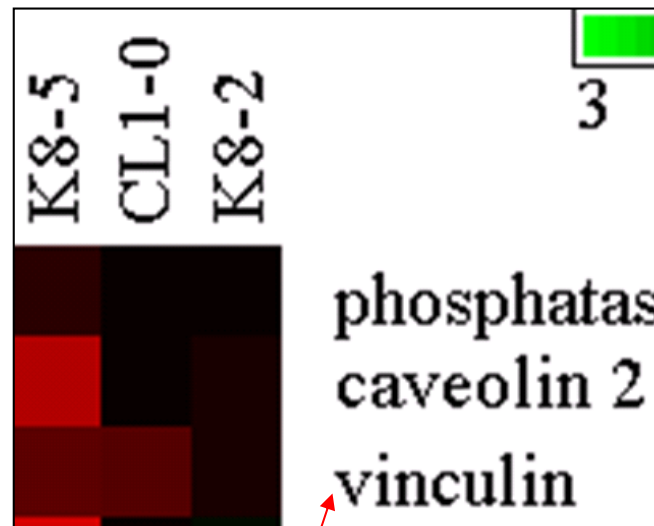
1. 選按圖片 => 滑鼠右鍵 => 設定圖格式 => 大小 => 縮放比例 => 高度設為300% => OK
2. 這跟放大投影片(或全螢幕)成300%是不一樣的。

向量圖放大變這樣:



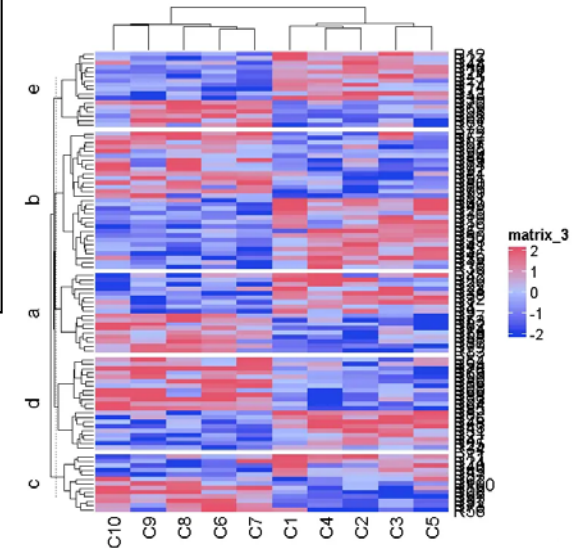
無鋸齒狀, 邊界會模糊的(或霧霧的是軟體自動平滑化的結果)

點陣圖放大變這樣:

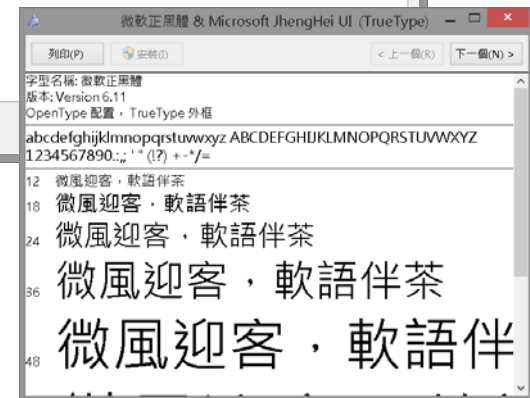
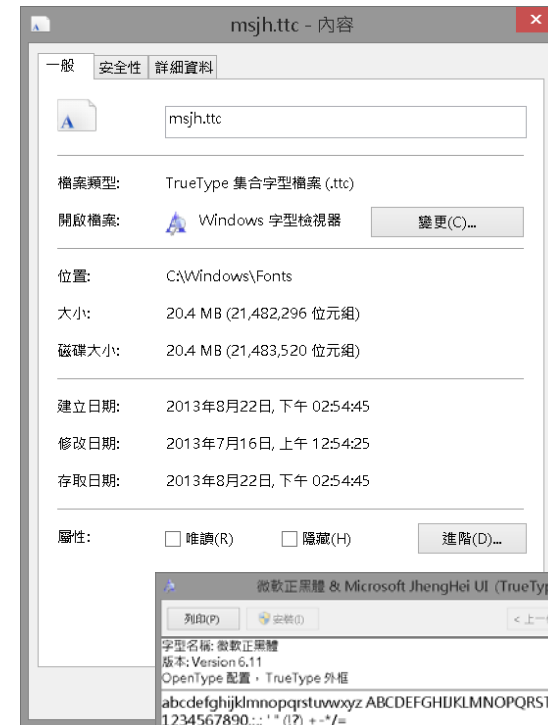
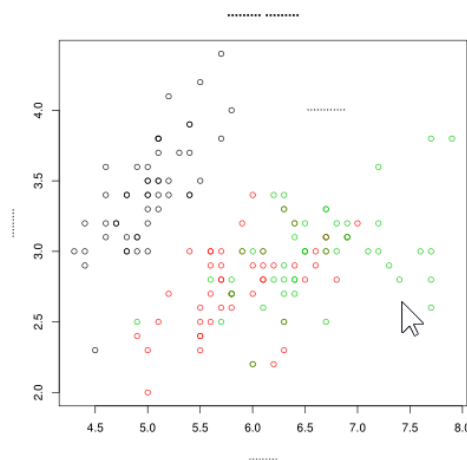
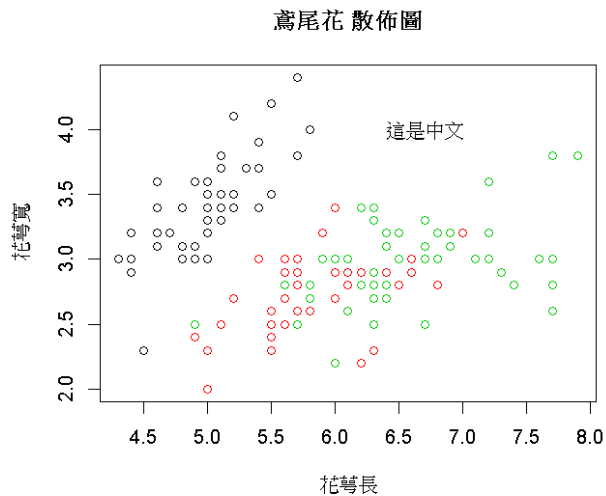


有鋸齒狀

R畫的圖很醜?



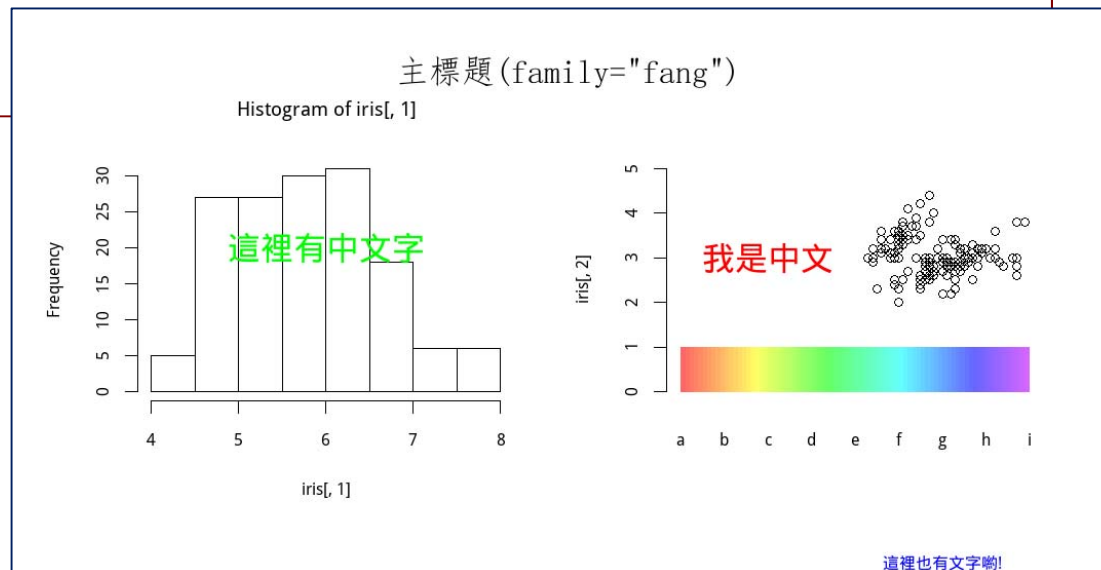
```
> pdf("iris.pdf") # jpeg, bmp OK
> plot(iris[, 1:2], xlab="花萼長", ylab="花萼寬", main="鳶尾花 散佈圖", col=iris[,5])
> text(6.7, 4, "這是中文")
> dev.off()
```



```
> #showtext: Using Fonts More Easily in R Graphs
> library(showtext)
> font_add("msjh", "msjh.ttc") ## 微軟正黑體
> showtext_auto(enable=TRUE)
```

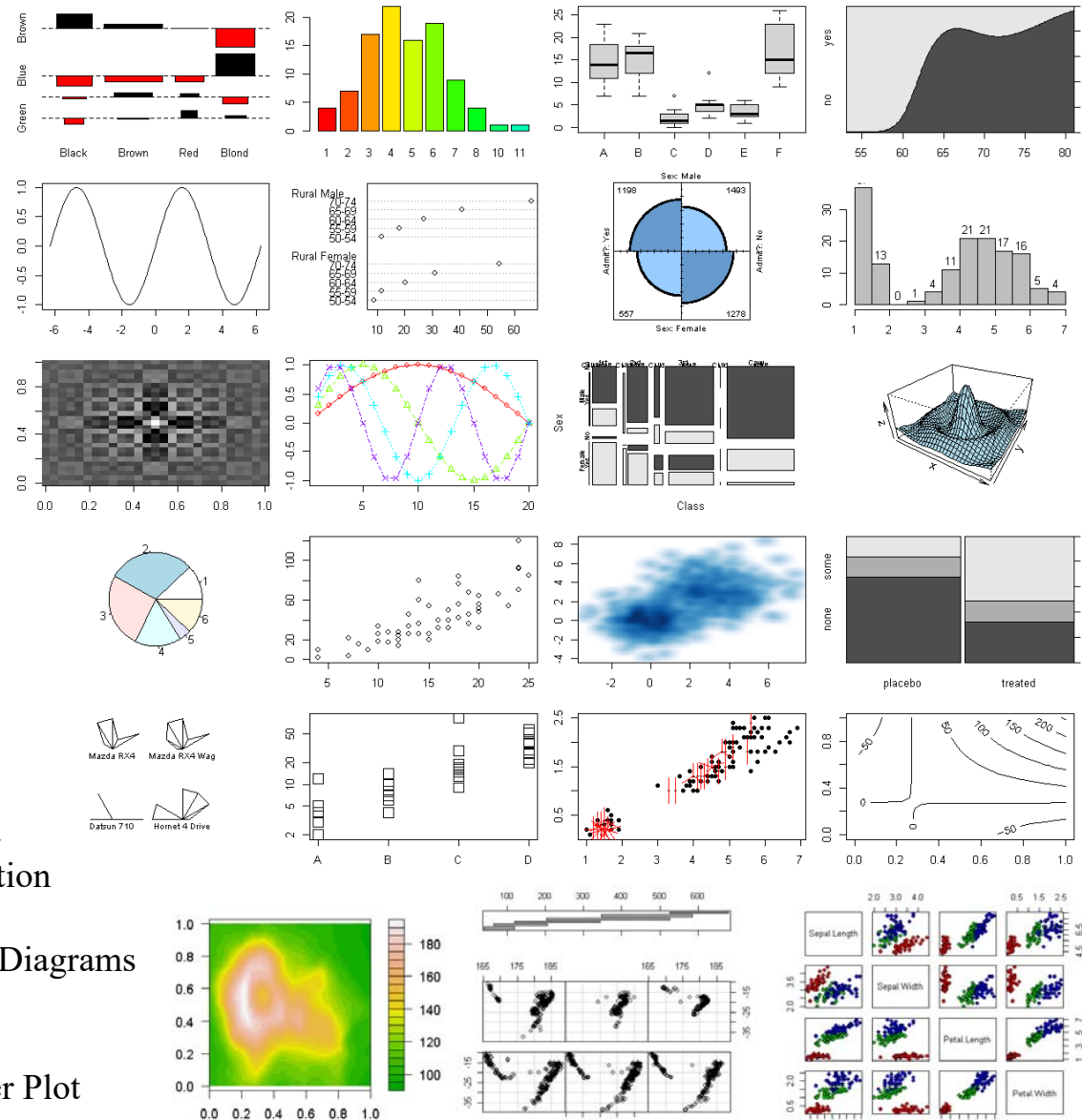
```

> pdf("test.pdf", width=12, height=8)
> # ?bmp (jpeg, png, tiff)
> par(family="msjh") ## 微軟正黑體
> # outer: 兩個圖對中間
> # line=-2 與圖形之空格
> par(mfrow=c(1,2), oma=c(2, 1, 2, 1))
> hist(iris[, 1])
> text(6, 20, "這裡有中文字", col="green", cex=2)
> plot(iris[, 1], iris[, 2], xaxt="n", bty="n", xlab="", xlim=c(0, 8), ylim=c(0, 5))
> axis(1, at=0:8, labels=letters[1:9], tick=FALSE)
> text(2, 3, "我是中文", col="red", cex=2)
> title('主標題(family="fang")', family="fang", outer=TRUE, line=-1, cex.main=2)
> mtext("這裡也有文字啲!", side=1, line=6, at=6, col="blue")
> mycolor <- rainbow(100, alpha=0.6)[1:80]
> rasterImage(t(mycolor), 0, 0, 8, 1, interpolate=FALSE)
> dev.off()
windows
    2
    
```



Plots:

- assocplot**: Association Plots
- barplot**: Bar Plots
- boxplot**: Box Plots
- cdplot**: Conditional Density Plots
- contour**: Display Contours
- coplot**: Conditioning Plots
- curve**: Draw Function Plots
- dotchart**: Cleveland's Dot Plots
- filled.contour**: Level (Contour) Plots
- fourfoldplot**: Fourfold Plots
- hist**: Histograms
- image**: Display a Color Image
- matplot**: Plot Columns of Matrices
- mosaicplot**: Mosaic Plots
- pairs**: Scatterplot Matrices
- persp**: Perspective Plots
- pie**: Pie Charts
- plot**: Generic X-Y Plotting
- smoothScatter**: Scatterplots with Smoothed Densities Color Representation
- spineplot**: Spine Plots and Spinograms
- stars**: Star (Spider/Radar) Plots and Segment Diagrams
- stem**: Stem-and-Leaf Plots
- stripchart**: 1-D Scatter Plots
- sunflowerplot**: Produce a Sunflower Scatter Plot





Decoration:

abline: Add Straight Lines to a Plot
arrows: Add Arrows to a Plot
axis.POSIXct: Date and Date-time Plotting Functions
axis: Add an Axis to a Plot
box: Draw a Box around a Plot
grid: Add Grid to a Plot
legend: Add Legends to Plots
lines: Add Connected Line Segments to a Plot
matlines: Plot Columns of Matrices
matpoints: Plot Columns of Matrices
mtext: Write Text into the Margins of a Plot
panel.smooth: Simple Panel Plot
points: Add Points to a Plot
polygon: Polygon Drawing
polypath: Path Drawing
rasterImage: Draw One or More Raster Images
rect: Draw One or More Rectangles
rug: Add a Rug to a Plot
segments: Add Line Segments to a Plot
symbols: Draw Symbols (Circles, Squares, Stars, Thermometers, Boxplots)
text: Add Text to a Plot
title: Plot Annotation
xspline: Draw an X-spline

Utilities:

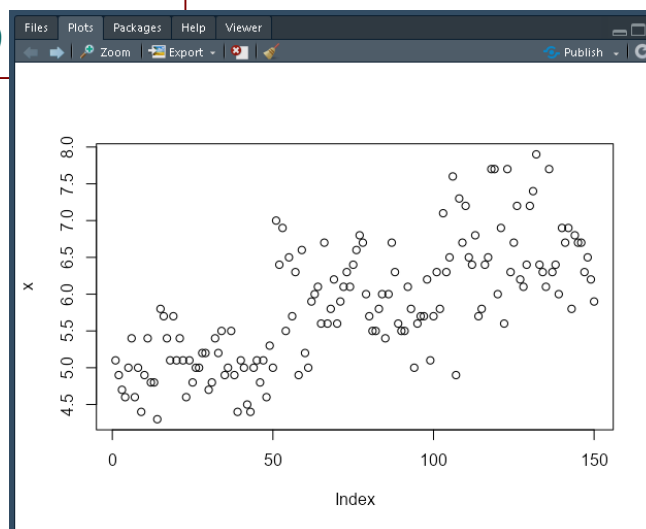
axTicks: Compute Axis Tickmark Locations
close.screen: Creating and Controlling Multiple Screens on a Single Device
erase.screen: Creating and Controlling Multiple Screens on a Single Device
frame: Create/Start a New Plot Frame
grconvertX: Convert between Graphics Coordinate Systems
grconvertY: Convert between Graphics Coordinate Systems
identify: Identify Points in a Scatter Plot
layout: Specifying Complex Plot Arrangements
lcm: Specifying Complex Plot Arrangements
locator: Graphical Input
screen: Creating and Controlling Multiple Screens on a Single Device
split.screen: Creating and Controlling Multiple Screens on a Single Device
strheight: Plotting Dimensions of Character Strings and Math Expressions
strwidth: Plotting Dimensions of Character Strings and Math Expressions

Parameters:

asp: Set up World Coordinates for Graphics Window
clip: Set Clipping Region
par: Set or Query Graphical Parameters
pch: Add Points to a Plot
xinch: Graphical Units
xlim: Set up World Coordinates for Graphics Window
xyinch: Graphical Units
yinch: Graphical Units
ylim: Set up World Coordinates for Graphics Window

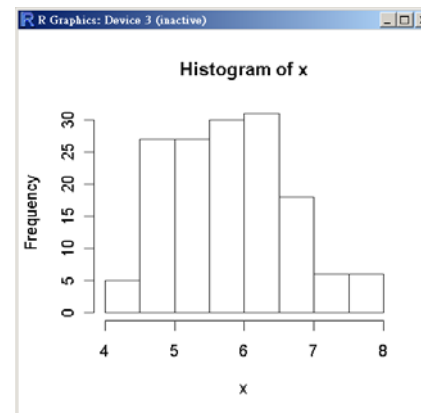
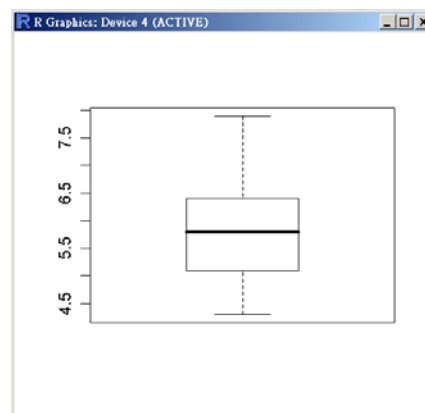
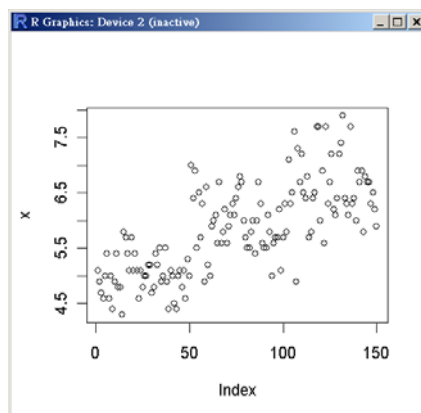
Plots for Single Samples

```
> x <- iris[,1]
> plot(x)
> hist(x)
> boxplot(x)
```



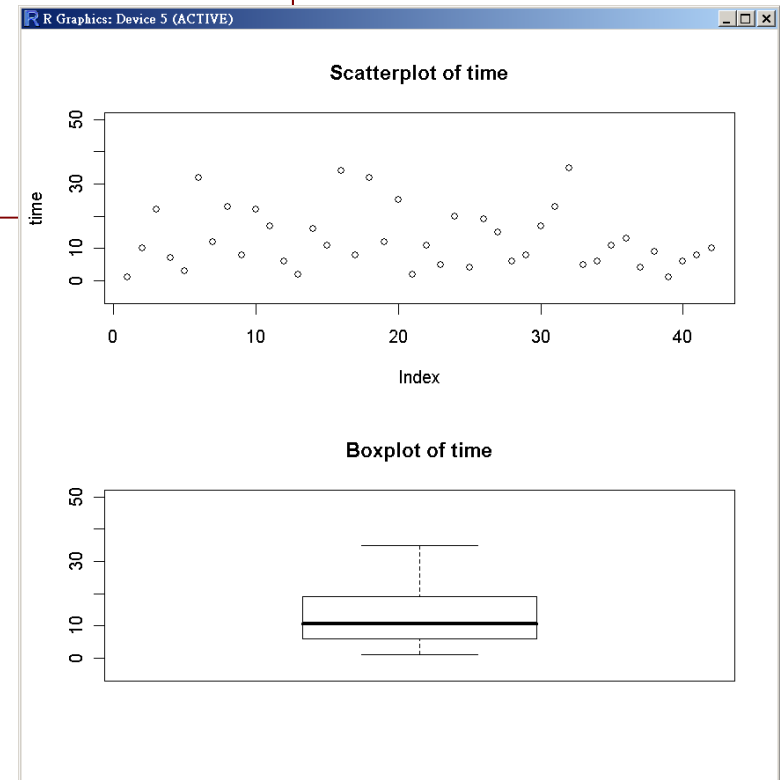
```
> title("Boxplot of time")
> hist(time, xlim=c(-5, 50))
> title("Histogram of time")
>
> hist(time, xlim=c(-5, 50))
> hist(time, xlim=c(-5, 50))
Error in plot.new() : figure margins too large
>
```

將繪圖視窗區域拉大



```
> head(gehan)
  pair time cens  treat
1    1    1    1 control
2    1   10    1   6-MP
3    2   22    1 control
4    2    7    1   6-MP
5    3    3    1 control
6    3   32    0   6-MP
```

```
> library(MASS)
> data(gehan)
> time <- gehan$time
>
> par(mfrow=c(2,1))
> plot(time, ylim=c(-5, 50), main="Scatterplot of time")
> boxplot(time, ylim=c(-5, 50), main="Boxplot of time")
>
> par(mfrow=c(2,1))
> s.title <- "Scatterplot of time"
> plot(time, ylim=c(-5, 50), main=s.title)
> b.title <- "Boxplot of time"
> boxplot(time, ylim=c(-5, 50), main=b.title)
```



- Standard Arguments: many high-level plot functions accept them.

```
> y <- iris[,1]
```

- `type="l"`: lines, `lwd`: line width

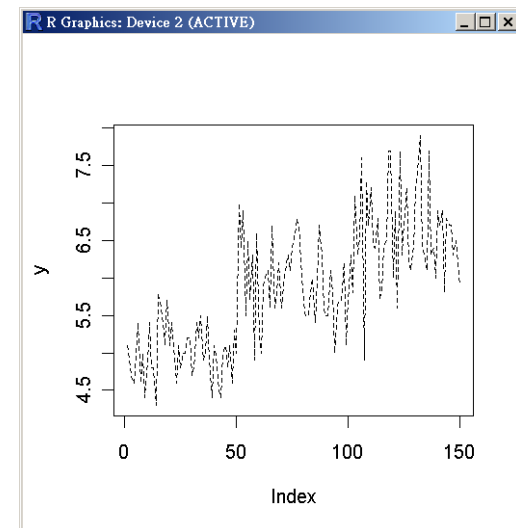
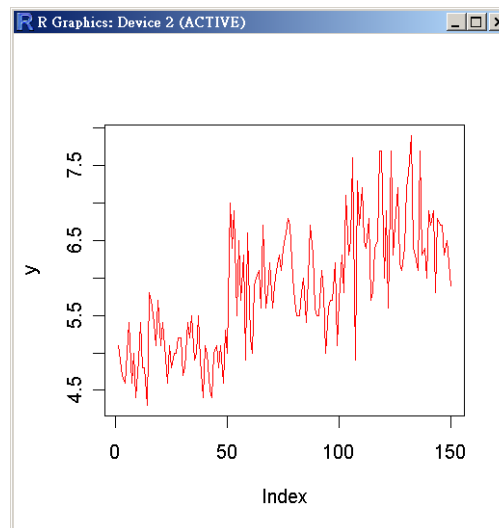
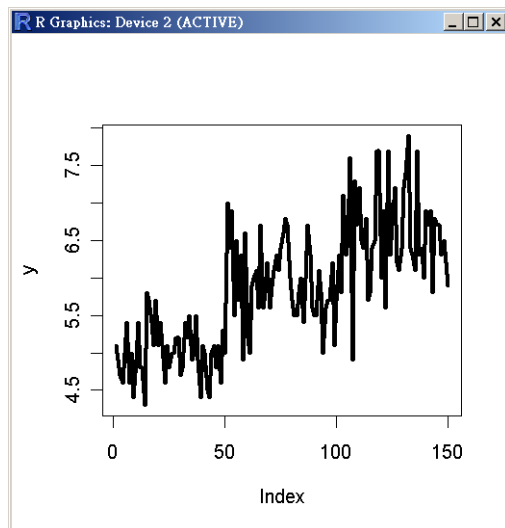
```
> plot(y, type="l", lwd=3)
```

- `col`: color 顏色

```
> plot(y, type="l", col="red")
```

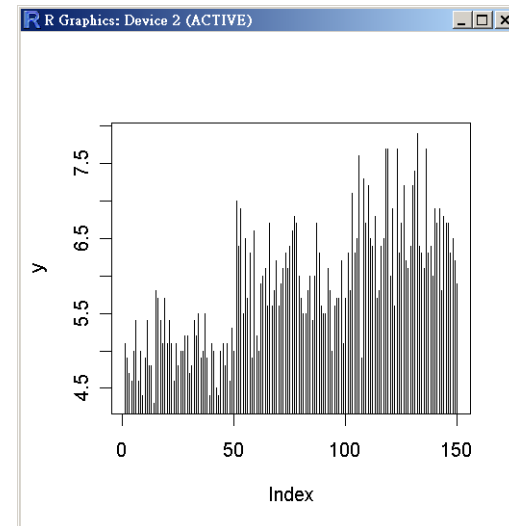
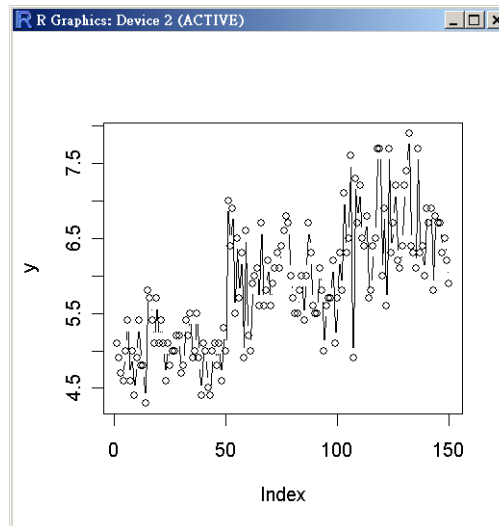
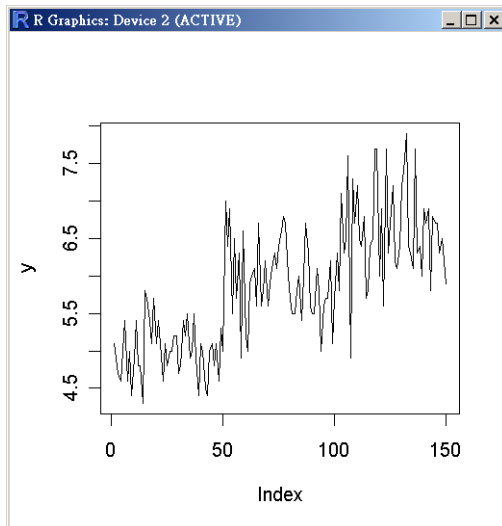
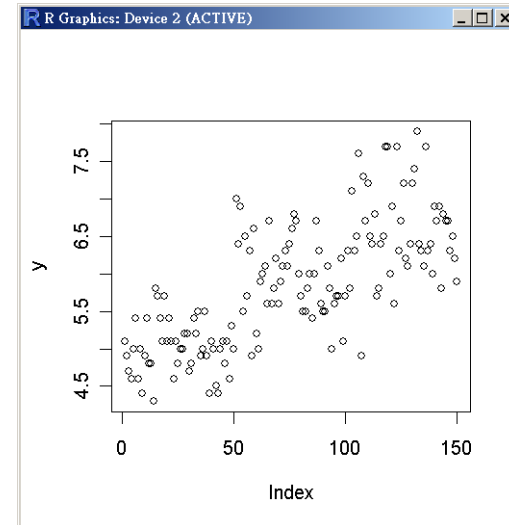
- `lty`: line type 線的型式

```
> plot(y, type="l", lty="dashed")
```

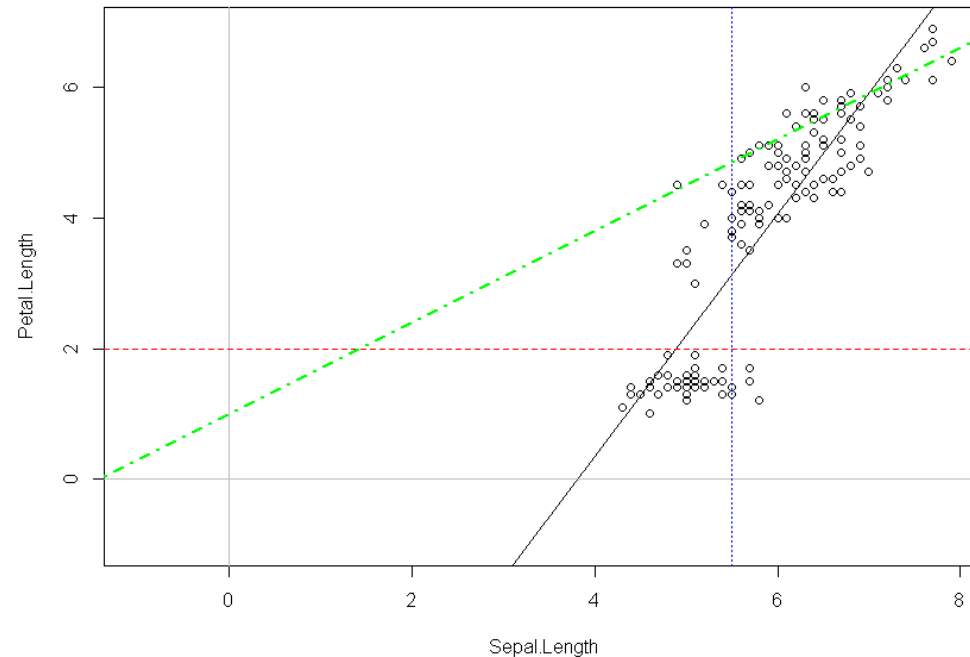


線的型式 (Line Type)

```
> y <- iris[,1]
> plot(y, type="p") # points
> plot(y, type="l") # lines
> plot(y, type="b") # both
> plot(y, type="h") #histogram-like
> plot(y, type="n") # none
```

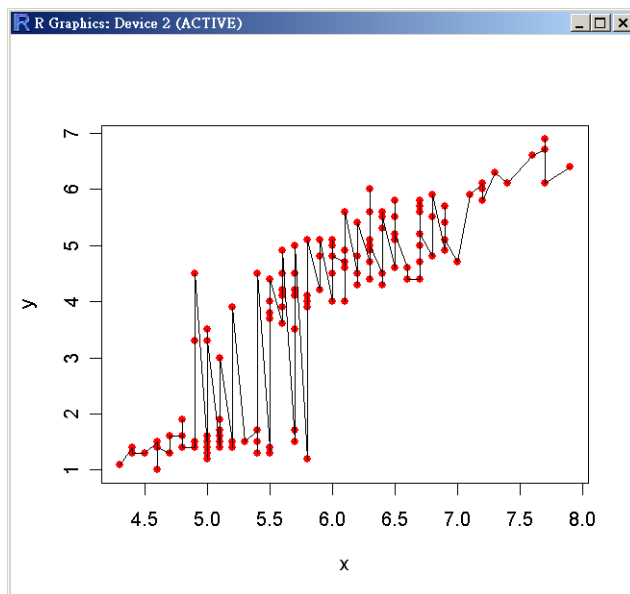
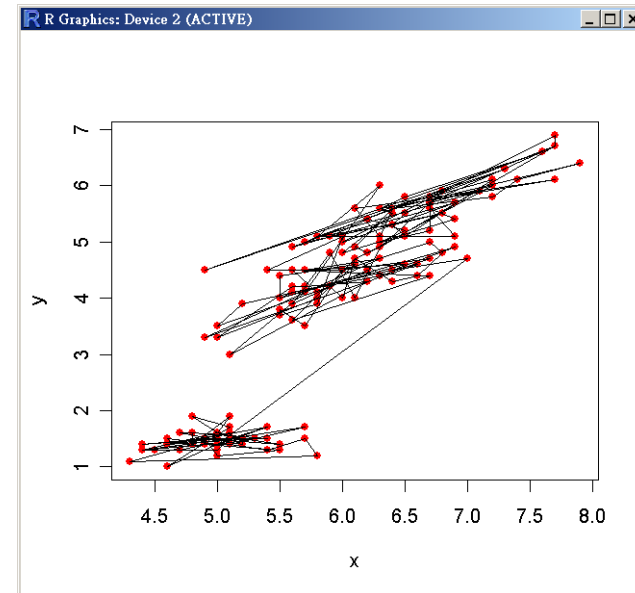


```
attach(iris)
plot(Sepal.Length, Petal.Length, xlim = c(-1, max(Sepal.Length)),
     ylim = c(-1, max(Petal.Length)))
abline(lm(Petal.Length ~ Sepal.Length), col = "black")
abline(h = 0, col = "grey")
abline(v = 0, col = "grey")
abline(h = 2, col = "red", lty = 2)
abline(v = 5.5, col = "blue", lty = 3)
abline(a = 1, b = 0.7, col = "green", lty = 4, lwd = 2)
detach(iris)
```



- Join the points on a scatterplot by lines: the trick is to ensure that **the points on the x axis are ordered**.
- if they are not ordered, the result is a mess.

```
> x <- iris[, 1]
> y <- iris[, 3]
> plot(x, y, pch=16, col="red")
> lines(x,y)
```

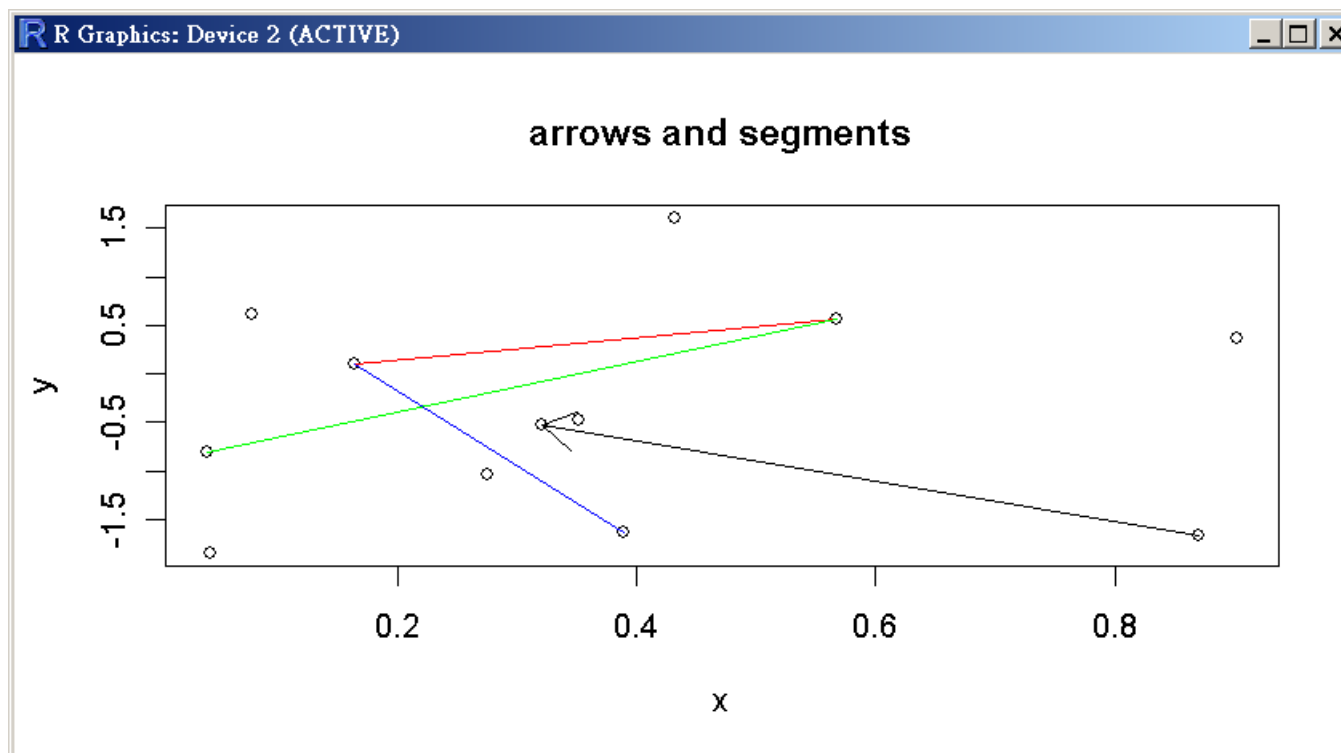


```
> sequence <- order(x)
> plot(x, y, pch=16, col="red")
> lines(x[sequence], y[sequence])
```

```

> x <- runif(12)
> y <- rnorm(12)
> plot(x, y, main="arrows and segments")
> arrows(x[1], y[1], x[2], y[2], col= "black", length=0.2)
> segments(x[3], y[3], x[4], y[4], col= "red")
> segments(x[3:4], y[3:4], x[5:6], y[5:6], col= c("blue", "green"))

```



座標系統 (Coordinate System)

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- The drawing of data **symbols**, **lines**, **text**, and so on in the plot region is relative to this user coordinate system.
- The scale on the axes: **xlim**, **ylim** or via **par()**.

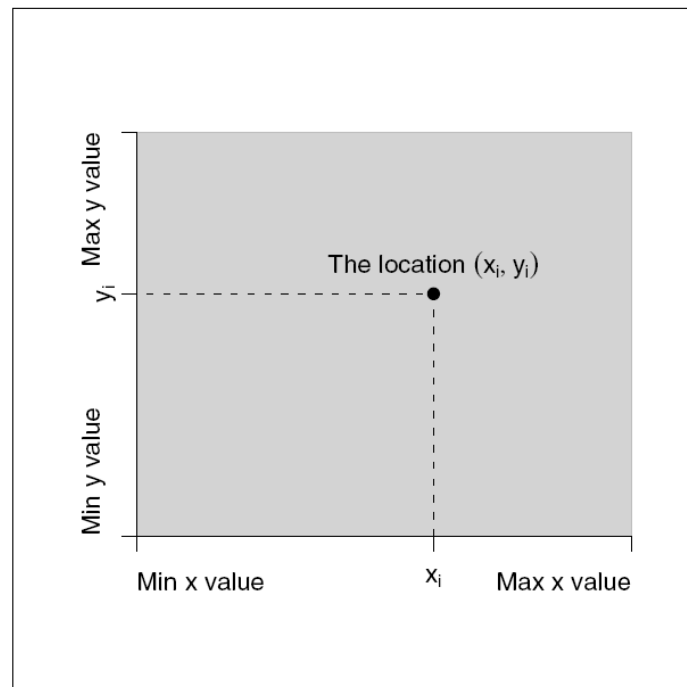
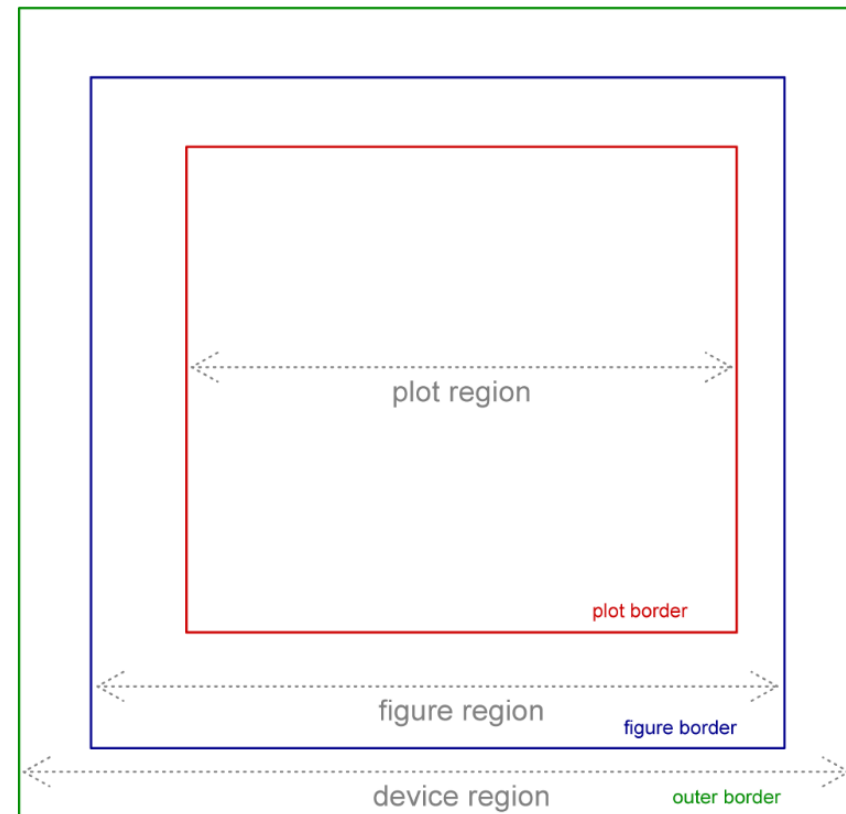


Figure 3.3

The user coordinate system in the plot region. Locations within this coordinate system are relative to the scales on the plot axes.



Murrell, P., 2005, R graphics, Chapman & Hall/CRC; 1 edition

Table 3.1

High-level traditional graphics state settings. This set of graphics state settings can be queried and set via the `par()` function *and* can be used as arguments to other graphics functions (e.g., `plot()` or `lines()`). Each setting is described in more detail in the relevant Section.

Setting	Description
adj	justification of text
ann	draw plot labels and titles?
bg	“background” color
bty	type of box drawn by <code>box()</code>
cex	size of text (multiplier)
cex.axis	size of axis tick labels
cex.lab	size of axis labels
cex.main	size of plot title
cex.sub	size of plot sub-title
col	color of lines and data symbols
col.axis	color of axis tick labels
col.lab	color of axis labels
col.main	color of plot title
col.sub	color of plot sub-title
fg	“foreground” color
font	font face (bold, italic) for text
font.axis	font face for axis tick labels
font.lab	font face for axis labels
font.main	font face for plot title
font.sub	font face for plot sub-title
gamma	gamma correction for colors
lab	number of ticks on axes
las	rotation of text in margins
lty	line type (solid, dashed)
lwd	line width
mgp	placement of axis ticks and tick labels
pch	data symbol type
srt	rotation of text in plot region
tck	length of axis ticks (relative to plot size)
tcl	length of axis ticks (relative to text size)
tmag	size of plot title (relative to other labels)
type	type of plot (points, lines, both)
xaxp	number of ticks on x-axis
xaxs	calculation of scale range on x-axis
xaxt	x-axis style (standard, none)
xpd	clipping region
yaxp	number of ticks on y-axis
yaxs	calculation of scale range on y-axis
yaxt	y-axis style (standard, none)

Murrell, P., 2005, R graphics, Chapman & Hall/CRC; 1 edition

> `par()` # list a complete list of the current graphics state.

- Setting will hold until a different setting is specified.

```
> par(c("col", "lty"))
```

```
> par(col="red", lty="dashed")
```

```
> y <- rnorm(20)
```

```
> plot(y, type="l") # dashed line
```

```
> plot(y, type="l", lty="solid") # solid line
```

```
> plot(y, type="l") # dashed line
```

```
> op <- par()
```

```
> par(col="red")
```

```
> plot(1,2)
```

```
> par(op)
```

- `par()` will affect all subsequent graphical output.

- High-level: `plot(..., col="red")`

- The setting will affect the output just for that plot.

- Low-level: `lines(..., col="red")`

- Control the appearance of a single piece of graphical output.

>?par

Setting	Description
ask	prompt user before new page?
family	font family for text
fig	location of figure region (normalized)
fin	size of figure region (inches)
lend	line end style
lheight	line spacing (multiplier)
ljoin	line join style
lmitre	line mitre limit
mai	size of figure margins (inches)
mar	size of figure margins (lines of text)
mex	line spacing in margins
mfcpl	number of figures on a page
mfg	which figure is used next
mfrow	number of figures on a page
new	has a new plot been started?
oma	size of outer margins (lines of text)
omd	location of inner region (normalized)
omi	size of outer margins (inches)
pin	size of plot region (inches)
plt	location of plot region (normalized)
ps	size of text (points)
pty	aspect ratio of plot region
usr	range of scales on axes
xlog	logarithmic scale on x-axis?
ylog	logarithmic scale on y-axis?

Table 3.2

Low-level traditional graphics state settings. This set of graphics state settings can be queried and set via the `par()` function. Each setting is described in more detail in the relevant Section.

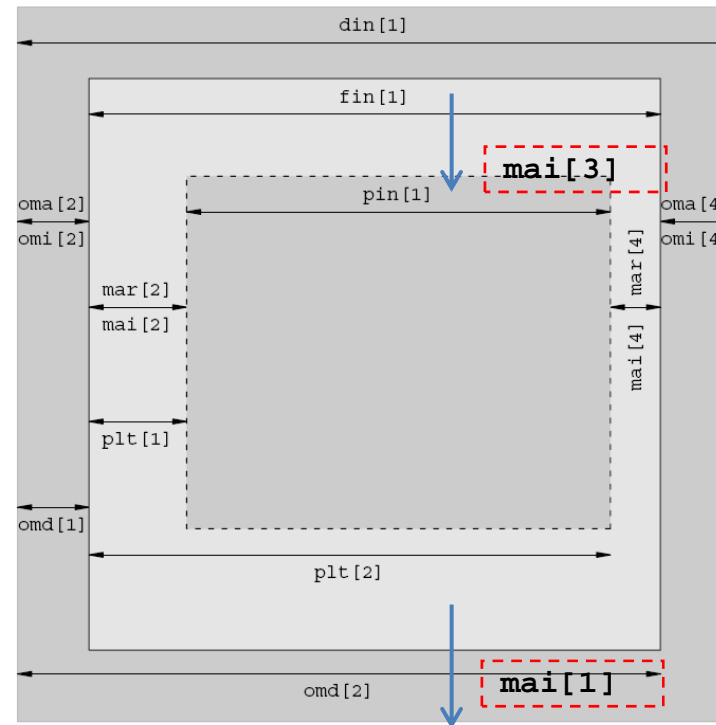
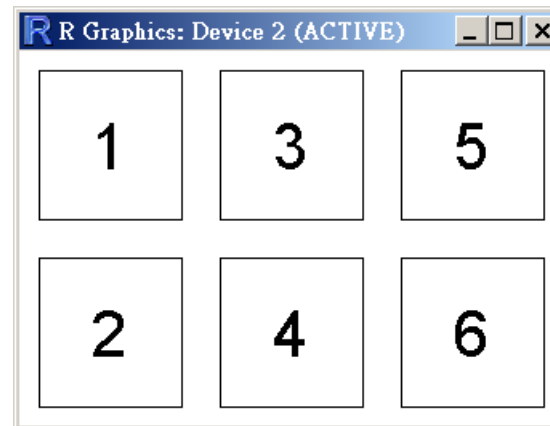
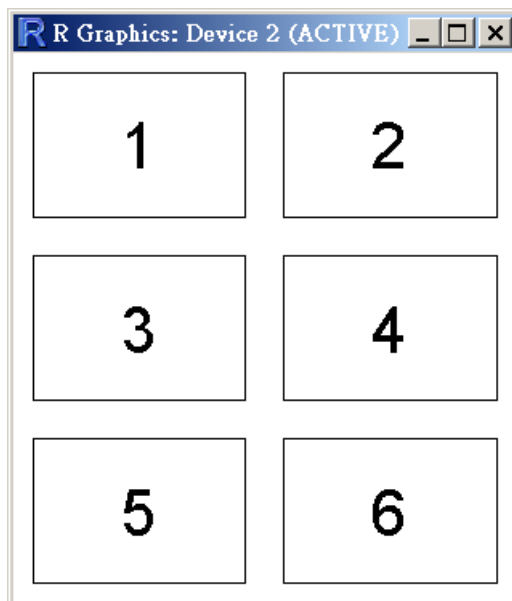


Figure 3.13

Graphics state settings controlling plot regions. These are some of the settings that control the widths and horizontal locations of the plot regions. For ease of comparison, this diagram has the same layout as Figure 3.1: the central grey rectangle represents the plot region, the lighter grey rectangle around that is the figure region, and the darker grey rectangle around that is the outer margins. A similar diagram could be produced for settings controlling heights and vertical locations.


```

> myplot <- function(n){
+   for(i in 1:n){
+     plot(1, type="n", xaxt="n", yaxt="n", xlab="", ylab="")
+     text(1, 1, labels=paste(i), cex=3)
+   }
+ }
>
> orig.par <- par(mai=c(0.1, 0.1, 0.1, 0.1), mfrow=c(3, 2))
> myplot(6)
> par(orig.par)
> par(mai=c(0.1, 0.1, 0.1, 0.1), mfcoll=c(2, 3))
> myplot(6)
    
```



- Specify the color of data symbols, lines, text that are drawn in the plot region.
- `col`, `fg`, `bg`
 - `col.axis` (axes), `col.lab` (label), `col.main` (titles), `col.sub` (sub-titles)
 - `fg`: color of axes and borders on plots.
 - `bg`: the color of the background for base graphics output. This color is used to fill the entire page.

```
> plot(1:10, rep(1, 10), pch=20, col=1:10, cex=5, xlab="", ylab="")
> text(1:10, rep(1.2, 10), labels=1:10)
```

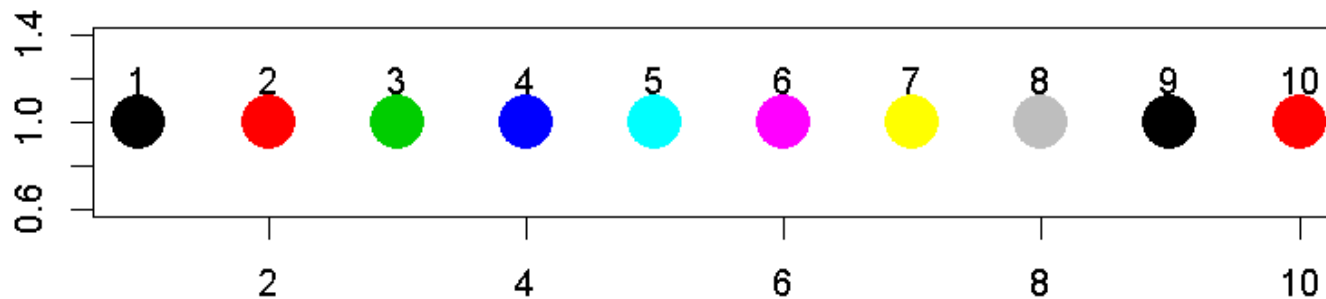


Table 3.4

Functions to generate color sets. R functions that can be used to generate coherent sets of colors

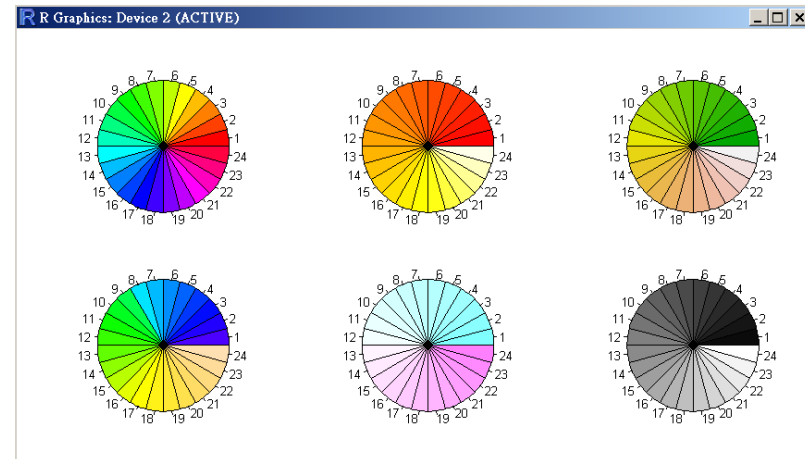
Name	Description
<code>rainbow()</code>	Colors vary from red through orange, yellow, green, blue, and indigo, to violet.
<code>heat.colors()</code>	Colors vary from white, through orange, to red.
<code>terrain.colors()</code>	Colors vary from white, through brown, to green.
<code>topo.colors()</code>	Colors vary from white, through brown then green, to blue.
<code>cm.colors()</code>	Colors vary from light blue, through white, to light magenta.
<code>grey()</code> or <code>gray()</code>	A set of shades of grey.

Murrell, P., 2005, R graphics, Chapman & Hall/CRC; 1 edition

Fill pattern

- `rect()`, `polygon()`, `hist()`, `barplot()`, `pie()`, `legend()`
- `rainbow(n)` # *n*: the number of colors

```
> par(mfrow=c(2,3))
> n <- 24
> pie(rep(1,n), col=rainbow(n))
> pie(rep(1,n), col=heat.colors(n))
> pie(rep(1,n), col=terrain.colors(n))
> pie(rep(1,n), col=topo.colors(n))
> pie(rep(1,n), col=cm.colors(n))
> pie(rep(1,n), col=grey(1:n/n))
```

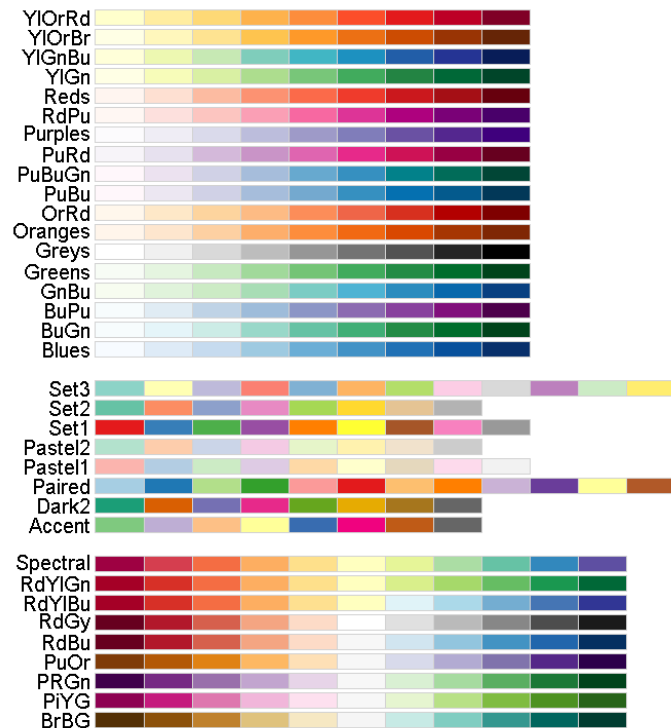


- **RColorBrewer**: Provides color schemes for maps (and other graphics) designed by Cynthia Brewer as described at <http://colorbrewer2.org>

```

brewer.pal(n, name)
display.brewer.pal(n, name)
display.brewer.all(n=NULL, type="all", select=NULL, exact.n=TRUE, colorblindFriendly=FALSE)
brewer.pal.info
# n: Number of different colors in the palette, mim=3, maximum depends
# name: A palette name from the lists below
# type: One of the string "div", "qual", "seq", or "all"

```



```

> library(RColorBrewer)
> brewer.pal.info

```

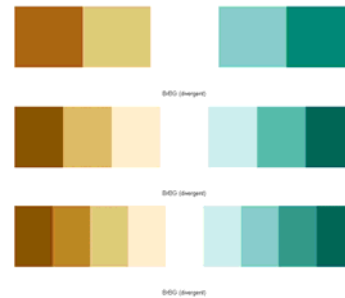
	maxcolors	category	colorblind
BrBG	11	div	TRUE
PiYG	11	div	TRUE
...			
Spectral	11	div	FALSE
Accent	8	qual	FALSE
Dark2	8	qual	TRUE
...			
YlOrBr	9	seq	TRUE
YlOrRd	9	seq	TRUE

```

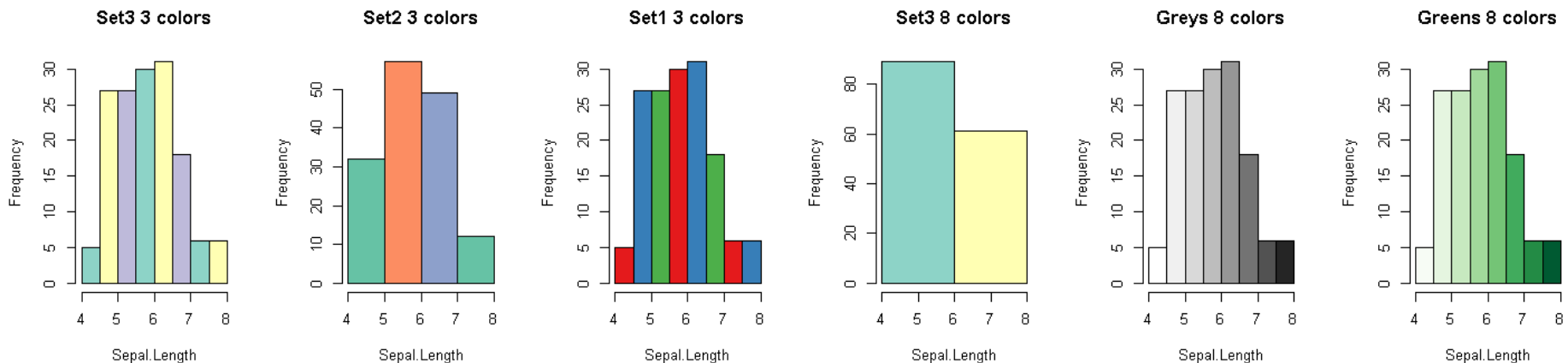
> display.brewer.all()

```

```
> display.brewer.pal(5, "BrBG")
> display.brewer.pal(7, "BrBG")
> display.brewer.pal(9, "BrBG")
```



```
> attach(iris)
> par(mfrow=c(1,6))
> hist(Sepal.Length, breaks=10, col=brewer.pal(3, "Set3"), main="Set3 3 colors")
> hist(Sepal.Length, breaks=3, col=brewer.pal(3, "Set2"), main="Set2 3 colors")
> hist(Sepal.Length, breaks=7, col=brewer.pal(3, "Set1"), main="Set1 3 colors")
> hist(Sepal.Length, breaks= 2, col=brewer.pal(8, "Set3"), main="Set3 8 colors")
> hist(Sepal.Length, col=brewer.pal(8, "Greys"), main="Greys 8 colors")
> hist(Sepal.Length, col=brewer.pal(8, "Greens"), main="Greens 8 colors")
```



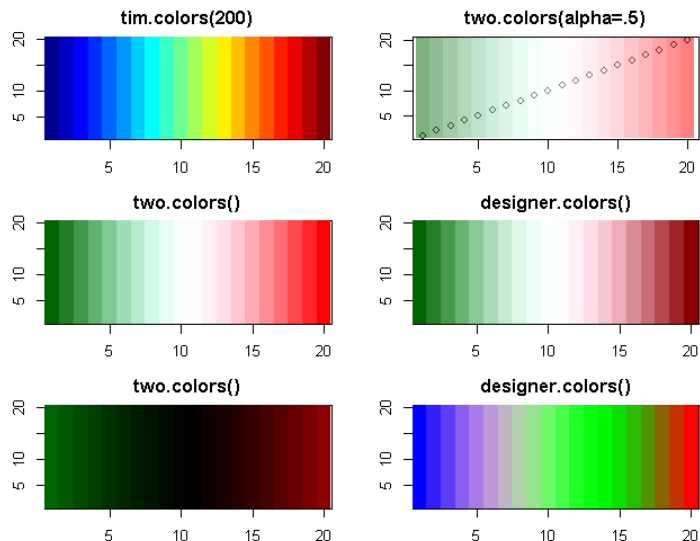
tim.colors {fields}: Some useful color tables for images and tools to handle them

```
> library(fields)
> par(mfcol=c(3,2))
> x <- 1:20
> y <- 1:20
> z <- outer(x, rep(1,20), "+")
> obj <- list(x=x, y=y, z=z)
> image(obj, col=tim.colors(200), main="tim.colors(200)")
> image(obj, col=two.colors(), main="two.colors()")
> image(obj, col=two.colors(start="darkgreen", end="darkred", middle="black"),
+       main="two.colors()")
> plot(x, y, main="two.colors(alpha=.5)")
> image(obj, col=two.colors(alpha=.5), add=TRUE)
> image(obj, col=designer.colors(), main="designer.colors()")
> coltab <- designer.colors(col=c("blue", "grey", "green", "red"))
> image(obj, col= coltab, main="designer.colors()")
```

```
> obj
$xc
[1] 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20

$y
[1] 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20

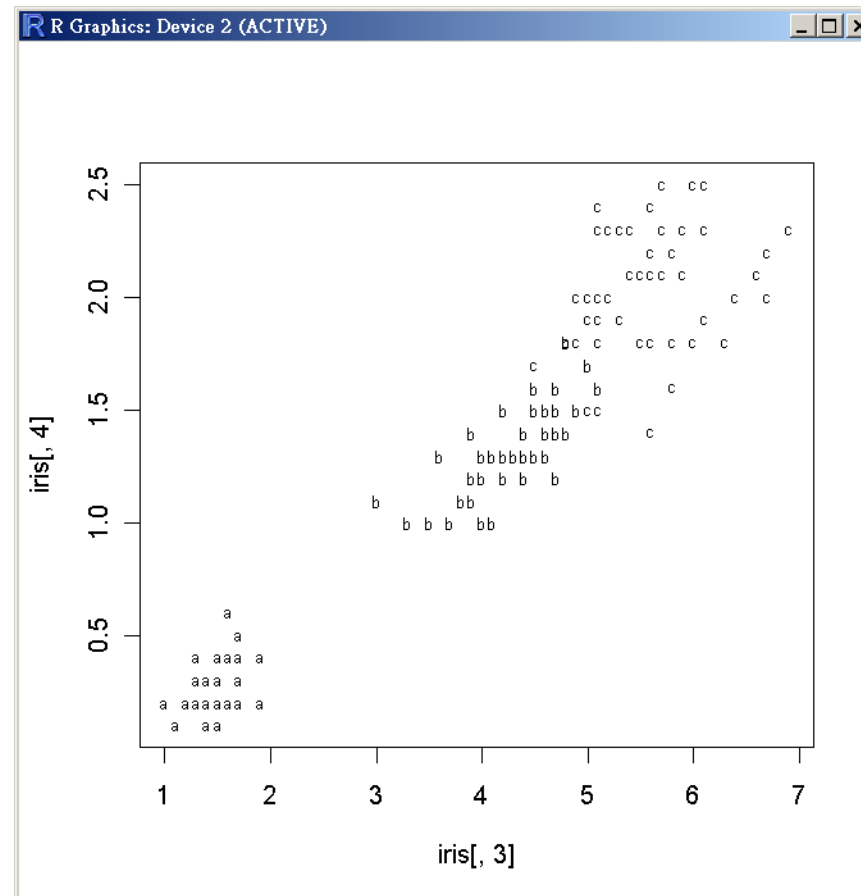
$z
      [,1] [,2] [,3] [,4] [,5] [,6] [,7] [,8] [,9] [,10] [,11] [,12]
[1,]  2   2   2   2   2   2   2   2   2   2   2   2
[2,]  3   3   3   3   3   3   3   3   3   3   3   3
[3,]  4   4   4   4   4   4   4   4   4   4   4   4
[4,]  5   5   5   5   5   5   5   5   5   5   5   5
[5,]  6   6   6   6   6   6   6   6   6   6   6   6
[6,]  7   7   7   7   7   7   7   7   7   7   7   7
[7,]  8   8   8   8   8   8   8   8   8   8   8   8
```



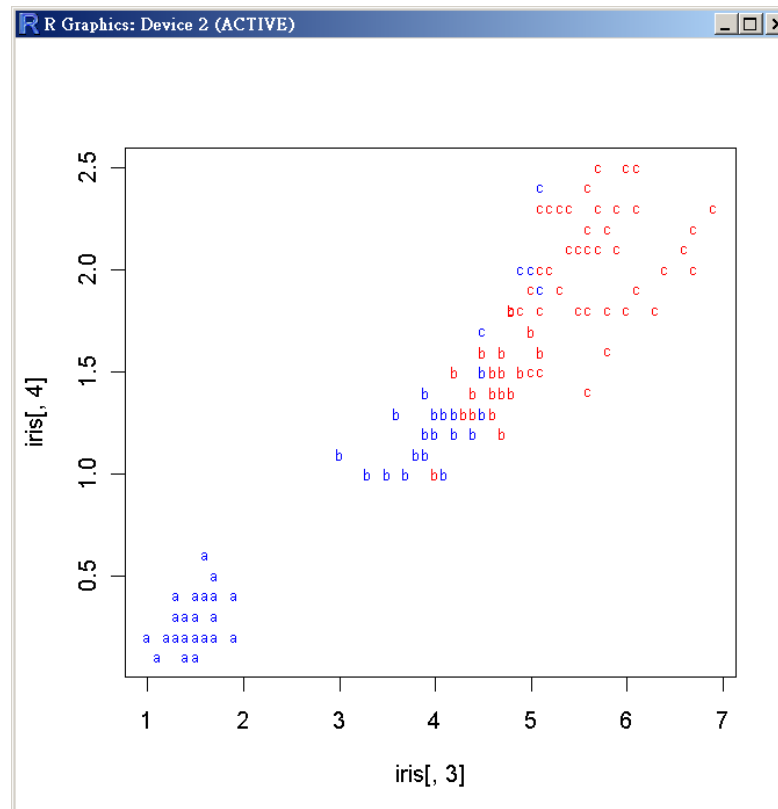
```
# fields: Tools for Spatial Data
tim.colors(n = 64, alpha=1.0)
larry.colors()
two.colors(n=256, start="darkgreen", end="red", middle="white",
alpha=1.0)
designer.colors(n=256, col= c("darkgreen", "white", "darkred"),
x=seq(0, 1, length(col)) ,alpha=1.0)

color.scale( z, col=tim.colors(256), zlim =NULL,
transparent.color="white", eps= 1e-8)
fieldsPlotColors( col,...)
```

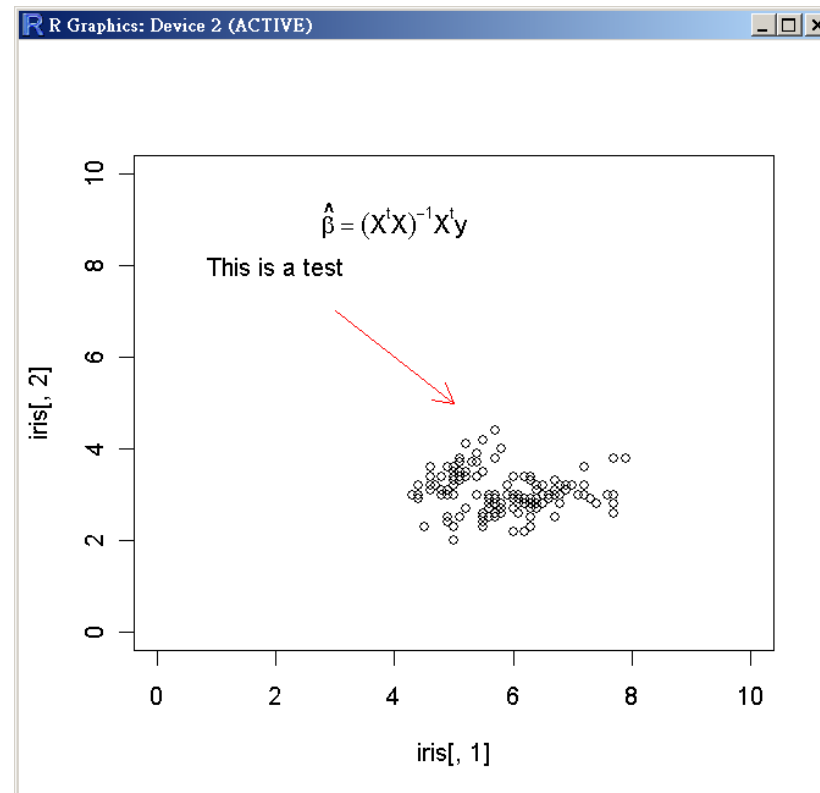
```
> plot(iris[,3], iris[,4], type="n")
> my.label <- c(rep("a", 50), rep("b", 50), rep("c", 50))
> text(iris[,3], iris[,4], labels=my.label, cex=0.7)
```



```
> plot(iris[,3], iris[,4], type="n")
> my.label <- c(rep("a", 50), rep("b", 50), rep("c", 50))
> text(iris[,3], iris[,4], my.label, cex=0.7,
      col=ifelse(iris[,1] > median(iris[,1]), "red", "blue"))
```




```
> plot(iris[,1], iris[,2], xlim=c(0, 10), ylim=c(0, 10))  
> text(2,8, "This is a test")  
> arrows(x0=3, y0=7, x1=5, y1=5, length = 0.15, col="red")  
> text(4, 9, expression(hat(beta) == (X^t * X)^{-1} * X^t * y))
```



A fixed set of 26 data symbols:

- **pch=numbers**
- **pch="a"**, a single character
- **pch="."**, small dot
- **cex**: size of data symbol
- **type**: how data is represented in a plot
 - **"p"**: data symbols are drawn at each (x,y) location
 - **"l"**: (x,y) locations are connected by lines
 - **"b"**: both data symbols and lines are drawn.
 - **"o"**: over-plot onlines
 - **"h"**: vertical lines are drawn from x-axis to the (x,y)
 - **"s"**, "S": city-block fashion, step
 - **"n"**: nothing is drawn

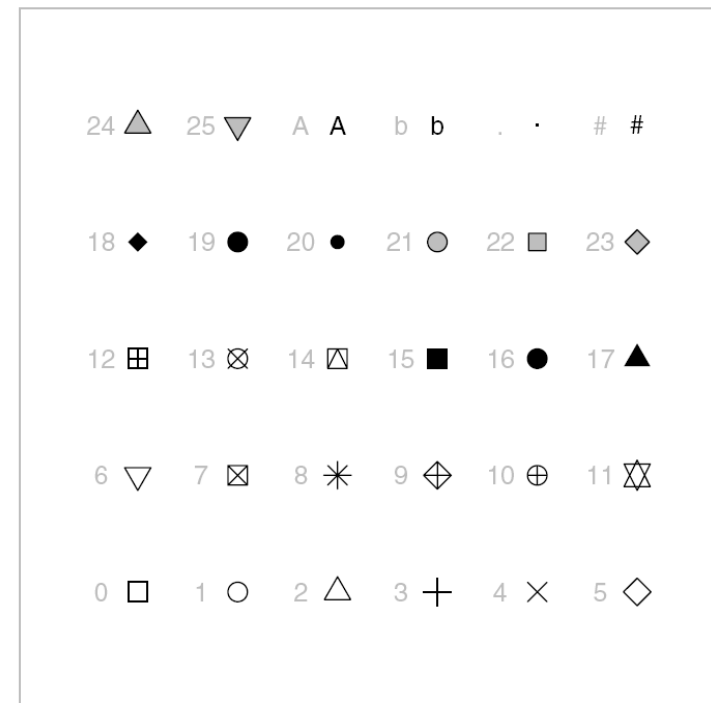


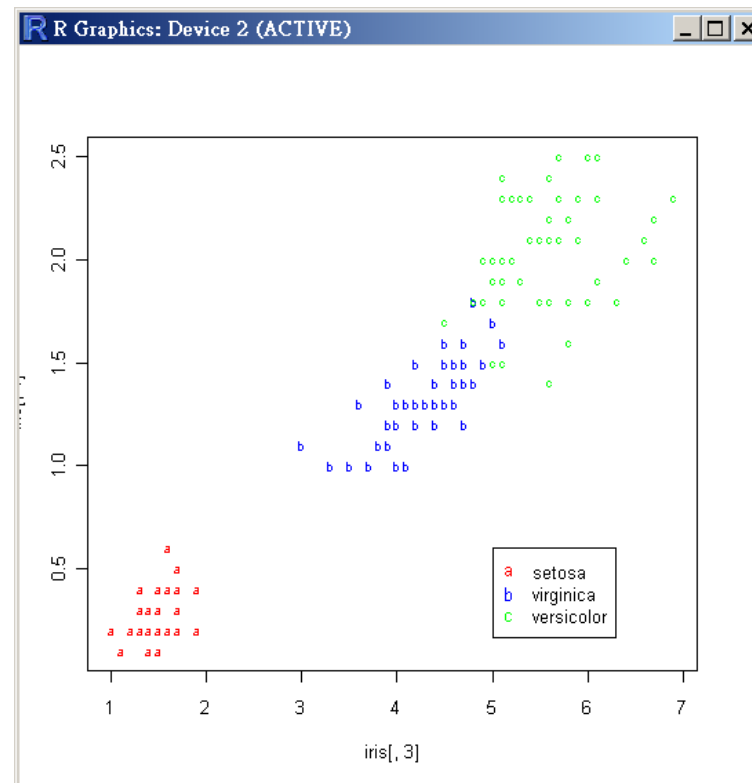
Figure 3.10

Data symbols available in R. A particular data symbol is selected by specifying an integer between 0 and 25 or a single character for the pch graphical setting. In the diagram, the relevant integer or character pch value is shown in grey to the left of the relevant symbol.

Murrell, P., 2005, R graphics, Chapman & Hall/CRC; 1 edition

Legend

```
> plot(iris[,3], iris[,4], type="n")
> my.label <- c(rep("a", 50), rep("b", 50), rep("c", 50))
> my.color <- c(rep("red", 50), rep("blue", 50), rep("green", 50))
> text(iris[,3], iris[,4], my.label, cex=0.7, col=my.color)
> legend(5, 0.6, legend=c("setosa","versicolor", "virginica"), pch = "abc",
        col=c("red","blue","green"))
```



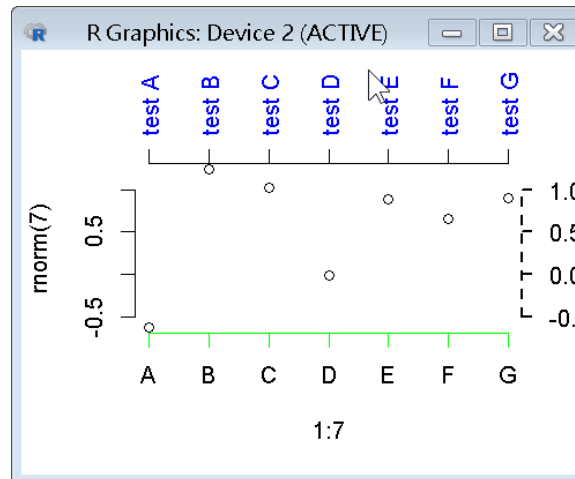
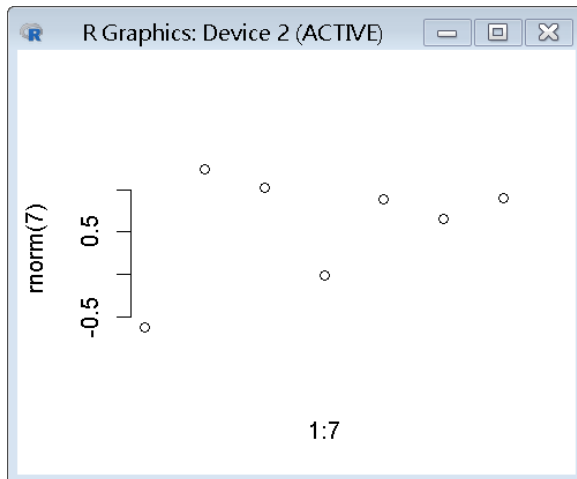
- 語法

```
axis(side, at = NULL, labels = TRUE, tick = TRUE,  
line = NA, pos = NA, outer = FALSE, font = NA,  
lty = "solid", lwd = 1, lwd.ticks = lwd, col = NULL,  
col.ticks = NULL, hadj = NA, padj = NA, ...)
```

- Arguments

- side: 為一整數 · 指定座標軸畫在圖形的哪一邊 · 1=below, 2=left, 3=above and 4=right.
- at: 指定圖上的tick-marks所在位置
- labels: 指定tick-marks的標號
- tick: (logical) tickmarks · axis line 需要畫出長來嗎?
- pos: (coordinate) axis line畫的位置
- hadj: adjustment for all labels parallel ('horizontal') to the reading direction.
- padj: adjustment for each tick label perpendicular to the reading direction
- las: labels are parallel (=0) or perpendicular(=2) to axis

```
plot(1:7, rnorm(7), xaxt = "n", frame = FALSE)
axis(1, 1:7, LETTERS[1:7], col = "green")
axis(3, 1:7, paste("test", LETTERS[1:7]), col.axis = "blue", las=2)
axis(4, lty=2, lwd = 2, las=2)
```

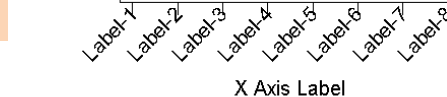
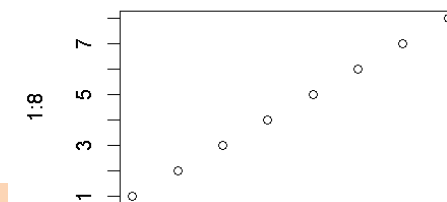
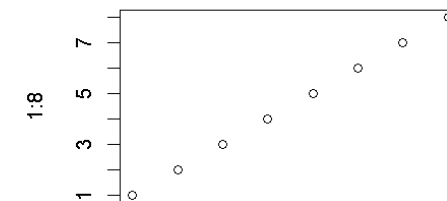
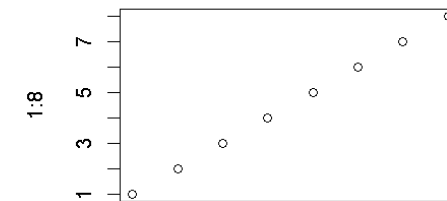


```
plot(1:8, xaxt = "n", xlab = "")
axis(1, labels = FALSE)
my.labels <- paste("Label", 1:8, sep = "-")
text(1:8, par("usr")[3] - 0.25, srt = 45, adj = 1,
     labels = my.labels, xpd = TRUE)
mtext(1, text = "X Axis Label", line = 3)
```

srt: string rotation

```
> par("usr")
[1] 0.72 8.28 0.72 8.28
```

xpd: all plotting is clipped to the figure region



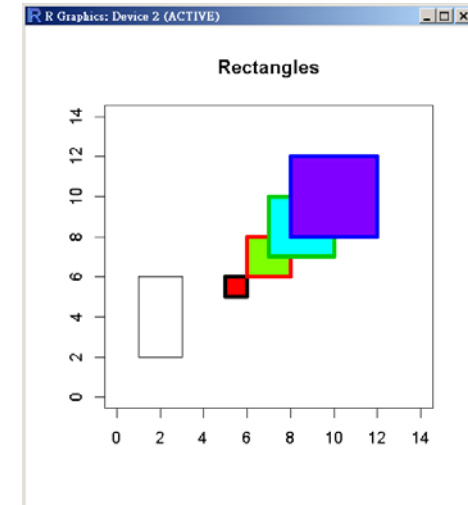
X Axis Label



矩形 · Draw Symbols

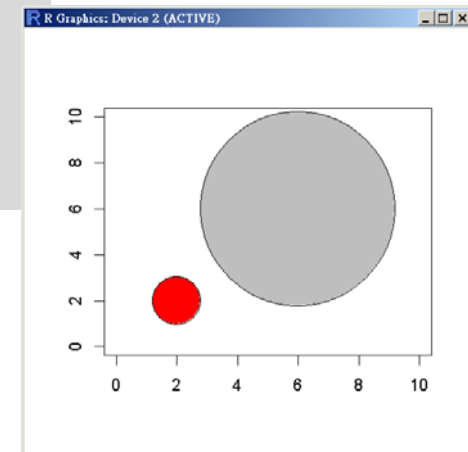
(Circles, Squares, Stars, Thermometers, Boxplots)

```
> plot(0, xlim=c(0,14), ylim=c(0, 14), type = "n",  
xlab = "", ylab = "", main = "Rectangles")  
> rect(1, 2, 3, 6)  
> n <- 0:3  
> rect(5+n, 5+n, 6+2*n, 6+2*n, col = rainbow(4),  
border = n+1, lwd=4)
```



```
symbols(x, y = NULL, circles, squares, rectangles, stars,  
thermometers, boxplots, inches = TRUE, add = FALSE,  
fg = par("col"), bg = NA,  
xlab = NULL, ylab = NULL, main = NULL,  
xlim = NULL, ylim = NULL, ...)
```

```
> symbols(x = c(2, 6), y = c(2, 6), circles = c(1,  
4), xlim=c(0, 10), ylim=c(0, 10), bg=c("red",  
"gray"), xlab="", ylab="")
```



See also "draw.ellipse {plotrix}"

```
> install.packages(c("tiff", "jpeg", "png", "fftwtools"),  
repos="http://cran.csie.ntu.edu.tw")
```

```
> library(EBImage) # (Repositories: BioC Software)
```

```
> Transformers <- readImage("Transformers07.jpg")
```

```
> (dims <- dim(Transformers))
```

```
[1] 300 421 3
```

```
> Transformers
```

```
Image
```

```
colorMode      : Color
```

```
storage.mode   : double
```

```
dim            : 300 421 3
```

```
frames.total   : 3
```

```
frames.render : 1
```

```
imageData(object)[1:5,1:6,1]
```

```
      [,1] [,2] [,3] [,4] [,5] [,6]
```

```
[1,] 0 0 0 0 0 0
```

```
[2,] 0 0 0 0 0 0
```

```
[3,] 0 0 0 0 0 0
```

```
[4,] 0 0 0 0 0 0
```

```
[5,] 0 0 0 0 0 0
```

```
> plot(c(0, dims[1]), c(0, dims[2]), type='n',
```

```
+ xlab="", ylab="")
```

```
> rasterImage(Transformers, 0, 0, dims[1], dims[2])
```

```
if (!requireNamespace("BiocManager", quietly = TRUE))  
  install.packages("BiocManager")
```

```
BiocManager::install("EBImage")
```

```
> #install.packages("jpeg")
```

```
> library(jpeg)
```

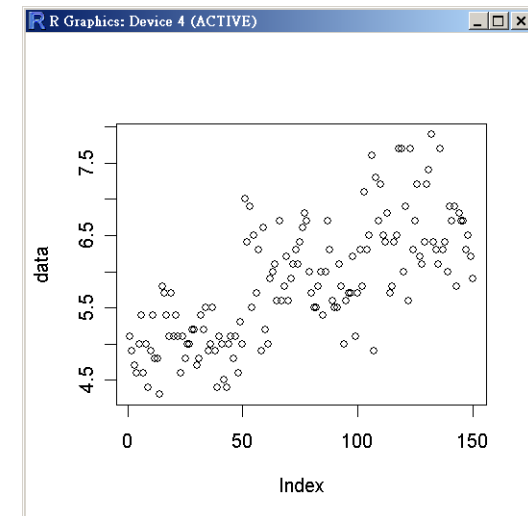
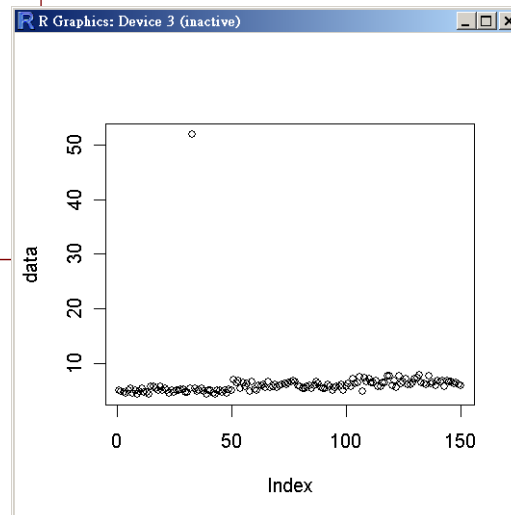
```
> Transformers <- readJPEG("Transformers07.jpg")
```



[https://en.wikipedia.org/wiki/Transformers_\(film\)](https://en.wikipedia.org/wiki/Transformers_(film))

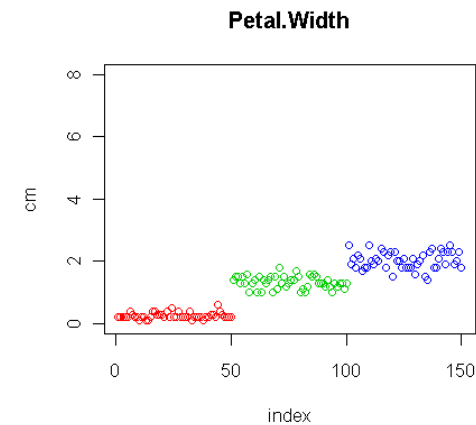
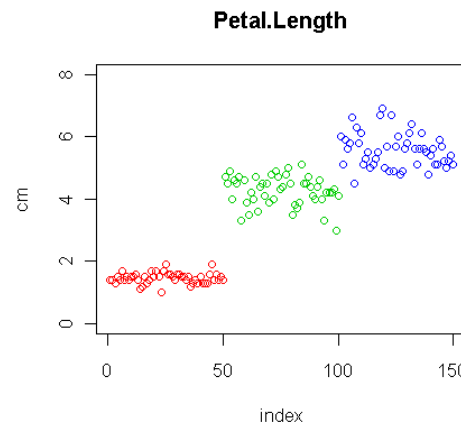
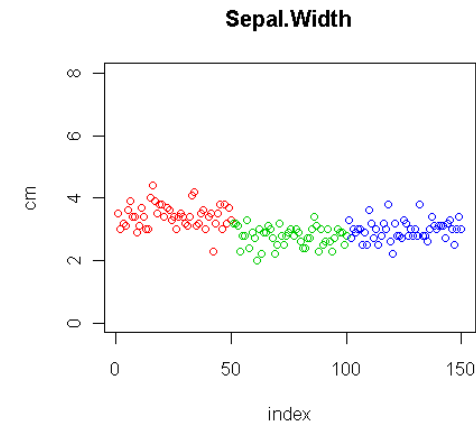
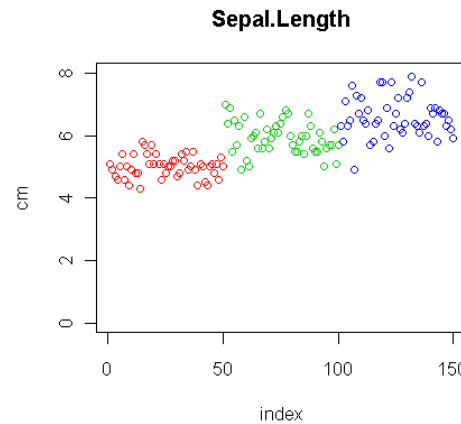
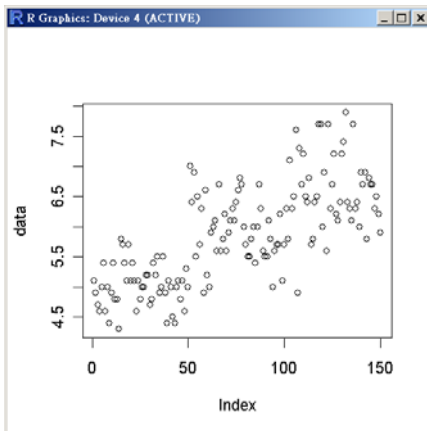
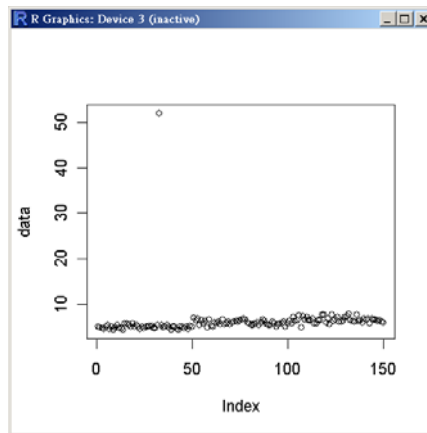
- Index plot takes a single argument which is a **continuous variable** and plots the **values** on the y axis, with the x coordinate determined by the **position** of the number in the vector.
- Useful for error checking.

```
> data <- iris[,1]
> data[33] <- data[33]*10
> plot(data)
> ind <- which(data>15)
> data[ind]
> data[ind] <- 5.2
> windows()
> plot(data)
```



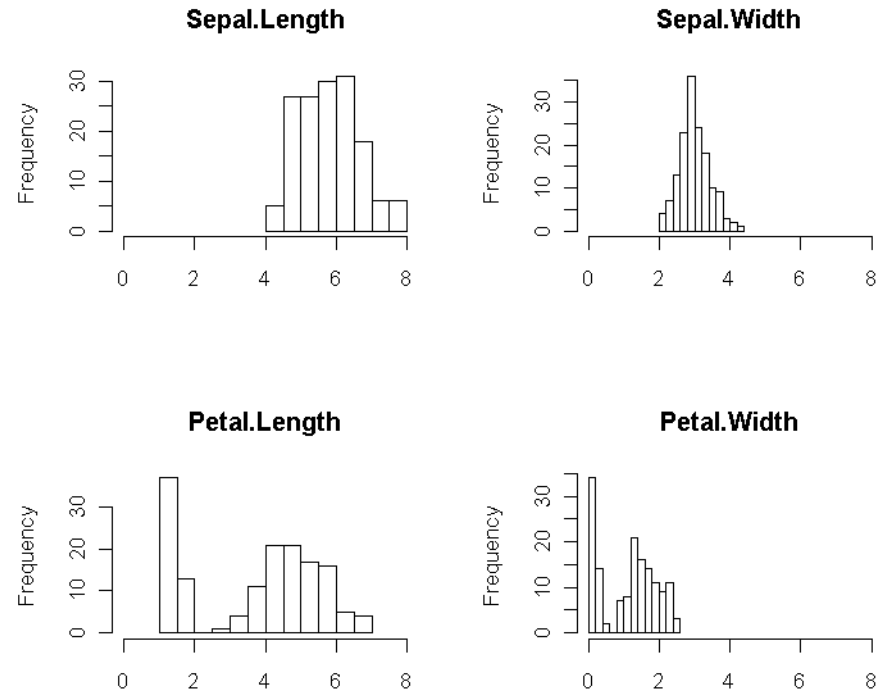
Index Plot

- Index plot takes a single argument which is a **continuous variable** and plots the **values** on the y axis, with the x coordinate determined by the **position** of the number in the vector.
- Useful for error checking.

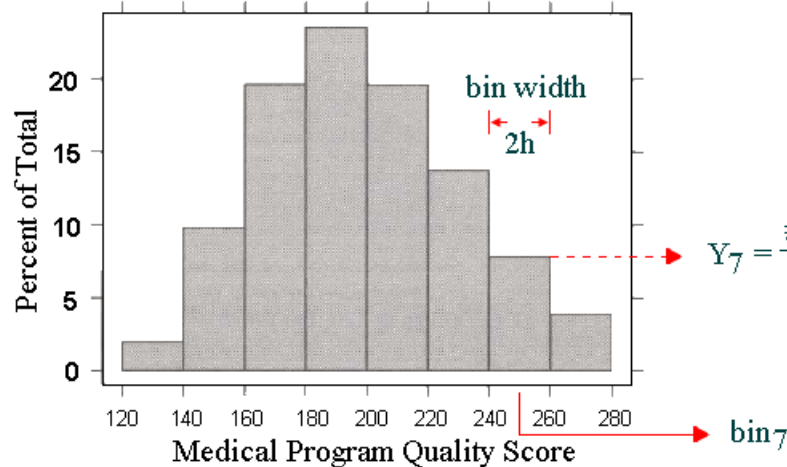


The histogram shows:

1. center of the data (location)
2. spread of the data (scale)
3. skewness of the data
4. presence of outliers
5. presence of multiple modes in the data.



O. Bin origin at 120, bin widths of 20.



```
par(mfrow=c(2, 2))
lapply(1:4, function(x) hist(iris[,x], xlim=c(0, 8),
ylim=c(0, 30), xlab="x", main=names(iris)[x]))
```

$$Y_7 = \frac{\text{\# observations within bin}_7}{(2h) n}$$

Changes in bin origin and bin widths affect the shape of the histogram

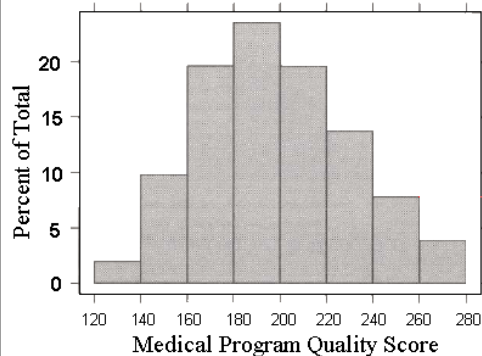
直方圖 (Histogram) (2/3)

- $1/2h$ adjusts the height of each bar so that the total area enclosed by the entire histogram is 1.
- The area covered by each bar can be interpreted as the probability of an observation falling within that bar.

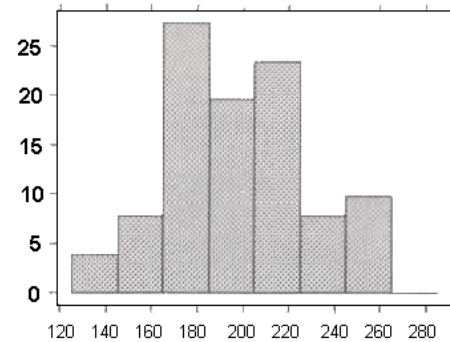
Disadvantage for displaying a variable's distribution:

- selection of **origin** of the bins.
- selection of **bin widths**.
- the very use of the bins is a distortion of information because any data **variability within** the bins cannot be displayed in the histogram.

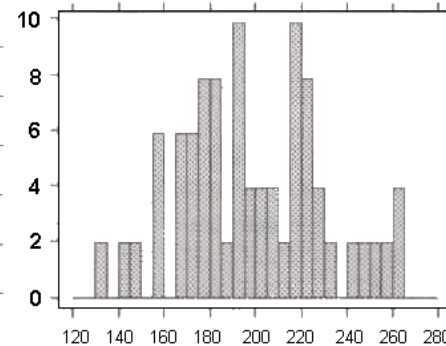
O. Bin origin at 120, bin widths of 20.



A. Bin origin at 125, bin widths of 20.



B. Bin origin at 120, bin widths of 5.



C. Bin origin at 120, bin widths of 10.

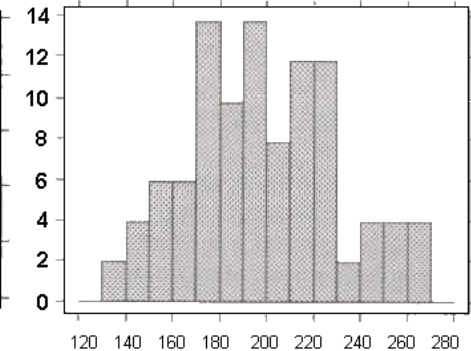
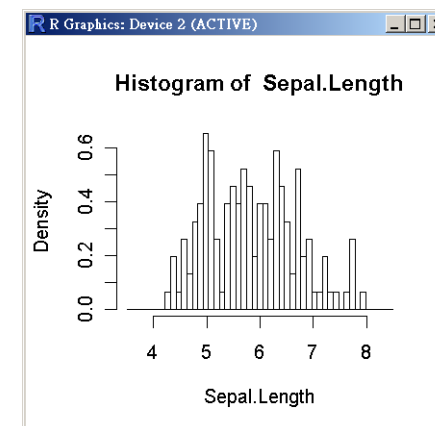
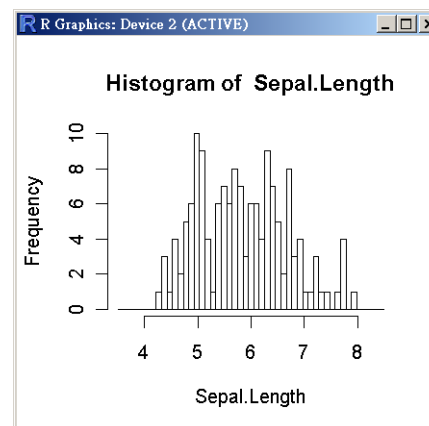
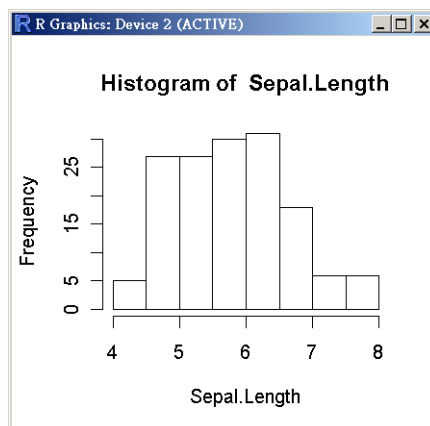


Figure Sources: Jacoby (1997).

直方圖 (Histogram) (3/3)

- Histogram are excellent for showing the mode, the spread and the symmetry (skew) of a set of data.
- Arguments
 - `breaks`
 - `pro=F` #default, 次數, `pro=T` #機率值

```
> lab <- names(iris)[1]
> title <- paste("Histogram of ", lab)
> hist(iris[,1], main=title, xlab=lab)
> range(iris[,1])
> hist(iris[,1], breaks=seq(3.5, 8.5, length=50),main=title, xlab=lab)
> hist(iris[,1], breaks=seq(3.5, 8.5, length=50),main=title, xlab=lab, pro=T)
```

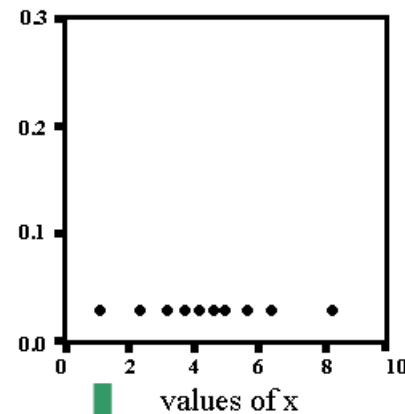


Density Plots (Smoothed Histograms) (1/3)

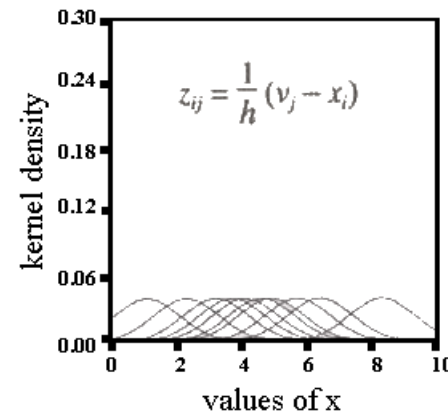
- Smoothed histograms overcome some of the disadvantages caused by the arbitrary, discrete bins used in traditional histogram.
- The relative height of the smooth curve corresponds to the local density.
- The overall height is adjusted so that the total area under the curve is approximately equal to 1.
- The area under the curve between any two points along the horizontal scale can be interpreted as the probability that an observation falls within that interval of data values.

Constructing a Smoothed Histogram (Jacoby, 1997)

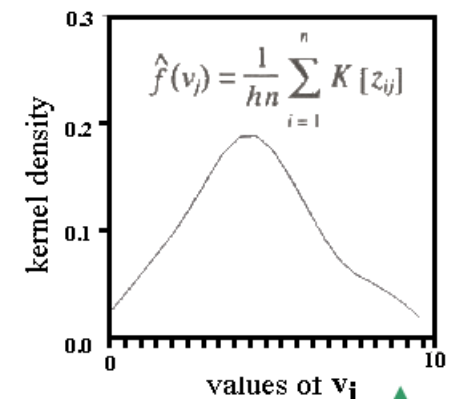
A. Unidimensional scatterplot of 10 data points



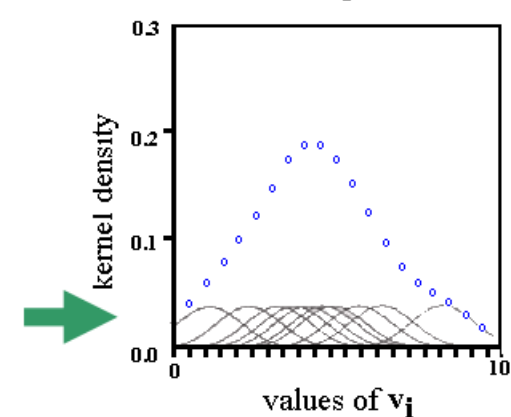
B. Data points shown as kernel densities



D. Final smoothed histogram



C. Summing kernel densities at the 20 v_i

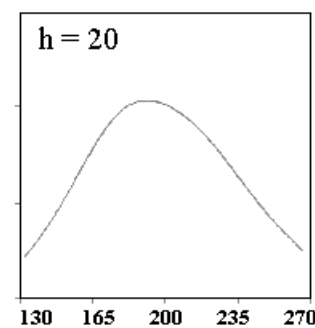
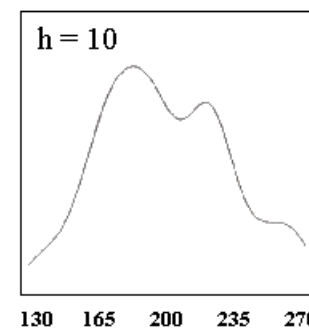
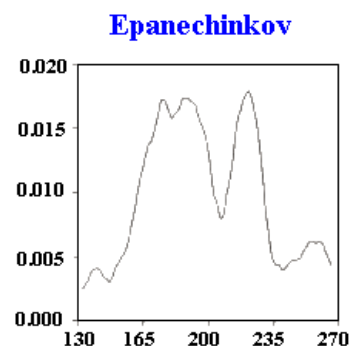
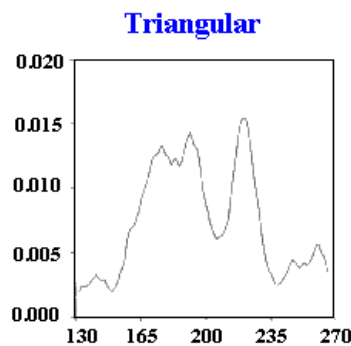
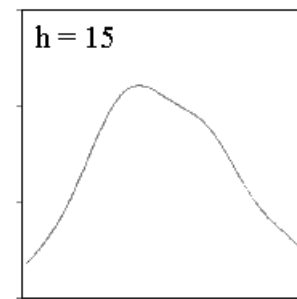
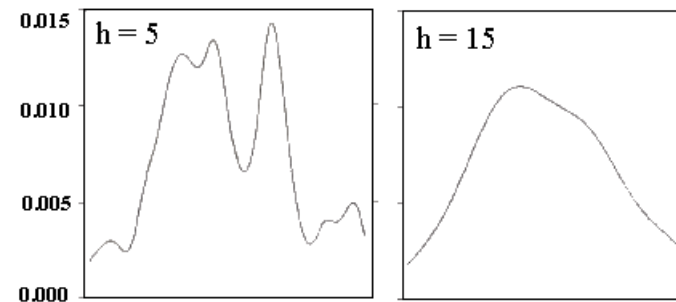
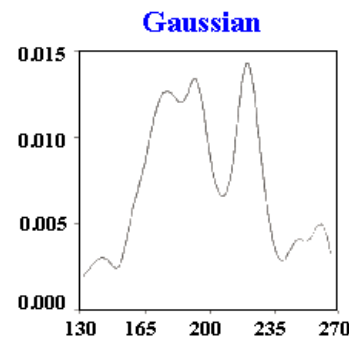
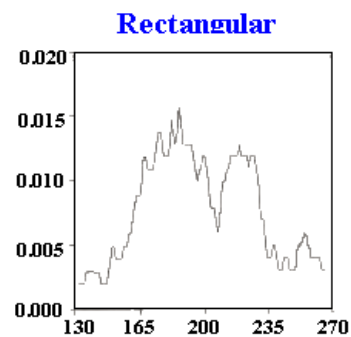
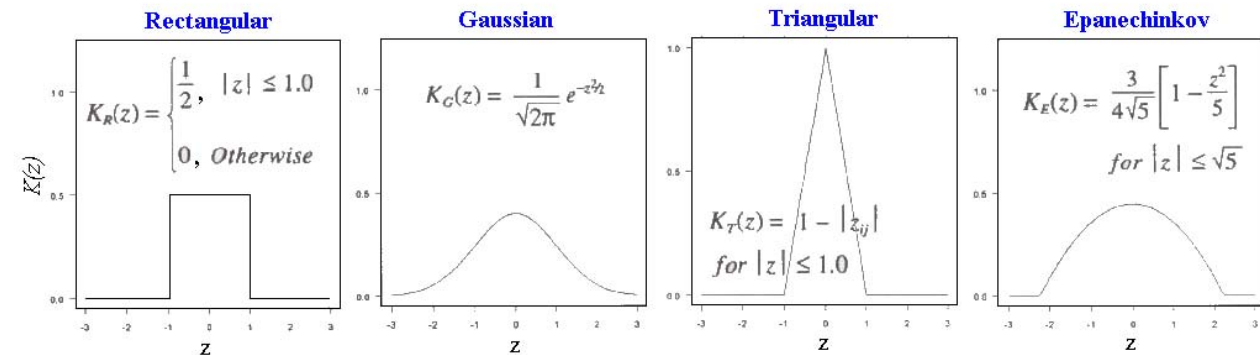


Density Plots (2/3)

- Selection of kernels
- Selection of bandwidth

$$\hat{f}(v_j) = \frac{1}{hn} \sum_{i=1}^n K[z_{ij}]$$

$$z_{ij} = \frac{1}{h} (v_j - x_i)$$



Figures modified from Jacoby (1997)

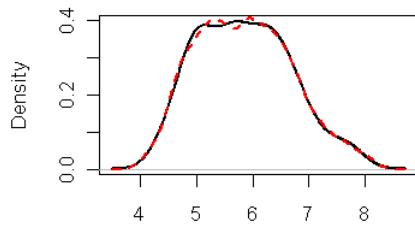
Density Plots (3/3)

```
density(x, bw = "nrd0", adjust = 1,
        kernel = c("gaussian", "epanechnikov", "rectangular",
                  "triangular", "biweight",
                  "cosine", "optcosine"),
        weights = NULL, window = kernel, width,
        give.Rkern = FALSE,
        n = 512, from, to, cut = 3, na.rm = FALSE, ...)
```

— gaussian
 - - - epanechnikov

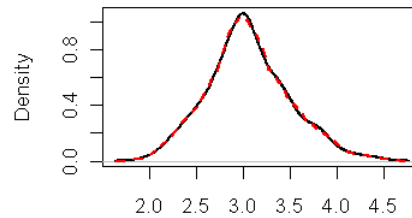
```
> plot(density(iris$Sepal.Length))
```

Sepal.Length



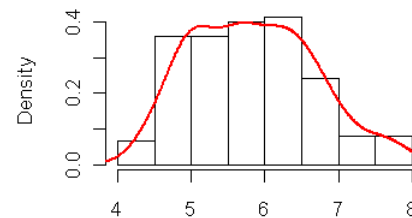
N = 150 Bandwidth = 0.2736

Sepal.Width

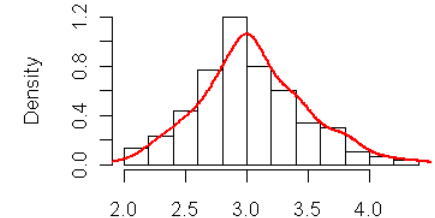


N = 150 Bandwidth = 0.1233

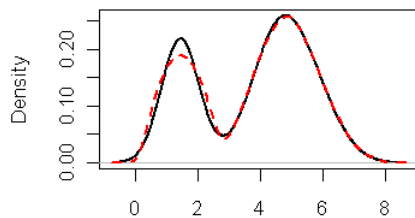
Sepal.Length



Sepal.Width

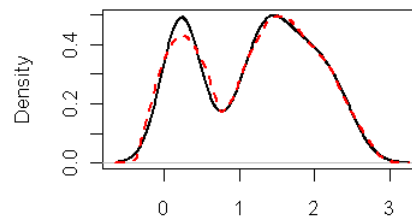


Petal.Length



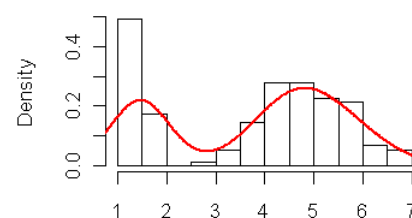
N = 150 Bandwidth = 0.5832

Petal.Width

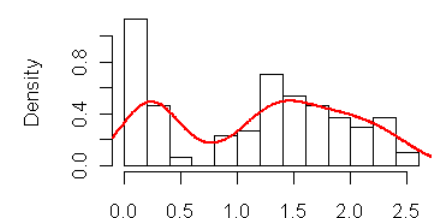


N = 150 Bandwidth = 0.2518

Petal.Length

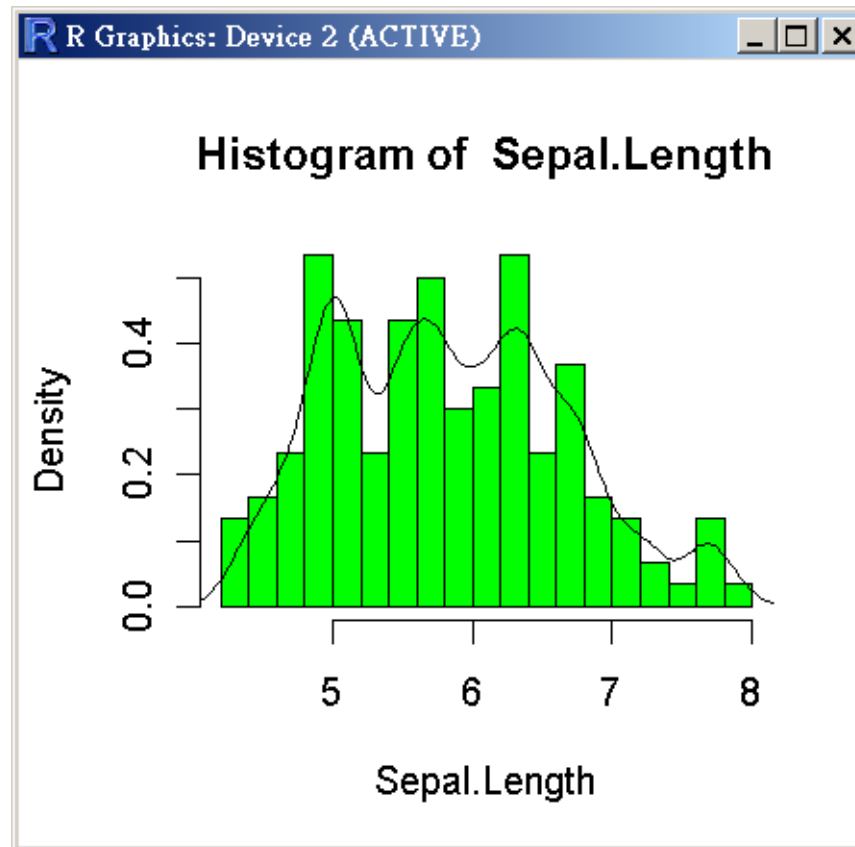


Petal.Width



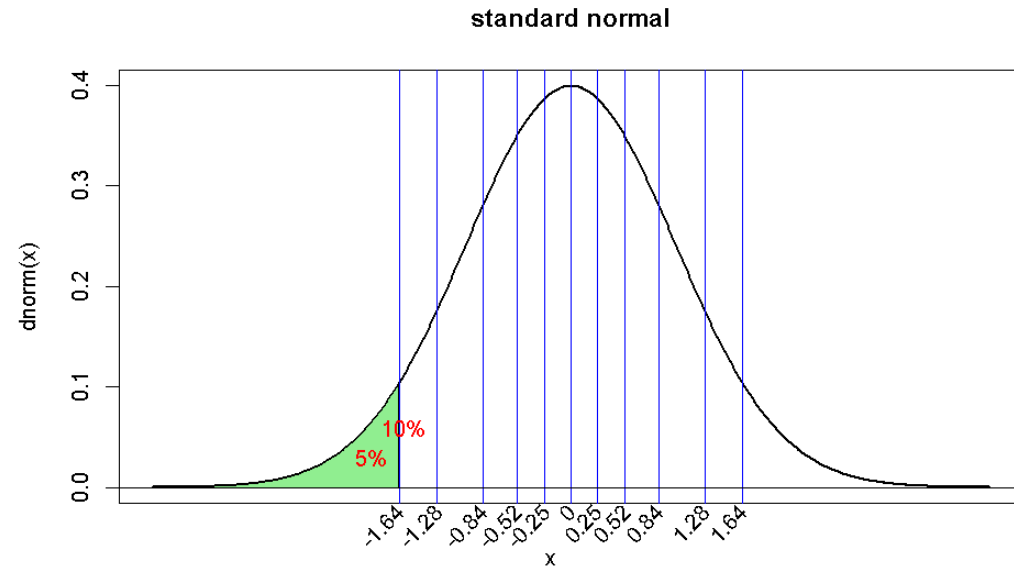
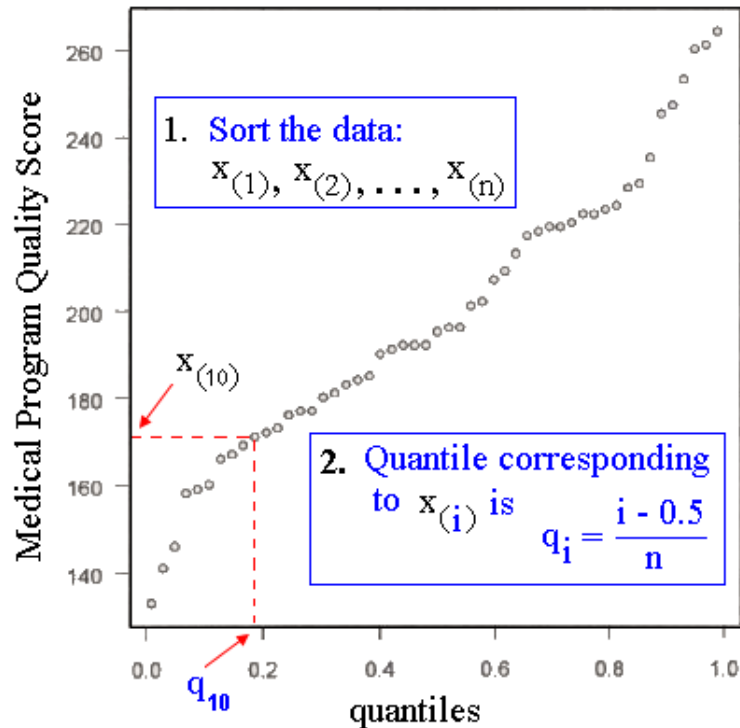
Density Estimation for Continuous Variables

```
> hist(iris[,1], breaks=15, main=title, xlab=lab, col="green", pro=T)
> lines(density(iris[,1], width=0.6, n=200))
```



Quantile Plots

The empirical quantiles

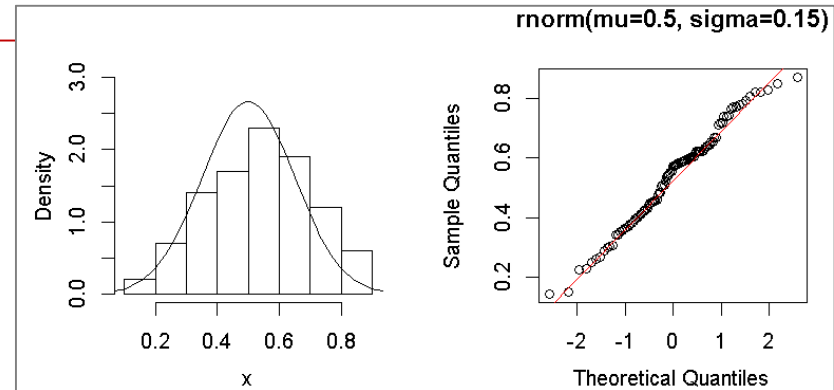


- 0.5 is subtracted from each i value to avoid extreme quantiles of exactly 0 or 1.
- The latter would cause problems if empirical quantiles were to be compared against quantiles derived from a theoretical asymptotic distribution such as the normal.
- This adjustment has no effect on the shape of any graphical display.

```

> par(mfrow = c(1, 2))
> set.seed(12345);
> n <- 100; mu <- 0.5; sigma <- 0.15
> x <- rnorm(n, mu, sigma)
> hist(x, freq=FALSE, ylim=c(0, 3), main="")
> y <- seq(0, 1, length = n)
> lines(y, dnorm(y, mu, sigma), type = 'l')
> qqnorm(x, main = "rnorm(mu=0.5, sigma=0.15)");
> qqline(x)

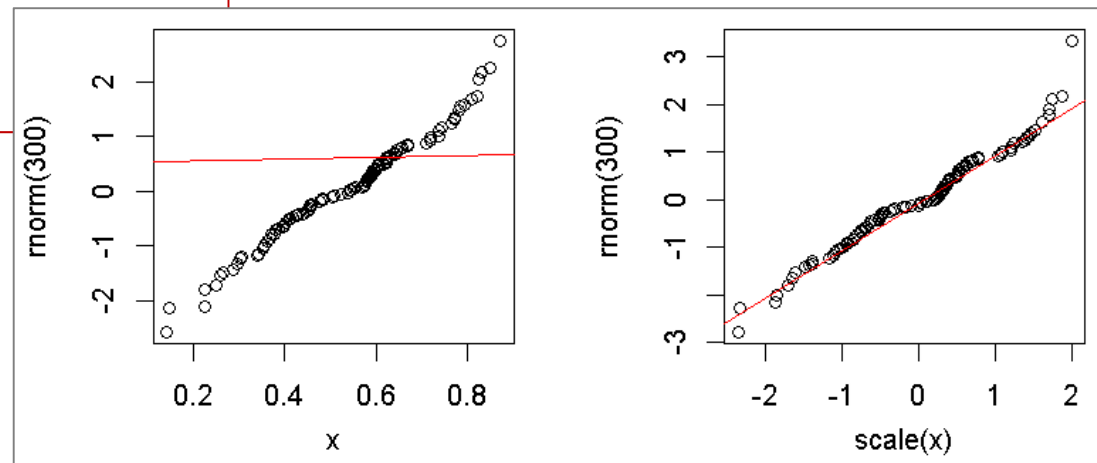
```



```

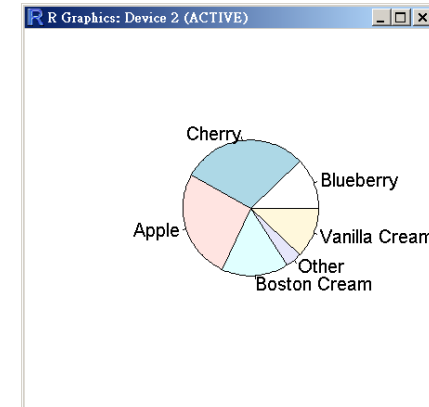
> qqplot(x, rnorm(300))
> qqline(x, col = 2)
> qqplot(scale(x), rnorm(300))
> qqline(scale(x), col = 2)

```

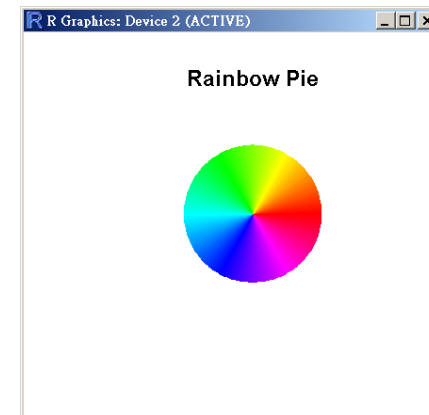
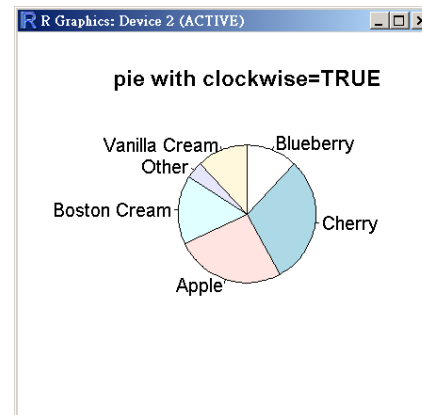
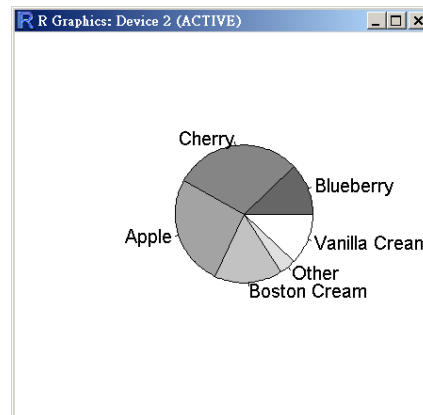
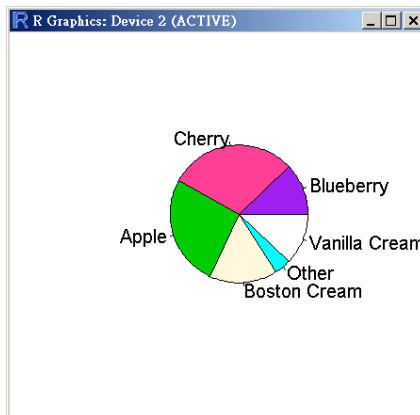


圓餅圖 (Pie Charts)

```
> pie.sales <- c(0.12, 0.3, 0.26, 0.16, 0.04, 0.12)
> sum(pie.sales)
> names(pie.sales) <- c("Blueberry", "Cherry",
  "Apple", "Boston Cream", "Other", "Vanilla Cream")
> pie(pie.sales) # default colours
```



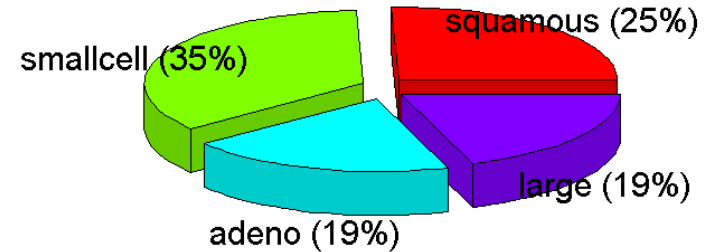
```
> pie(pie.sales, col = c("purple", "violetred1", "green3", "cornsilk", "cyan", "white"))
> pie(pie.sales, col = gray(seq(0.4,1.0,length=6)))
> pie(pie.sales, clockwise=TRUE, main="pie with clockwise=TRUE")
> pie(rep(1,200), labels="", col=rainbow(200), border=NA, main = "Rainbow Pie")
```



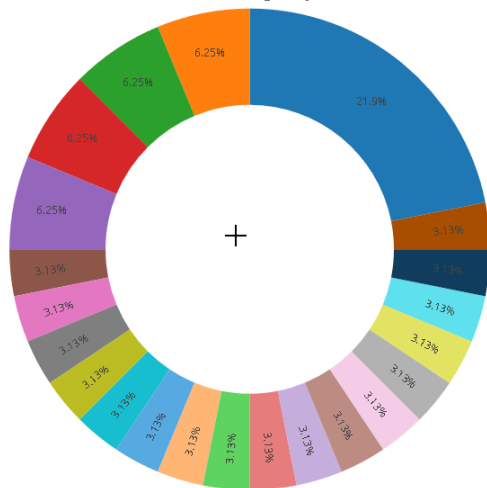
```

> library(plotrix)
> library(survival)
> head(veteran)
  trt celltype time status karno diagtime age prior
1   1 squamous  72      1    60         7  69     0
2   1 squamous 411      1    70         5  64    10
3   1 squamous 228      1    60         3  38     0
4   1 squamous 126      1    60         9  63    10
5   1 squamous 118      1    70        11  65    10
6   1 squamous  10      1    20         5  49     0

> slices <- summary(veteran$celltype)
> p <- floor(100*slices/sum(slices))
> pie3D(slices, labels=paste0(names(slices), " (",p, "%)"), explode=0.1)
    
```

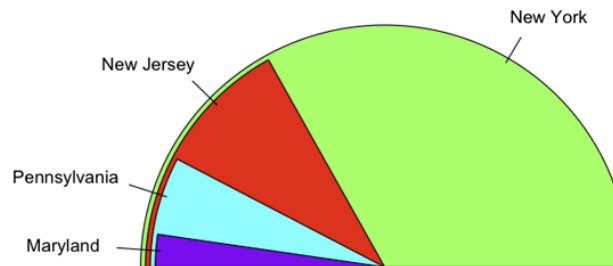


Donut charts using Plotly

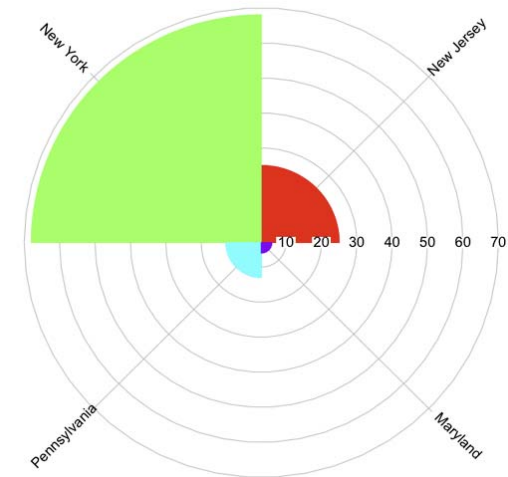


<https://plot.ly/r/pie-charts/>

fan.plot

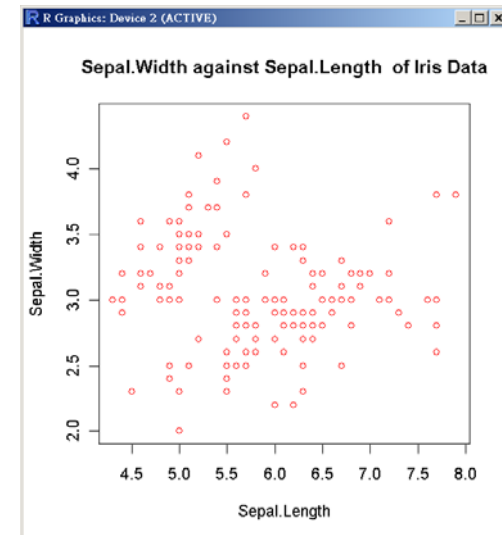


radial.pie

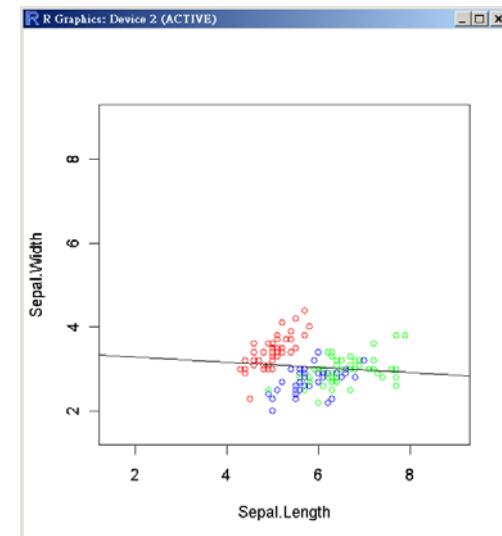


`plot(x, y)` 或 `plot(y~x)`

```
> xlab <- names(iris)[1]
> ylab <- names(iris)[2]
> title <- paste(ylab, "against", xlab, " of Iris
  Data")
> x <- iris[,1]
> y <- iris[,2]
> plot(x, y, col="red", xlab=xlab, ylab=ylab,
  main=title)
```



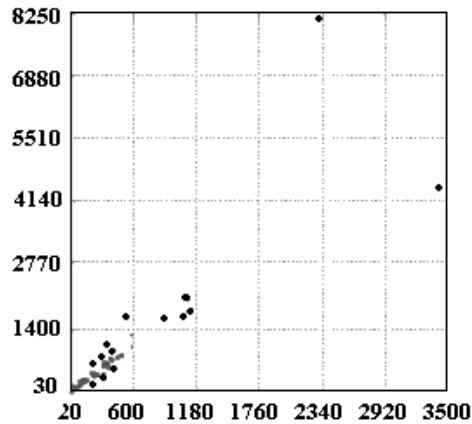
```
> range(x)
> range(y)
> plot(y~x, xlab=xlab, ylab=ylab, xlim=c(1.5,9),
  ylim=c(1.5,9), type="n")
> points(x[1:50], y[1:50], col="red")
> points(x[51:100], y[51:100], col="blue")
> points(x[101:150], y[101:150], col="green")
> abline(lm(y~x))
```



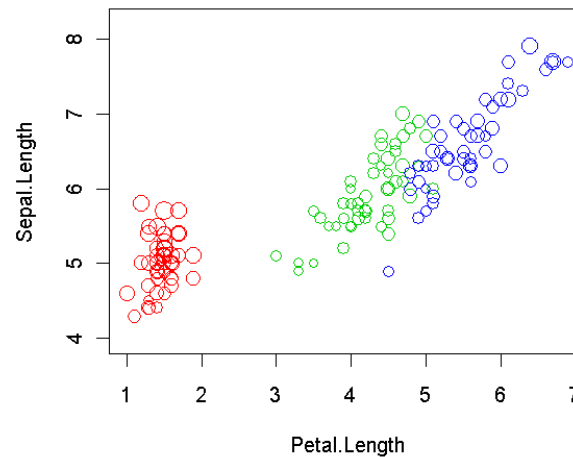
Extensions of Scatterplots

With a smoothing curve

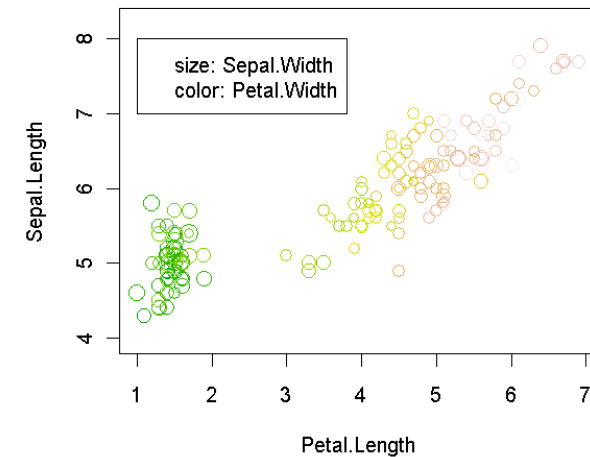
Poorly-Constructed



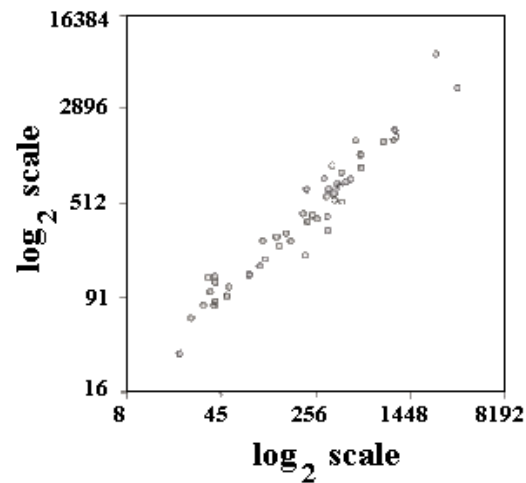
Bubbleplot: Sepal.Width



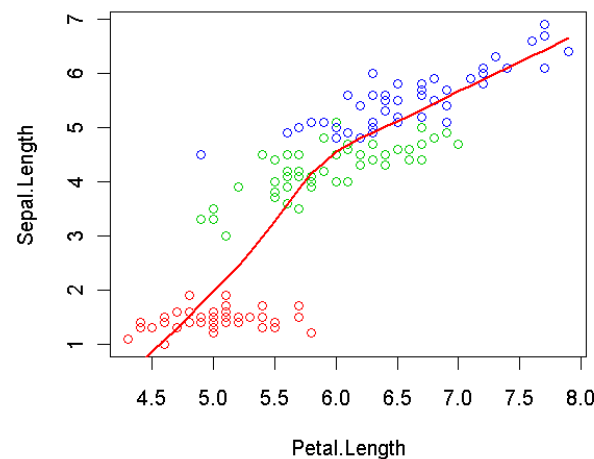
Color Bubbleplot



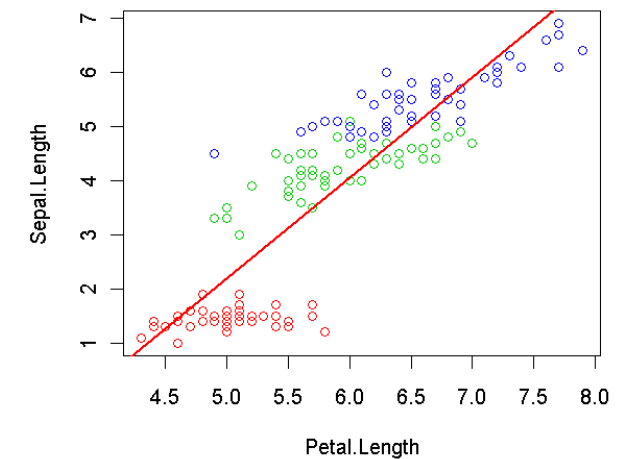
Better



LOWESS

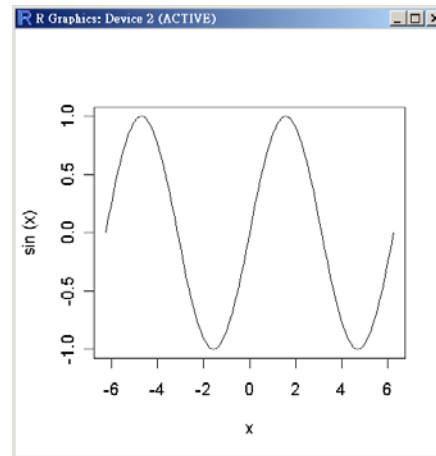
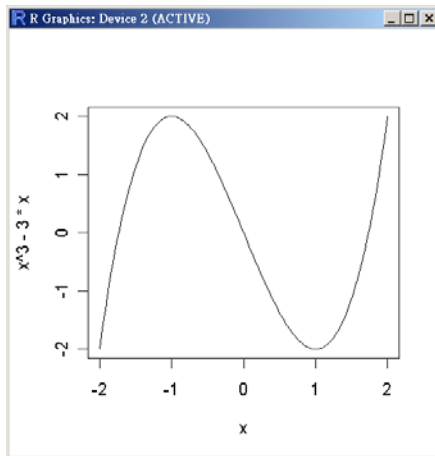


Simple Linear Regression



curve {graphics}: 畫2D數學函數圖

```
> curve(x^3-3*x, -2, 2)
> curve(sin, -2*pi, 2*pi)
```



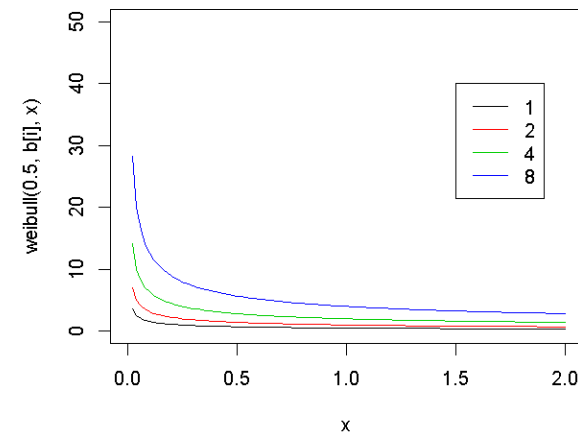
```
> x <- seq(-2, 2, 0.01)
> y <- x^3-3*x
> plot(x, y, type="l")
```

The probability density function of a Weibull random variable is:^[1]

$$f(x; \lambda, k) = \begin{cases} \frac{k}{\lambda} \left(\frac{x}{\lambda}\right)^{k-1} e^{-(x/\lambda)^k}, & x \geq 0, \\ 0, & x < 0, \end{cases}$$

where $k > 0$ is the *shape parameter* and $\lambda > 0$ is the *scale parameter* of the distribution. Its

alpha=5



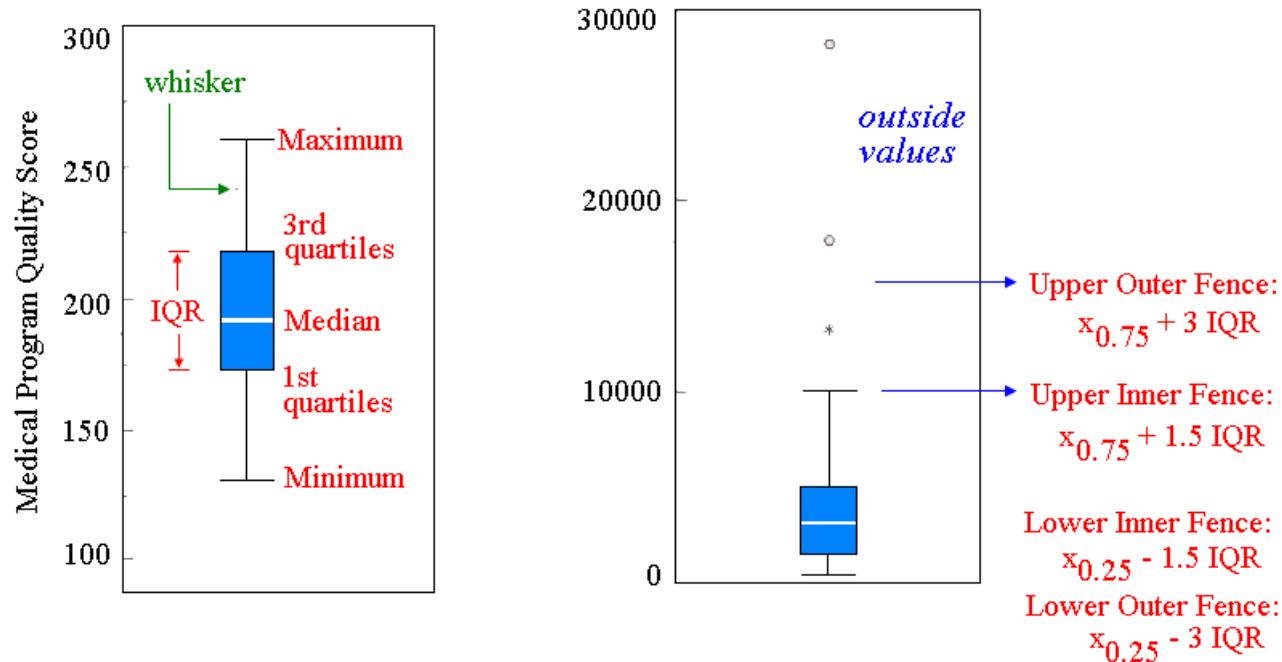
$$f(x; k, b) = bkx^{k-1}e^{-bx^k},$$

the hazard function is $h(x; k, b) = bkx^{k-1}$,

```
> weibull <- function(alpha, beta, x){
+   alpha * beta * (x^(alpha-1))
+ }
>
> b <- c(1, 2, 4, 8)
> for(i in 1:length(b)) {
+   curve(weibull(0.5, b[i], x), from=0, to=2,
+         add=(i!=1),
+         col=i, ylim=c(0, 50), main="alpha=.5")
+ }
>
> legend(1.5, 40, legend=b, col=1:length(b), lty=1)
```

盒形圖 (Boxplot)

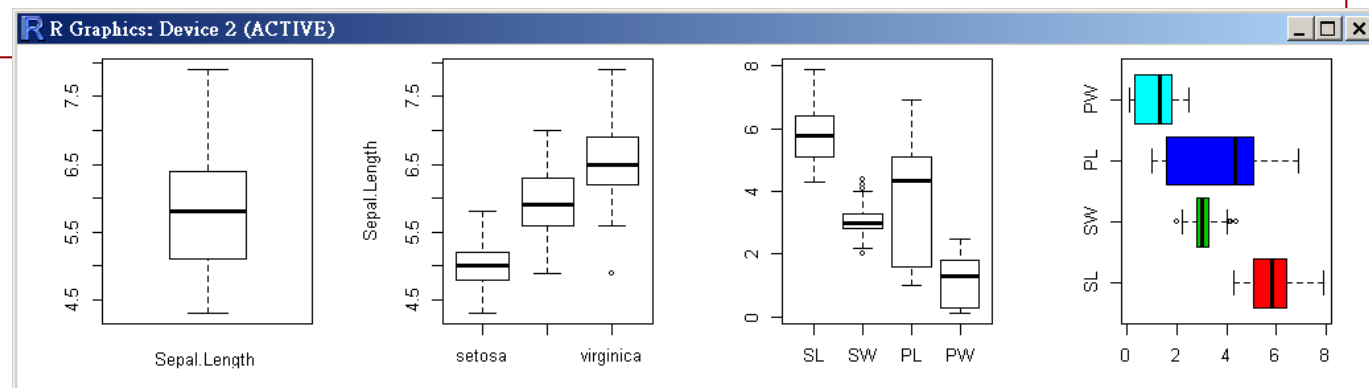
- Plotting with a categorical explanatory variable: boxplot.
- Categorical variables are factors with two or more levels.
- Boxplot
 - The horizontal line shows the median.
 - The bottom and top of the box show the 25th and 75th percentiles.
 - The vertical dashed lines are called the "whiskers".
 - Either maximum or 1.5 times the IQR.
- boxplot not only show the **location and spread of data** but also indicate **skewness**.



boxplot(x, ...)

```
## S3 method for class 'formula'
boxplot(formula, data = NULL, ..., subset, na.action = NULL)
## Default S3 method:
boxplot(x, ..., range = 1.5, width = NULL, varwidth = FALSE,
        notch = FALSE, outline = TRUE, names, plot = TRUE,
        border = par("fg"), col = NULL, log = "",
        pars = list(boxwex = 0.8, staplewex = 0.5, outwex = 0.5),
        horizontal = FALSE, add = FALSE, at = NULL)
```

```
> par(mfrow=c(1,4))
> names(iris)
[1] "Sepal.Length" "Sepal.Width" "Petal.Length" "Petal.Width" "Species"
> attach(iris)
> boxplot(Sepal.Length, xlab="Sepal.Length")
> boxplot(Sepal.Length~Species, ylab="Sepal.Length")
> names(iris) <- c("SL", "SW", "PL", "PW", "Species")
> boxplot(iris[,which(sapply(iris, is.numeric))])
> boxplot(iris[,which(sapply(iris, is.numeric))], horizontal=T, col=2:8)
> detach(iris)
```

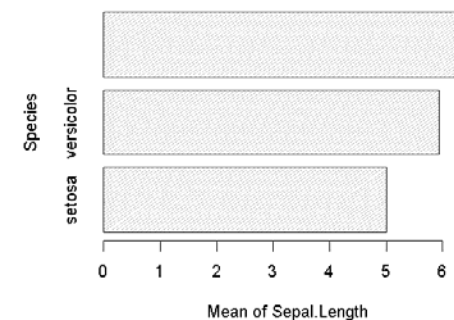
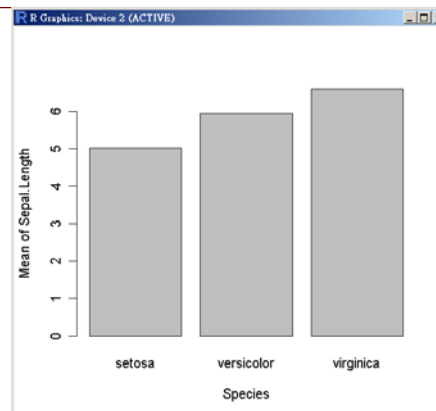


長條圖 (barplot)

```
barplot(height, width = 1, space = NULL,
        names.arg = NULL, legend.text = NULL, beside = FALSE,
        horiz = FALSE, density = NULL, angle = 45,
        col = NULL, border = par("fg"),
        main = NULL, sub = NULL, xlab = NULL, ylab = NULL,
        xlim = NULL, ylim = NULL, xpd = TRUE, log = "",
        axes = TRUE, axisnames = TRUE,
        cex.axis = par("cex.axis"), cex.names = par("cex.axis"),
        inside = TRUE, plot = TRUE, axis.lty = 0, offset = 0,
        add = FALSE, args.legend = NULL, ...)
```

```
> data(iris)
> means <- tapply(iris$Sepal.Length, iris$Species, mean)
> barplot(means, xlab="Species", ylab="Mean of Sepal.Length")
> # density: a vector giving the density of shading lines
> barplot(means, ylab="Species", xlab="Mean of Sepal.Length", density=20,
          horiz=TRUE)
```

```
> means
  setosa versicolor virginica
  5.006   5.936   6.588
```



長條圖 (barplot)

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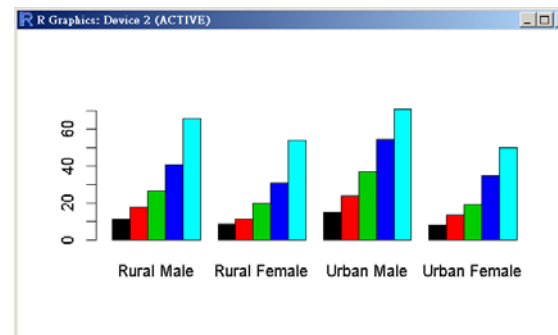
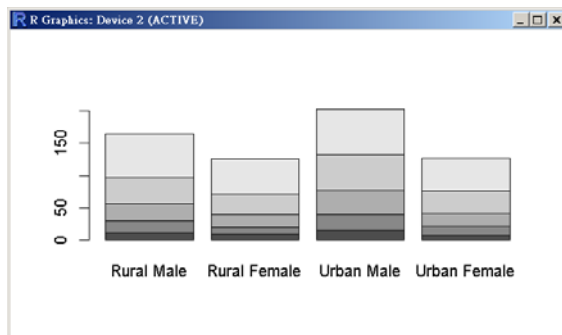
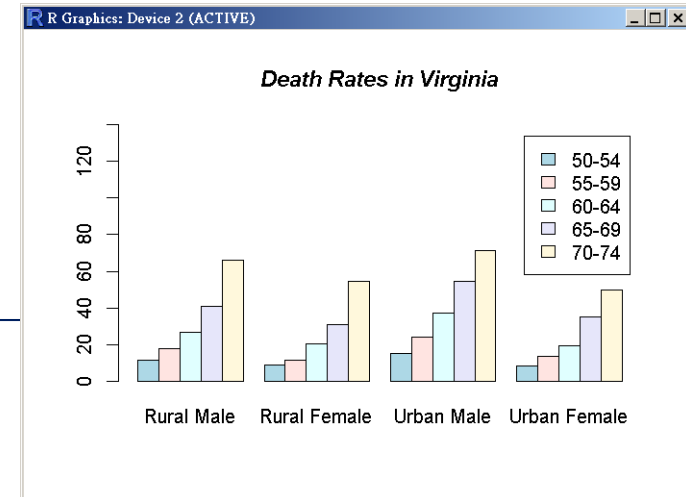
Death Rates in Virginia (1940): The death rates are measured per 1000 population per year. They are cross-classified by age group (rows) and population group (columns). The age groups are: 50–54, 55–59, 60–64, 65–69, 70–74 and the population groups are Rural/Male, Rural/Female, Urban/Male and Urban/Female.

```
> VADeaths
```

	Rural Male	Rural Female	Urban Male	Urban Female
50-54	11.7	8.7	15.4	8.4
55-59	18.1	11.7	24.3	13.6
60-64	26.9	20.3	37.0	19.3
65-69	41.0	30.9	54.6	35.1
70-74	66.0	54.3	71.1	50.0

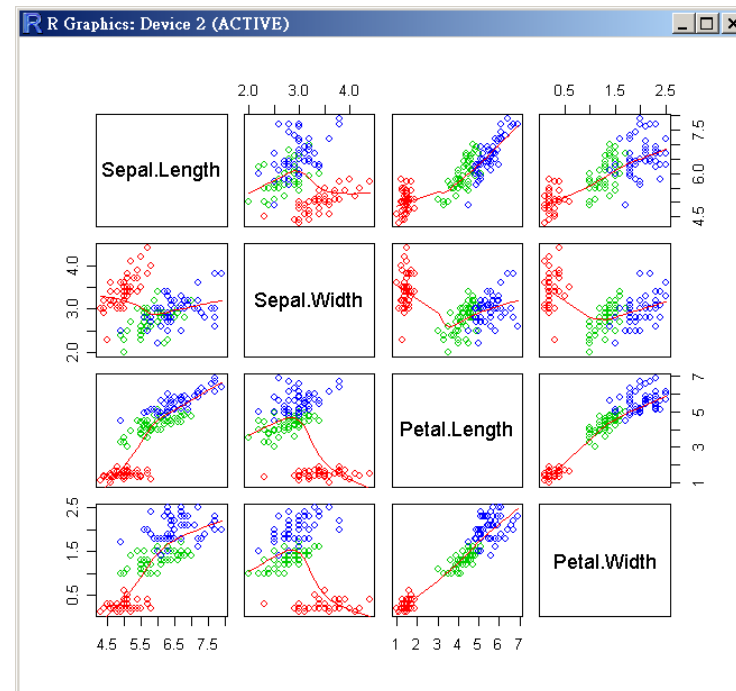
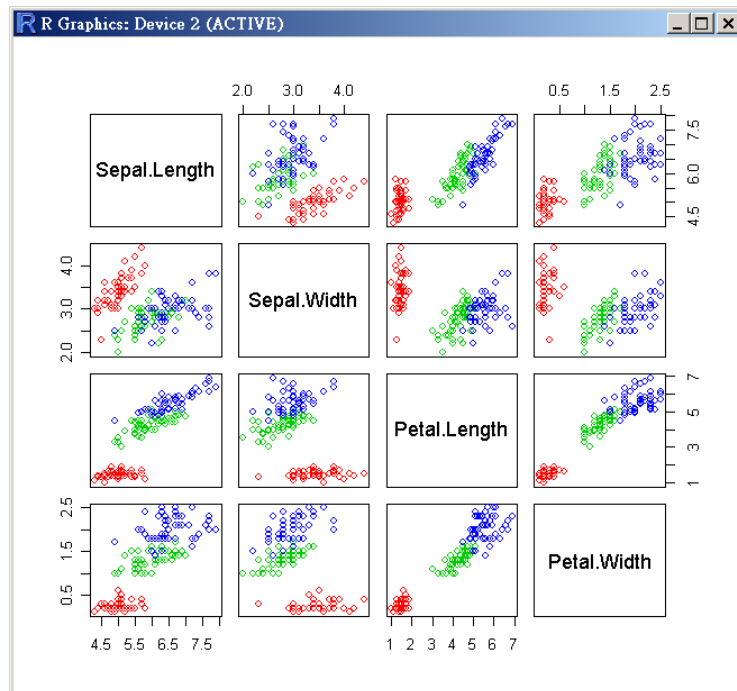
```
barplot(VADeaths)
barplot(VADeaths, beside = TRUE, col=1:5)

barplot(VADeaths, beside = TRUE,
        col = c("lightblue", "mistyrose", "lightcyan",
                "lavender", "cornsilk"),
        legend = rownames(VADeaths), ylim = c(0, 140))
title(main = "Death Rates in Virginia", font.main = 4)
```



散佈圖矩陣 (Scatterplot Matrices)^{60/86}

```
> pairs(iris[,1:4], col=as.integer(iris[,5])+1)
> pairs(iris[,1:4], col=as.integer(iris[,5])+1,
       panel=panel.smooth)
```

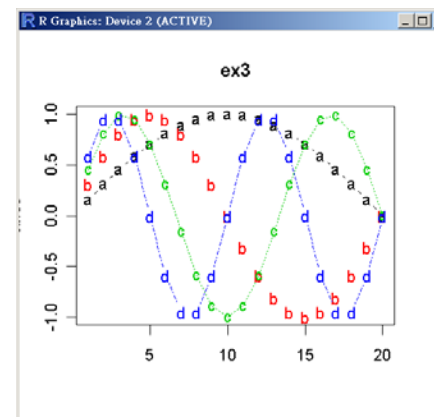
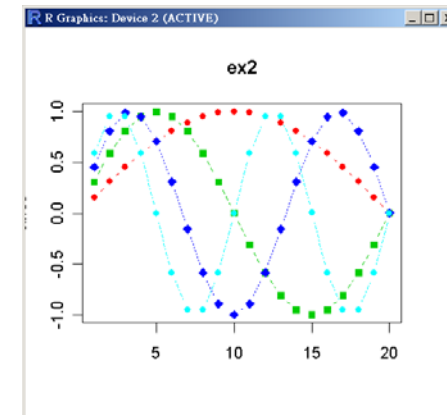
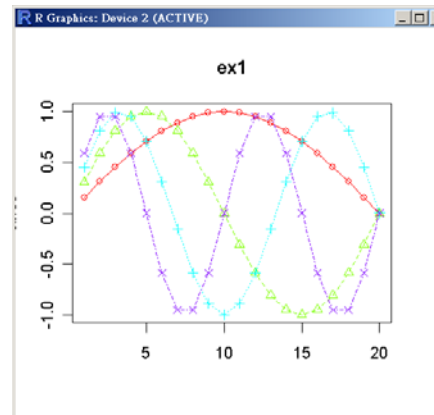


See also: conditioning plot (`coplot`)

Plot Columns of Matrices

```
sines <- outer(1:20, 1:4, function(x, y) sin(x / 20 * pi * y))
dim(sines)
sines
matplot(sines, pch = 1:4, type = "o", col = rainbow(ncol(sines)), main="ex1")
matplot(sines, pch = 21:23, type = "b", col = 2:5, bg= 2:5, main="ex2")
```

```
> sines <- outer(1:20, 1:4, function(x, y) sin(x / 20 * pi * y))
> sines
      [,1]      [,2]      [,3]      [,4]
[1,] 1.564345e-01 3.090170e-01 4.539905e-01 5.877853e-01
[2,] 3.090170e-01 5.877853e-01 8.090170e-01 9.510565e-01
[3,] 4.539905e-01 8.090170e-01 9.876883e-01 9.510565e-01
[4,] 5.877853e-01 9.510565e-01 9.510565e-01 5.877853e-01
[5,] 7.071068e-01 1.000000e+00 7.071068e-01 1.224606e-16
[6,] 8.090170e-01 9.510565e-01 3.090170e-01 -5.877853e-01
[7,] 8.910065e-01 8.090170e-01 -1.564345e-01 -9.510565e-01
[8,] 9.510565e-01 5.877853e-01 -5.877853e-01 -9.510565e-01
[9,] 9.876883e-01 3.090170e-01 -8.910065e-01 -5.877853e-01
[10,] 1.000000e+00 1.224606e-16 -1.000000e+00 -2.449213e-16
[11,] 9.876883e-01 -3.090170e-01 -8.910065e-01 5.877853e-01
[12,] 9.510565e-01 -5.877853e-01 -5.877853e-01 9.510565e-01
[13,] 8.910065e-01 -8.090170e-01 -1.564345e-01 9.510565e-01
[14,] 8.090170e-01 -9.510565e-01 3.090170e-01 5.877853e-01
[15,] 7.071068e-01 -1.000000e+00 7.071068e-01 3.673819e-16
[16,] 5.877853e-01 -9.510565e-01 9.510565e-01 -5.877853e-01
[17,] 4.539905e-01 -8.090170e-01 9.876883e-01 -9.510565e-01
[18,] 3.090170e-01 -5.877853e-01 8.090170e-01 -9.510565e-01
[19,] 1.564345e-01 -3.090170e-01 4.539905e-01 -5.877853e-01
[20,] 1.224606e-16 -2.449213e-16 3.673819e-16 -4.898425e-16
> |
```



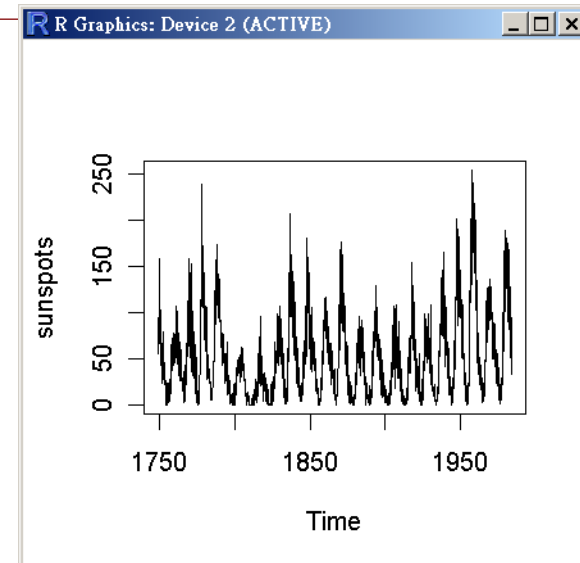
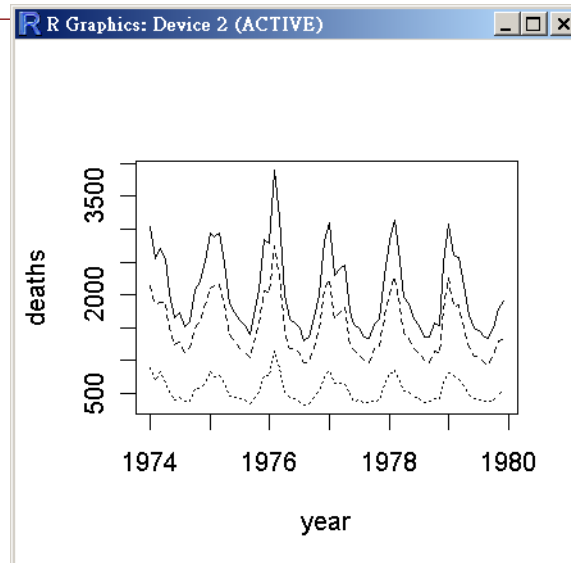
Exercise: adding legends?

- `ts.plot`: simple vector of numbers
- `plot.ts` works for plotting objects inheriting from class `ts`.

```
> data(UKlungDeaths) # total, male, female death
> ts.plot(ldeaths, mdeaths, fdeaths, xlab="year", ylab="deaths", lty=c(1:3))

> data(sunspots)
> plot(sunspots) # sunspots is ts class
> class(sunspots)
[1] "ts"
> is.ts(sunspots)
[1] TRUE
```

```
data(UKgas)
attach(UKgas)
names(UKgas)
```



Time Series Plots

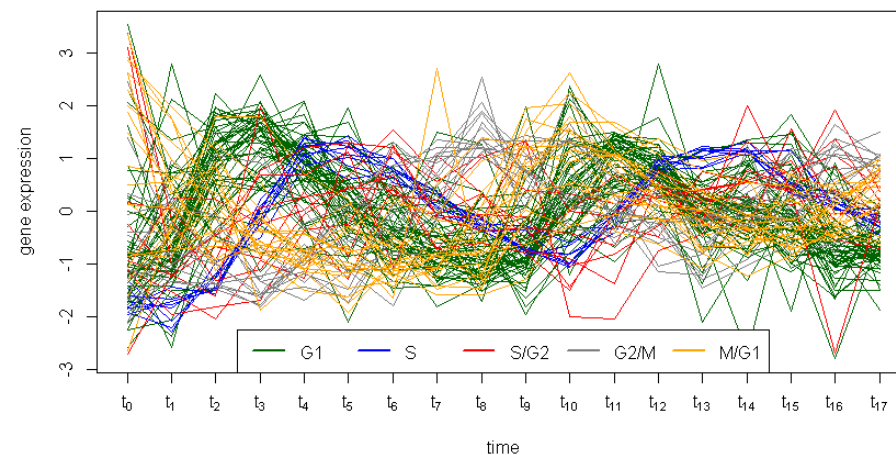
```

cell.raw <- read.table("trad_alpha103.txt", row.names=1, header=T)
head(cell.raw)
cell.xdata <- t(scale(t(cell.raw[,2:19]), center=T, scale=T))
y.C <- as.integer(cell.raw[,1])
table(y.C)
no.cluster <- length(unique(y.C))
p <- ncol(cell.raw) - 1
cellcycle.color <- c("darkgreen", "blue", "red", "gray50", "orange")
ycolors <- cellcycle.color[y.C+1]
my.pch <- c(1:no.cluster)[y.C+1]
phase <- c("G1", "S", "S/G2", "G2/M", "M/G1")
matplot(t(cell.xdata), lty=1, type = "l", ylab="gene expression",
        col=ycolors, xlab="time", main="Time series", xaxt="n")
time.label <- parse(text=paste("t[",0:p,"]",sep=""))
axis(1, 1:(p+1), time.label)
legend("bottom", legend=phase, col=cellcycle.color, lty=1, horiz = T, lwd=2)

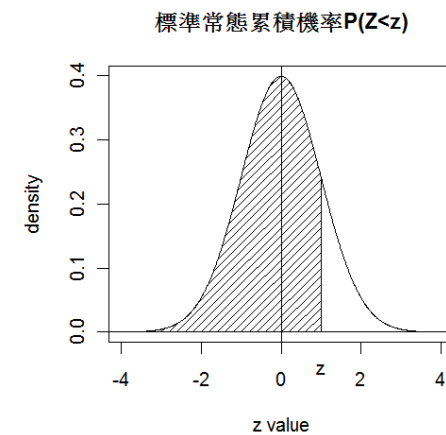
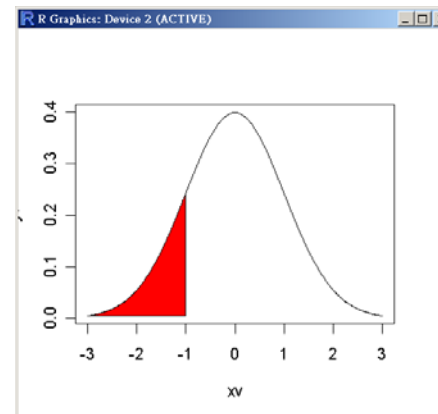
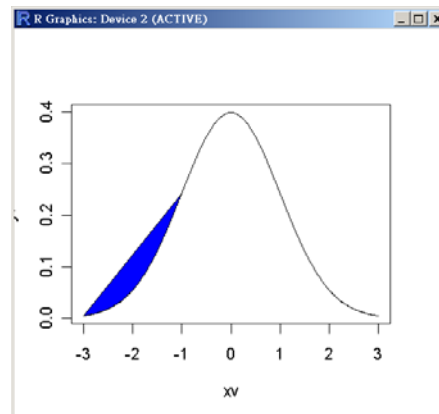
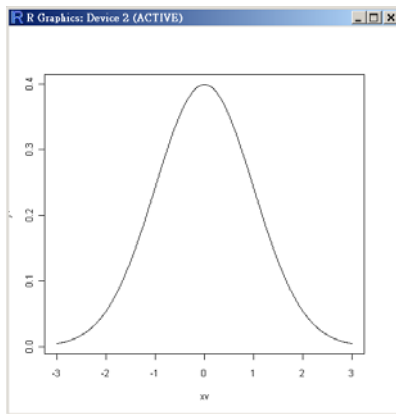
```

	A	B	C	D	E	F	G
1	UID	Phase	alpha0	alpha7	alpha14	alpha21	alpha28
2	YAR007C_G1	0	-0.48	-0.42	0.87	0.92	0.67
3	YBL035C_G1	0	-0.39	-0.58	1.08	1.21	0.52
4	YBR023C_G1	0	0.87	0.25	-0.17	0.18	-0.13
5	YBR067C_G1	0	1.57	1.03	1.22	0.31	0.16
6	YBR088C_G1	0	-1.15	-0.86	1.21	1.62	1.12
7	YBR278W_G1	0	0.04	-0.12	0.31	0.16	0.17
8	YCL055W_G1	0	2.95	0.45	-0.40	-0.66	-0.59
9	YDL003W_G1	0	-1.22	-0.74	1.34	1.50	0.63
10	YDL055C_G1	0	-0.73	-1.06	-0.79	-0.02	0.16
11	YDL102W_G1	0	-0.58	-0.40	0.13	0.58	-0.09
12	YDL164C_G1	0	-0.50	-0.42	0.66	1.05	0.68
13	YDL197C_G1	0	-0.86	-0.29	0.42	0.46	0.30
14	YDL227C_G1	0	-0.16	0.29	0.17	-0.28	-0.02
15	YDR052C_G1	0	-0.36	-0.03	-0.03	-0.08	-0.23

Time series



```
# draw a polygon using mouse
> plot(1)
> locations <- locator(6)
> polygon(locations, col="lavender")
```



```
> xv <- seq(-3, 3, 0.01)
> yv <- dnorm(xv)
> plot(xv, yv, type="l")

> polygon(c(xv[xv <= -1]), c(yv[xv <= -1]), col="blue")

> plot(xv, yv, type="l")
> polygon(c(xv[xv <= -1], -1), c(yv[xv <= -1], yv[xv== -3]), col="red")
```




Radar Plot using **fmsb** Package

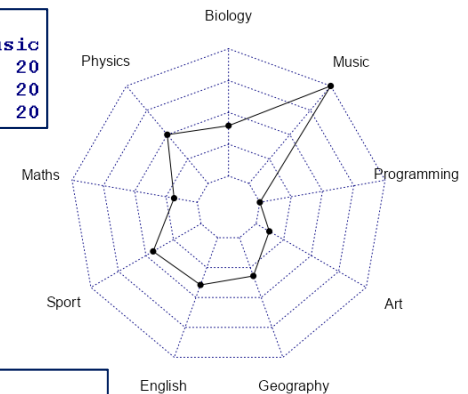
```
scores <- data.frame(
  row.names = c("Student.1", "Student.2", "Student.3"),
  Biology = c(7.9, 3.9, 9.4),
  Physics = c(10, 20, 0),
  Maths = c(3.7, 11.5, 2.5),
  Sport = c(8.7, 20, 4),
  English = c(7.9, 7.2, 12.4),
  Geography = c(6.4, 10.5, 6.5),
  Art = c(2.4, 0.2, 9.8),
  Programming = c(0, 0, 20),
  Music = c(20, 20, 20)
)
```

- library(fmsb)
- The data should be organized as follow:
- The row 1 must contain the maximum values for each variable
- The row 2 must contain the minimum values for each variable
- Data for cases or individuals should be given starting from row 3
- The number of columns or variables must be more than 2.

```
max.min <- data.frame(
  Biology = c(20, 0), Physics = c(20, 0), Maths = c(20, 0),
  Sport = c(20, 0), English = c(20, 0), Geography = c(20, 0),
  Art = c(20, 0), Programming = c(20, 0), Music = c(20, 0)
)
rownames(max.min) <- c("Max", "Min")
max.min
```

```
> scores
```

	Biology	Physics	Maths	Sport	English	Geography	Art	Programming	Music
Student.1	7.9	10	3.7	8.7	7.9	6.4	2.4	0	20
Student.2	3.9	20	11.5	20.0	7.2	10.5	0.2	0	20
Student.3	9.4	0	2.5	4.0	12.4	6.5	9.8	20	20

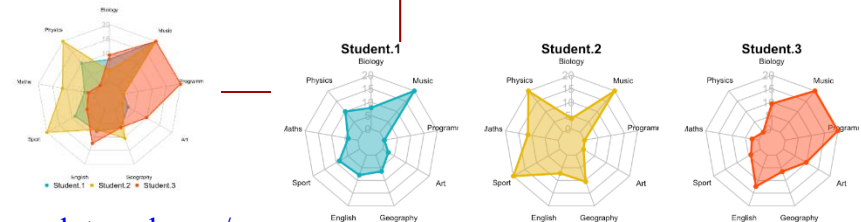


```
mydata <- rbind(max.min, scores)
mydata
```

```
> mydata
```

	Biology	Physics	Maths	Sport	English	Geography	Art	Programming	Music
Max	20.0	20	20.0	20.0	20.0	20.0	20.0	20	20
Min	0.0	0	0.0	0.0	0.0	0.0	0.0	0	0
Student.1	7.9	10	3.7	8.7	7.9	6.4	2.4	0	20
Student.2	3.9	20	11.5	20.0	7.2	10.5	0.2	0	20
Student.3	9.4	0	2.5	4.0	12.4	6.5	9.8	20	20

```
library(fmsb)
student1.data <- mydata[c("Max", "Min", "Student.1"), ]
student1.data
radarchart(student1.data)
```

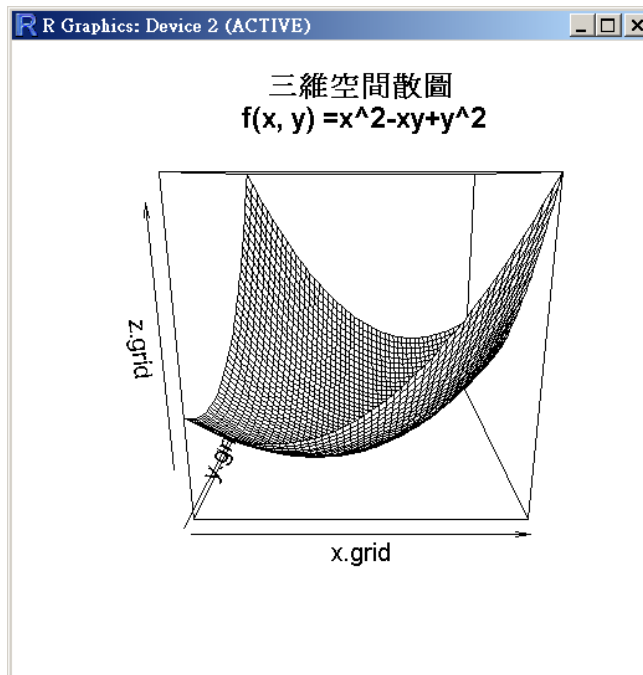


Beautiful Radar Chart in R using FMSB and GGPlot Packages

<https://www.datanovia.com/en/blog/beautiful-radar-chart-in-r-using-fmsb-and-ggplot-packages/>

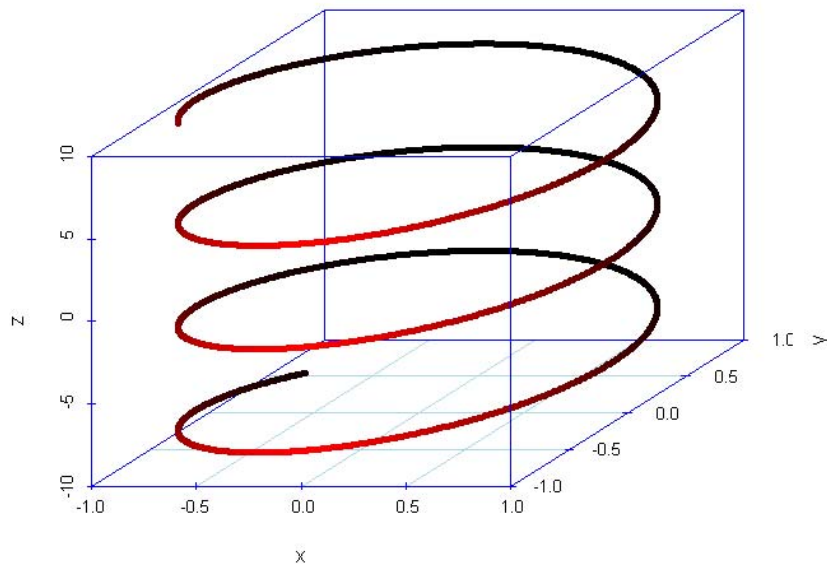
- 雙變數函數在三維空間的散佈圖
- NOTE: x-y平面上，格點數目愈多，散佈圖愈密集。
- plot $f(x, y) = x^2 - xy + y^2$ 。

```
> ploy <- function(x, y){x^2-x*y+y^2}
> x.grid <- seq(-3, 3, length=50)
> y.grid <- seq(-3, 3, length=50)
> z.grid <- outer(x.grid, y.grid, FUN=ploy)
> ploy.title <- paste("三維空間散圖\n", "f(x, y) =x^2-xy+y^2")
> persp(x.grid, y.grid, z.grid, main= ploy.title)
```

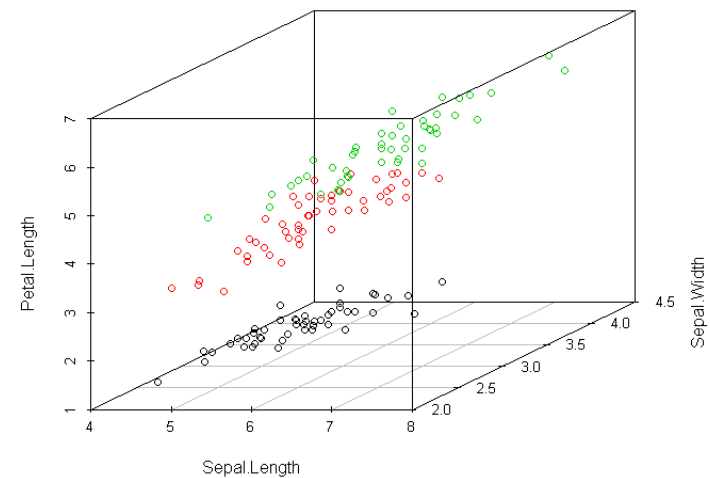


```
> library(scatterplot3d)
> z <- seq(-10, 10, 0.01)
> x <- cos(z)
> y <- sin(z)
> scatterplot3d(x, y, z, highlight.3d=TRUE, col.axis="blue",
  col.grid="lightblue", main="scatterplot3d - 1", pch=20)
```

scatterplot3d - 1



```
> scatterplot3d(iris[,1:3],
  color=as.integer(iris[,5]))
```



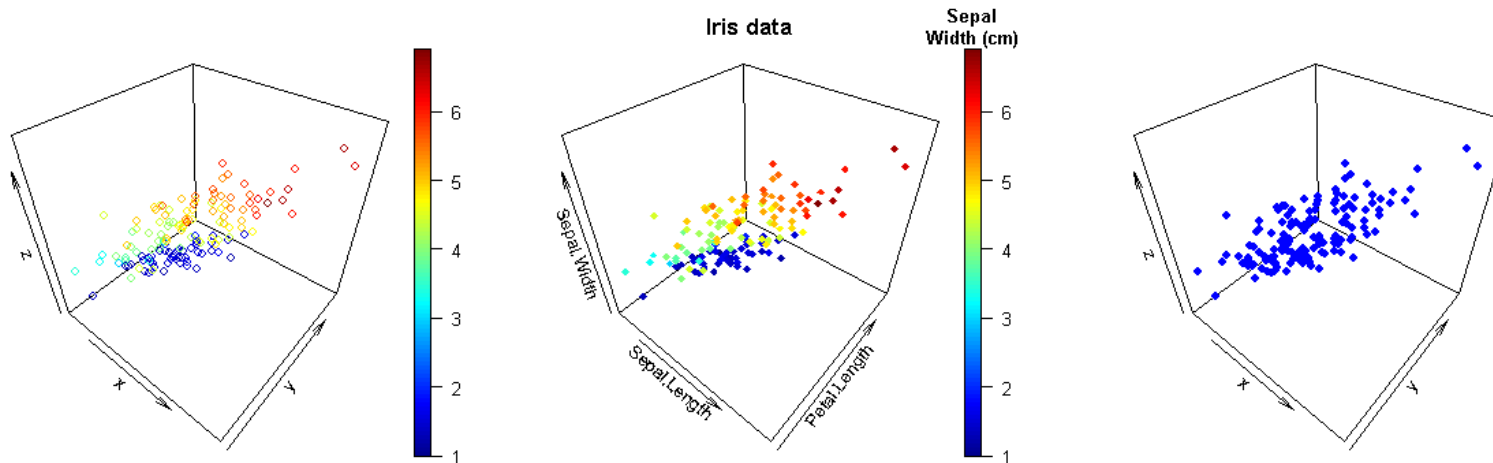
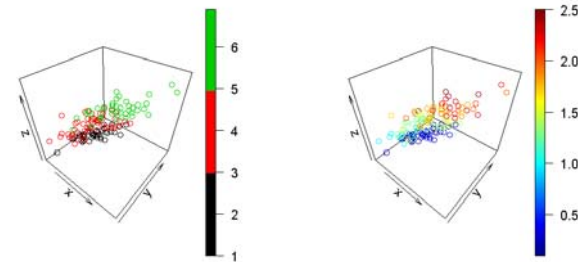
plot3D package

```
install.packages("plot3D")
library("plot3D")
```

```
x <- iris$Sepal.Length
y <- iris$Sepal.Width
z <- iris$Petal.Length
w <- iris$Petal.Width
s <- iris$Species
```

```
> scatter3D(x, y, z, col = as.integer(s))
> scatter3D(x, y, z, colvar = w)
```

```
par(mfrow = c(1,3), mai = c(0.3, 0.3, 0.3, 0.3))
scatter3D(x, y, z)
scatter3D(x, y, z, pch = 18, clab = c("Sepal", "Width (cm)"), main = "Iris data",
          xlab = "Sepal.Length", zlab = "Petal.Length", ylab = "Sepal.Width")
scatter3D(x, y, z, colvar = as.integer(s), col = "blue", pch = 19, cex = 1)
```



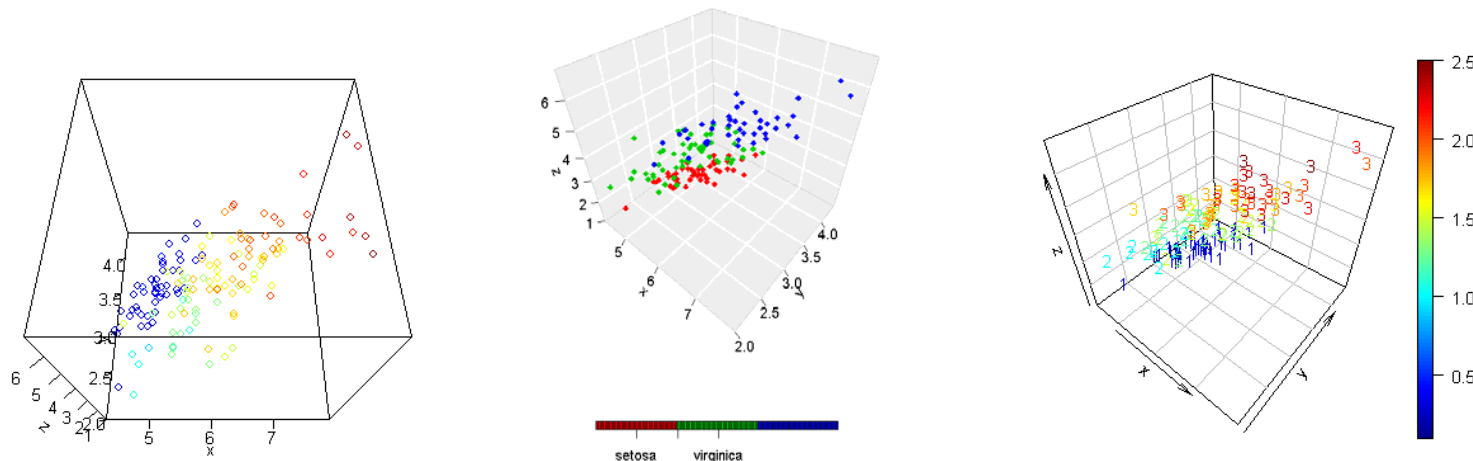
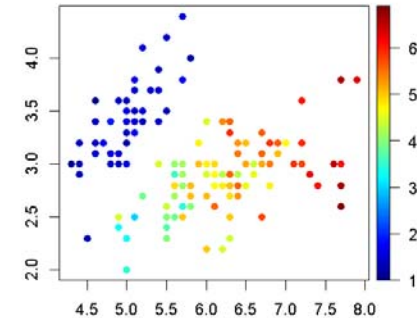
<http://www.sthda.com/english/wiki/impressive-package-for-3d-and-4d-graph-r-software-and-data-visualization>

```

par(mfrow = c(1,3), mai = c(0.3, 0.3, 0.2, 0.3))
# grey background with white grid lines
scatter3D(x, y, z, bty = "f", colkey = FALSE, ticktype = "detailed",
          theta=0, phi=60)
# theta: the azimuthal direction, phi: the co-latitude.
scatter3D(x, y, z, bty = "g", pch = 18,
          col = as.integer(s) + 1,
          ticktype = "detailed",
          colkey = list(at = c(2, 3, 4), side = 1,
                        addlines = TRUE, length = 0.5, width = 0.5,
                        labels = c("setosa", "versicolor", "virginica")),
          text3D(x, y, z, labels = as.integer(s), colvar = w, bty = "b2")
# "b2": back panels and grid lines are visible

# hist3D, text3D
# scatter2D(x, y, colvar = z, pch = 16)

```



More examples: <http://www.sthda.com/english/wiki/impressive-package-for-3d-and-4d-graph-r-software-and-data-visualization>

3D visualization device system (OpenGL)

```

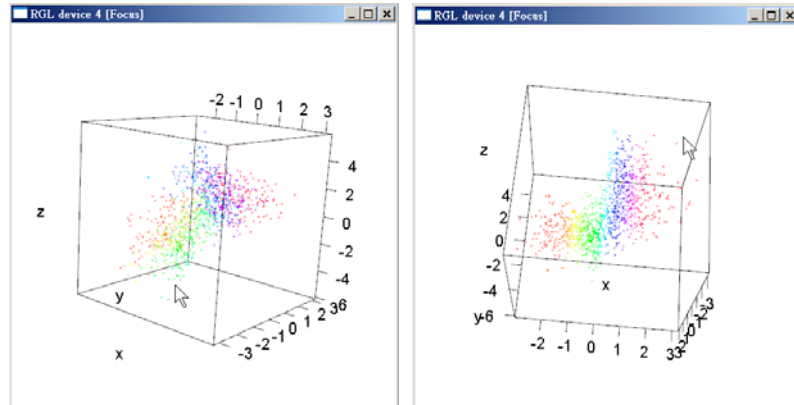
> library(rgl)
> demo(rgl)
> open3d()
> x <- sort(rnorm(1000))
> y <- rnorm(1000)
> z <- rnorm(1000) + atan2(x,y)
> plot3d(x, y, z, col=rainbow(1000), size=2)
>
> M <- par3d("userMatrix")
> play3d(par3dinterp(userMatrix=list(M, rotate3d(M, pi/2, 1, 0, 0),
+ rotate3d(M, pi/2, 0, 1, 0)), duration=4)
+

```

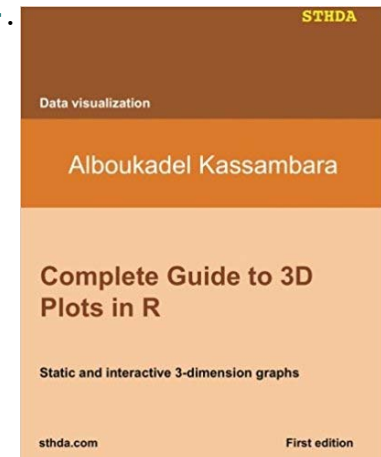
```

> M
      [,1]      [,2]      [,3] [,4]
[1,] -0.98849827 -0.1478247 -0.0319229  0
[2,] -0.01036166 -0.1443882  0.9894670  0
[3,] -0.15087697  0.9784170  0.1411958  0
[4,]  0.00000000  0.0000000  0.0000000  1

```



- 截取靜態2d圖: `rgl.postscript {rgl}`, 或 `rgl.snapshot {rgl}`.
- 存出動態圖: `writeWebGL {rgl}`.



STHDA: rgl

<http://www.sthda.com/english/wiki/a-complete-guide-to-3d-visualization-device-system-in-r-r-software-and-data-visualization>

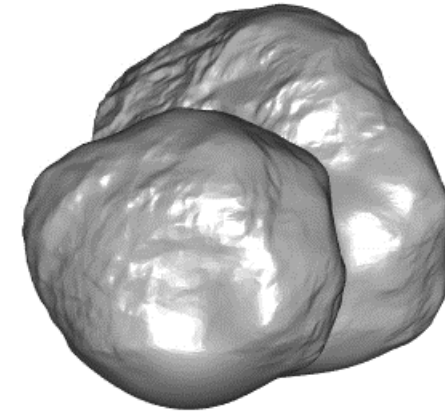
<http://www.sthda.com/english/download/3-ebooks/6-complete-guide-to-3d-plots-in-r/>

Explore a comet with R's "rgl" package

December 24, 2014

<http://blog.revolutionanalytics.com/2014/12/explore-a-comet-with-rs-rgl-package.html>

"Last month, the Philae lander touched down on comet Churyumov–Gerasimenko. In the process, the lander and the orbiting Rosetta probe captured detailed data on the geometry of the comet, which the ESA published as a shape file. ..."



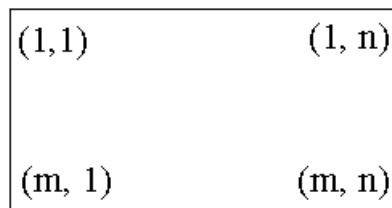
<https://en.wikipedia.org/wiki/67P/Churyumov%E2%80%93Gerasimenko>

```
> open3d()
> # comet <- readOBJ(url("http://sci.esa.int/science-e/www/object/doc.cfm?fobjectid=54726"))
> comet <- readOBJ("ESA_Rosetta_OSIRIS_67P_SHAP2P.obj")
> class(comet)
[1] "mesh3d" "shape3d"
> str(comet)
List of 6
 $ vb          : num [1:4, 1:31456] -0.394 0.402 0.443 1 -0.163 ...
 $ it          : num [1:3, 1:62908] 14327 6959 18747 8258 15598 ...
 $ primitivetype: chr "triangle"
 $ material     : NULL
 $ normals     : NULL
 $ texcoords   : NULL
 - attr(*, "class")= chr [1:2] "mesh3d" "shape3d"
> shade3d(comet, col="gray")
```

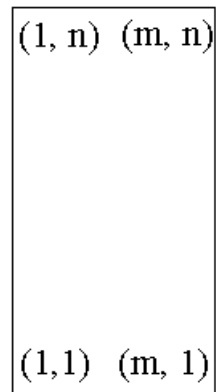
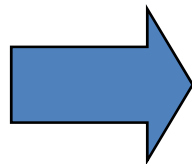
```
# it: indices for triangular faces
# ib: indices for quad faces
# vb: matrix of vertices: 4xn matrix (rows
x, y, z, h) or equivalent vector, where h
indicates scaling of each plotted quad
```

- `image(x, ...)`
- `image(x, y, z, zlim, xlim, ylim, col = heat.colors(12), add = FALSE, xaxs = "i", yaxs = "i", xlab, ylab, breaks, oldstyle = FALSE, ...)`

Data Matrix

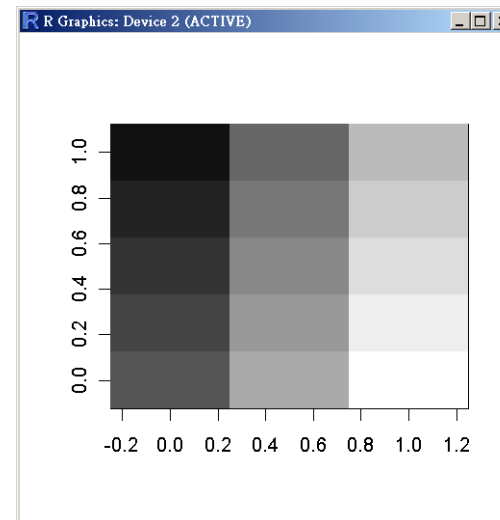
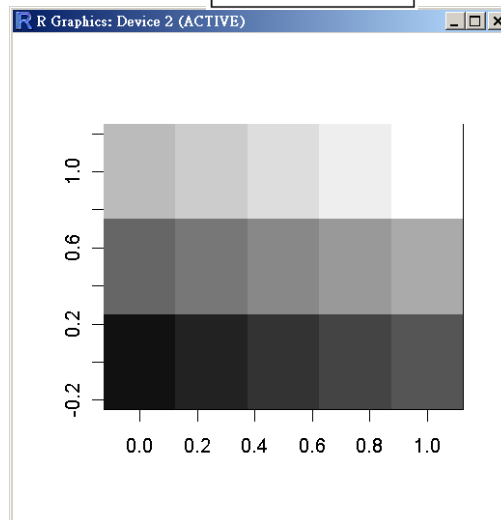


Image



```
> my.data <- matrix(c(1:15), ncol=3, nrow=5)
> my.data
> image(my.data, col=grey(1:15/15))
> image(t(my.data)[,nrow(my.data):1],
col=grey(1:15/15))
```

```
> my.data
  [,1] [,2] [,3]
[1,]  1   6  11
[2,]  2   7  12
[3,]  3   8  13
[4,]  4   9  14
[5,]  5  10  15
```

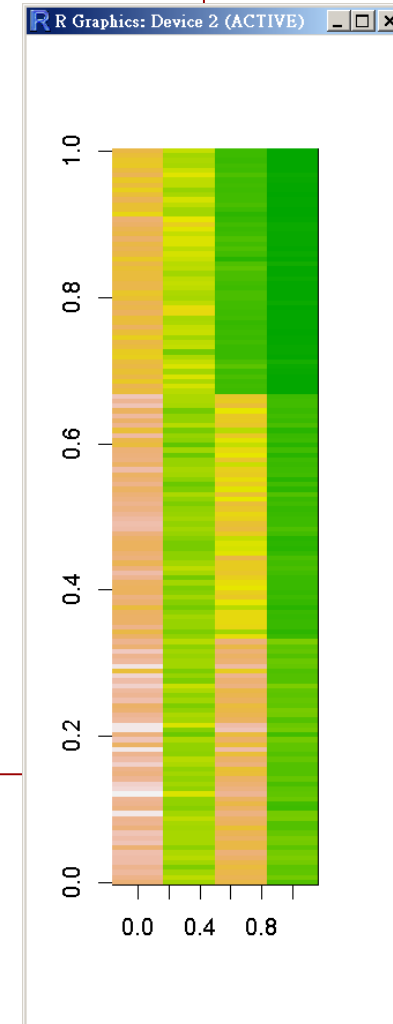



```
> image(t(as.matrix(iris[,1:4]))[,150:1], col=terrain.colors(100))
> head(iris[,1:4])
```

	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width
1	5.1	3.5	1.4	0.2
2	4.9	3.0	1.4	0.2
3	4.7	3.2	1.3	0.2
4	4.6	3.1	1.5	0.2
5	5.0	3.6	1.4	0.2
6	5.4	3.9	1.7	0.4

```
> tail(iris[,1:4])
```

	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width
145	6.7	3.3	5.7	2.5
146	6.7	3.0	5.2	2.3
147	6.3	2.5	5.0	1.9
148	6.5	3.0	5.2	2.0
149	6.2	3.4	5.4	2.3
150	5.9	3.0	5.1	1.8





pheatmap: Pretty Heatmaps

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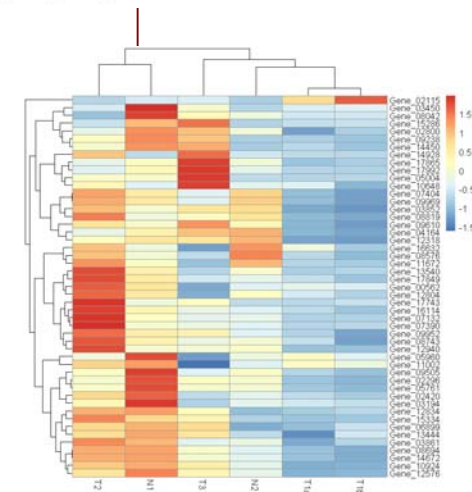
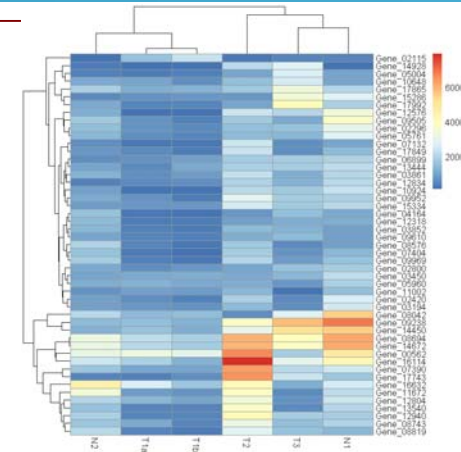
Implementation of heatmaps that offers more control over dimensions and appearance.

選讀

```
> library(pheatmap)
> DESeq_subset <- read.csv("DESeq_subset.csv")
> dim(DESeq_subset)
[1] 49 7
> head(DESeq_subset)
      X  T1a  T1b  T2  T3  N1  N2
1 Gene_00562 32314 29693 66140 17973 47994 30878
2 Gene_02115 15261 23301 1944 4578 4087 1072
...
6 Gene_03194 7611 6806 13506 5727 25020 9235
> DESeq.X <- as.matrix(DESeq_subset[,2:ncol(DESeq_subset)])
> colnames(DESeq.X)
[1] "T1a" "T1b" "T2" "T3" "N1" "N2"
> rownames(DESeq.X) <- DESeq_subset[,1]
> dimnames(DESeq.X)
[[1]]
 [1] "Gene_00562" "Gene_02115" "Gene_02296" "Gene_02420" ...
[46] "Gene_17743" "Gene_17849" "Gene_17865" "Gene_17992"

[[2]]
[1] "T1a" "T1b" "T2" "T3" "N1" "N2"

> str(DESeq.X)
int [1:49, 1:6] 32314 15261 6730 ...
- attr(*, "dimnames")=List of 2
 ..$ : chr [1:49] "Gene_00562"...
 ..$ : chr [1:6] "T1a" "T1b" "T2" "T3" ...
> pheatmap(DESeq.X)
```



```
> DESeq.X.std <- t(apply(DESeq.X, 1, scale))
> class(DESeq.X.std)a
[1] "matrix"
> dimnames(DESeq.X.std) <- dimnames(DESeq.X)
> pheatmap(DESeq.X.std) # note the color spectrum
```

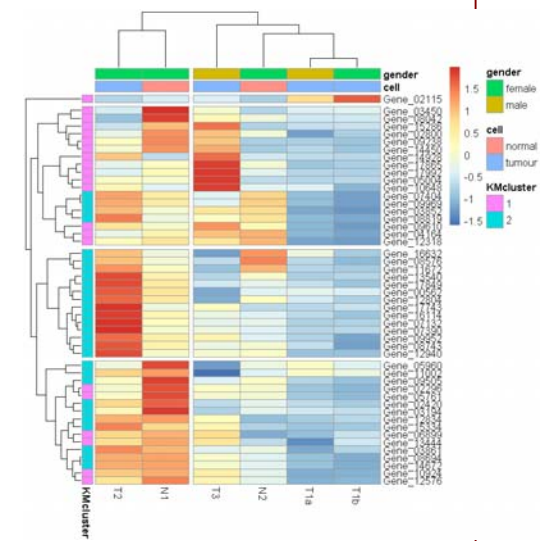
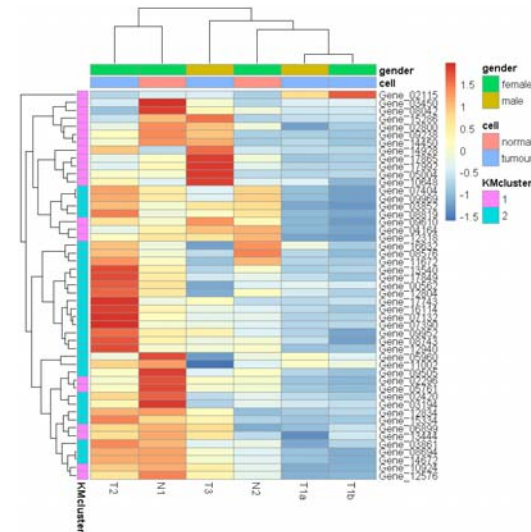
Making a heatmap in R with the pheatmap package

<https://davetang.org/muse/2018/05/15/making-a-heatmap-in-r-with-the-pheatmap-package/>

```

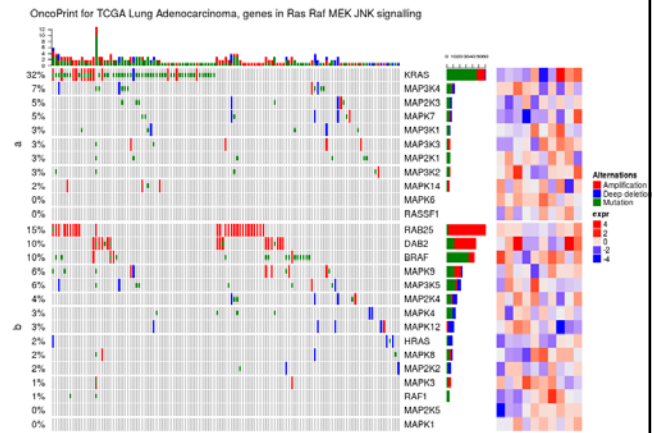
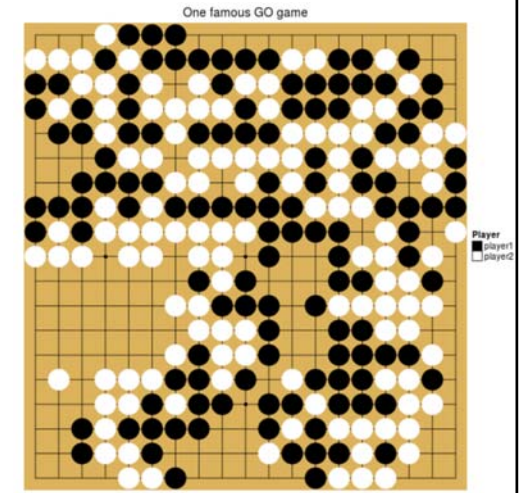
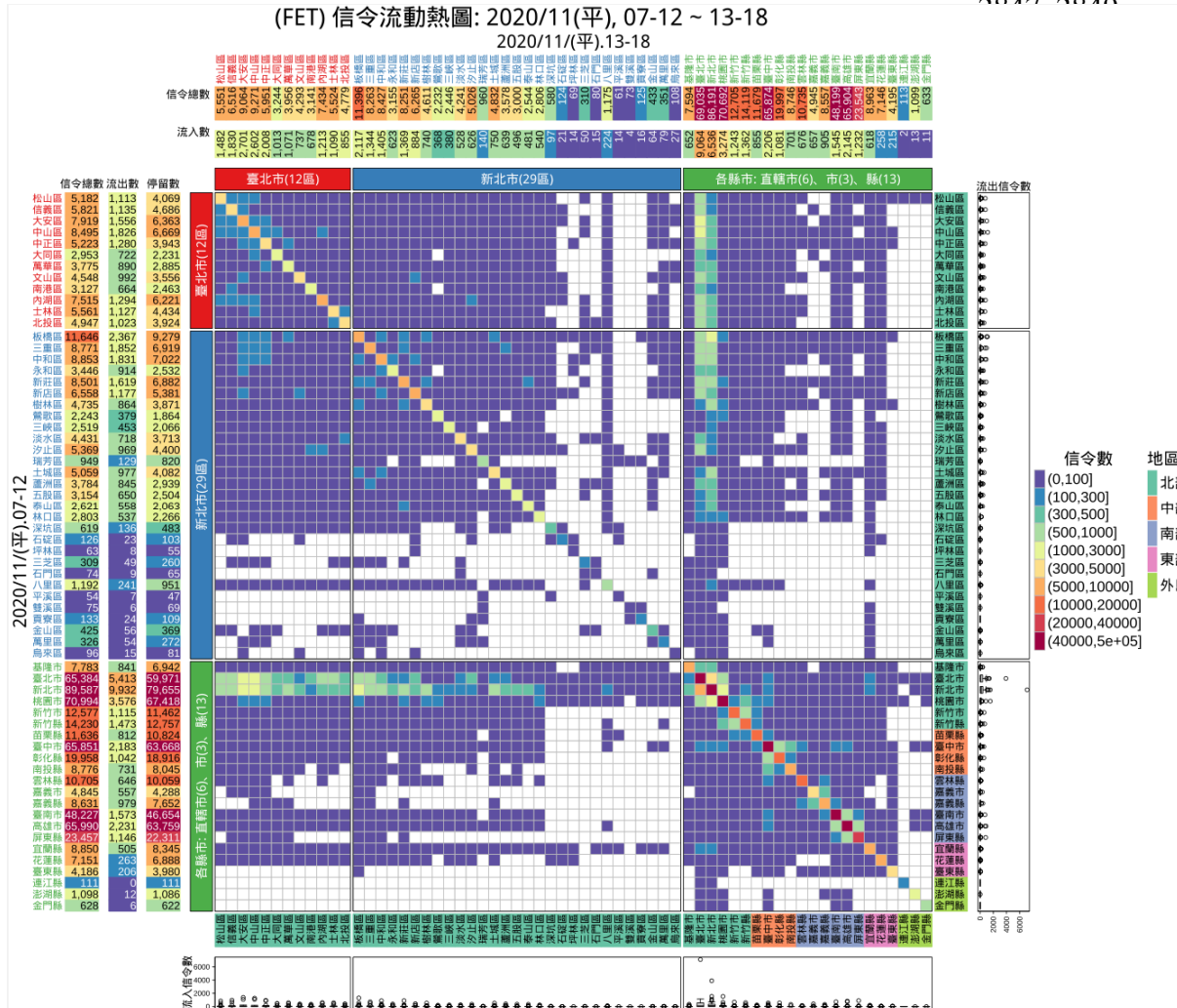
> sample.group <- data.frame(cell = rep(c("tumour", "normal"), c(4,2)),
+                             gender = sample(c("male", "female"), 6, replace=T))
> row.names(sample.group) <- colnames(DESeq.X)
> sample.group
      cell gender
T1a tumour  male
T1b tumour female
T2  tumour female
T3  tumour  male
N1  normal female
N2  normal female
> km <- as.character(kmeans(DESeq.X.std, 2)$cluster)
> gene.cluster <- data.frame(KMcluster = km)
> row.names(gene.cluster) <- rownames(DESeq.X)
> head(gene.cluster)
      KMcluster
Gene_00562      2
Gene_02115      1
Gene_02296      1
Gene_02420      2
Gene_02800      1
Gene_03194      2
>
> pheatmap(DESeq.X.std,
+          annotation_row = gene.cluster,
+          annotation_col = sample.group)
>
> pheatmap(DESeq.X.std,
+          annotation_row = gene.cluster,
+          annotation_col = sample.group,
+          cutree_rows = 4,
+          cutree_cols = 2)

```



<https://jokergoo.github.io/ComplexHeatmap-reference/book/>
<http://bioconductor.org/packages/release/bioc/html/ComplexHeatmap.html>

Zuguang Gu, Roland Eils, Matthias Schlesner, Complex heatmaps reveal patterns and correlations in multidimensional genomic data, Bioinformatics, Volume 32, Issue 18, 15 September 2016, Pages 2847-2850




visualize multiple genomic alteration events by heatmap


D3 JavaScript Network Graphs from R

<https://christophergandrud.github.io/networkD3/>


networkD3:

D3 JavaScript Network Graphs from R

CRAN 0.4 Dev-version: 0.4 Fork me on 

Christopher Gandrud, JJ Allaire, Kent Russell, & CJ Yetman Issues/suggestions 

2017-03-18



About

This started as a port of Christopher Gandrud's R package [d3Network](#) for creating [D3](#) network graphs to the [htmlwidgets](#) framework. The htmlwidgets framework greatly simplifies the package's syntax for exporting the graphs, improves integration with [RStudio's](#) Viewer Pane, [RMarkdown](#), and [Shiny web apps](#). See [below](#) for examples.

It currently supports the following types of network graphs:

- Force directed networks with [simpleNetwork](#) and [forceNetwork](#)
- Sankey diagrams with [sankeyNetwork](#)
- Radial networks with [radialNetwork](#)
- Dendro networks with [dendroNetwork](#)

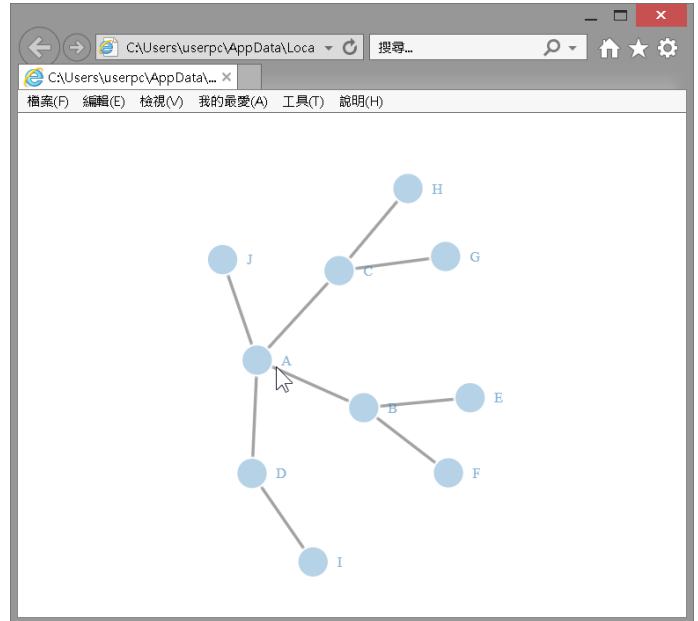
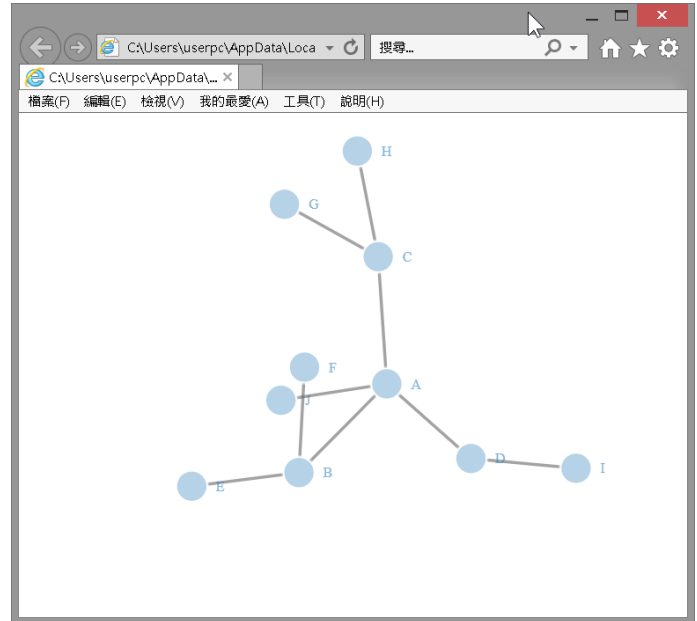
- **networkD3** supports the following types of network graphs:
 - Force directed networks with **simpleNetwork** and **forceNetwork**
 - Sankey diagrams with **sankeyNetwork**
 - Radial networks with **radialNetwork**
 - Dendro networks with **dendroNetwork**

```

> library(networkD3)
> src <- c("A", "A", "A", "A", "B", "B", "C", "C", "D")
> target <- c("B", "C", "D", "J", "E", "F", "G", "H", "I")
> networkData <- data.frame(src, target)
> simpleNetwork(networkData)
  
```

```

> networkData
src target
1 A B
2 A C
3 A D
4 A J
5 B E
6 B F
7 C G
8 C H
9 D I
  
```



桑基圖 (Sankey Diagram)

<http://www.sankey-diagrams.com/>

Google Sankey diagram

全部 圖片 影片 新聞 地圖 更多 設定 工具 查看已儲存的圖片 安全搜尋

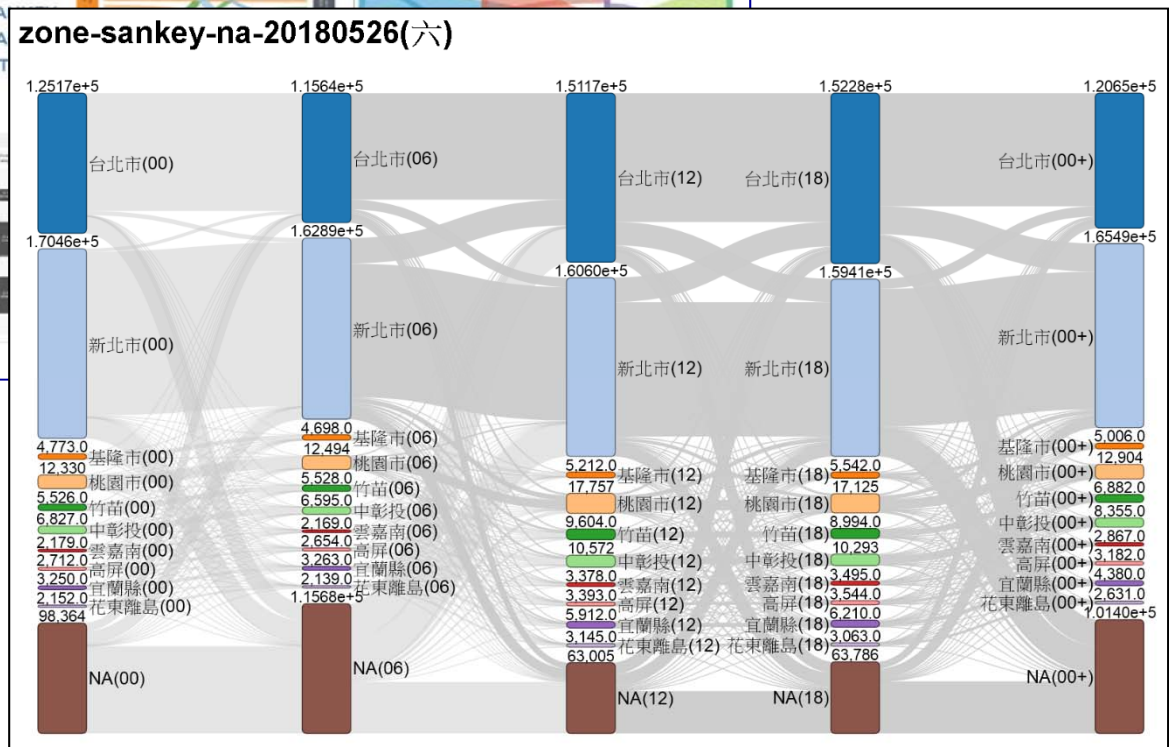
tableau python oil generator flow energy ktoc diagram maker

Arrows point to flow direction
Line width = Flow amount

Light Energy 10 J
Electrical Energy 100 J
Heat Energy 90 J

SA
DIA
TUT

zone-sankey-na-20180526(六)





Create a D3 JavaScript Sankey diagram

Usage

```
sankeyNetwork(Links, Nodes, Source, Target, Value, NodeID, NodeGroup = NodeID,  
  LinkGroup = NULL, units = "",  
  colourScale = JS("d3.scaleOrdinal(d3.schemeCategory20);"), fontSize = 7,  
  fontFamily = NULL, nodeWidth = 15, nodePadding = 10, margin = NULL,  
  height = NULL, width = NULL, iterations = 32, sinksRight = TRUE)
```

Arguments

- **Links**: a data frame object with the links between the nodes. It should include the Source and Target for each link. An optional Value variable can be included to specify how close the nodes are to one another.
- **Nodes**: a data frame containing the node id and properties of the nodes. If no ID is specified then the nodes must be in the same order as the Source variable column in the Links data frame. Currently only grouping variable is allowed.
- **Source**: character string naming the network source variable in the Links data frame.
- **Target**: character string naming the network target variable in the Links data frame.
- **Value**: character string naming the variable in the Links data frame for how far away the nodes are from one another.
- **NodeID**: character string specifying the node IDs in the Nodes. data frame. Must be 0-indexed.
- **NodeGroup**: character string specifying the node groups in the Nodes. Used to color the nodes in the network.
- **LinkGroup**: character string specifying the groups in the Links. Used to color the links in the network.



sankeyNetwork {networkD3}: Create a D3 JavaScript Sankey diagram

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```
> URL <- paste0(
+   "https://cdn.rawgit.com/christophergandrud/networkD3/",
+   "master/JSONdata/energy.json")
> Energy <- jsonlite::fromJSON(URL)
> str(Energy)
List of 2
 $ nodes:'data.frame': 48 obs. of 1 variable:
  ..$ name: chr [1:48] "Agricultural 'waste'" "Bio-conversion" "Liquid" "Losses" ...
 $ links:'data.frame': 68 obs. of 3 variables:
  ..$ source: int [1:68] 0 1 1 1 1 6 7 8 10 9 ...
  ..$ target: int [1:68] 1 2 3 4 5 2 4 9 9 4 ...
  ..$ value : num [1:68] 124.729 0.597 26.862 280.322 81.144 ...
```

```
> lapply(Energy, head)
```

\$nodes

	name
1	Agricultural 'waste'
2	Bio-conversion
3	Liquid
4	Losses
5	Solid
6	Gas

\$links

	source	target	value
1	0	1	124.729
2	1	2	0.597
3	1	3	26.862
4	1	4	280.322
5	1	5	81.144
6	6	2	35.000

```
> sankeyNetwork(Links = Energy$links, Nodes = Energy$nodes,
+   Source = "source", Target = "target",
+   Value = "value", NodeID = "name",
+   fontSize = 12, nodeWidth = 30)
```

sankeyNetwork

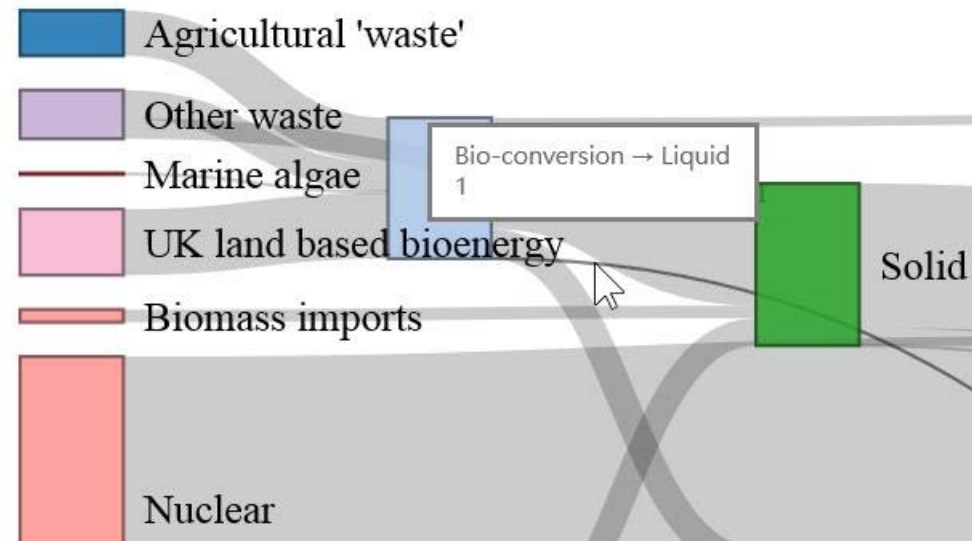
```
> Energy
```

```
$nodes
```

	name
1	Agricultural 'waste'
2	Bio-conversion
3	Liquid
4	Losses
5	Solid
6	Gas
7	Biofuel imports
8	Biomass imports
...	
46	UK land based bioenergy
47	Wave
48	Wind

```
$links
```

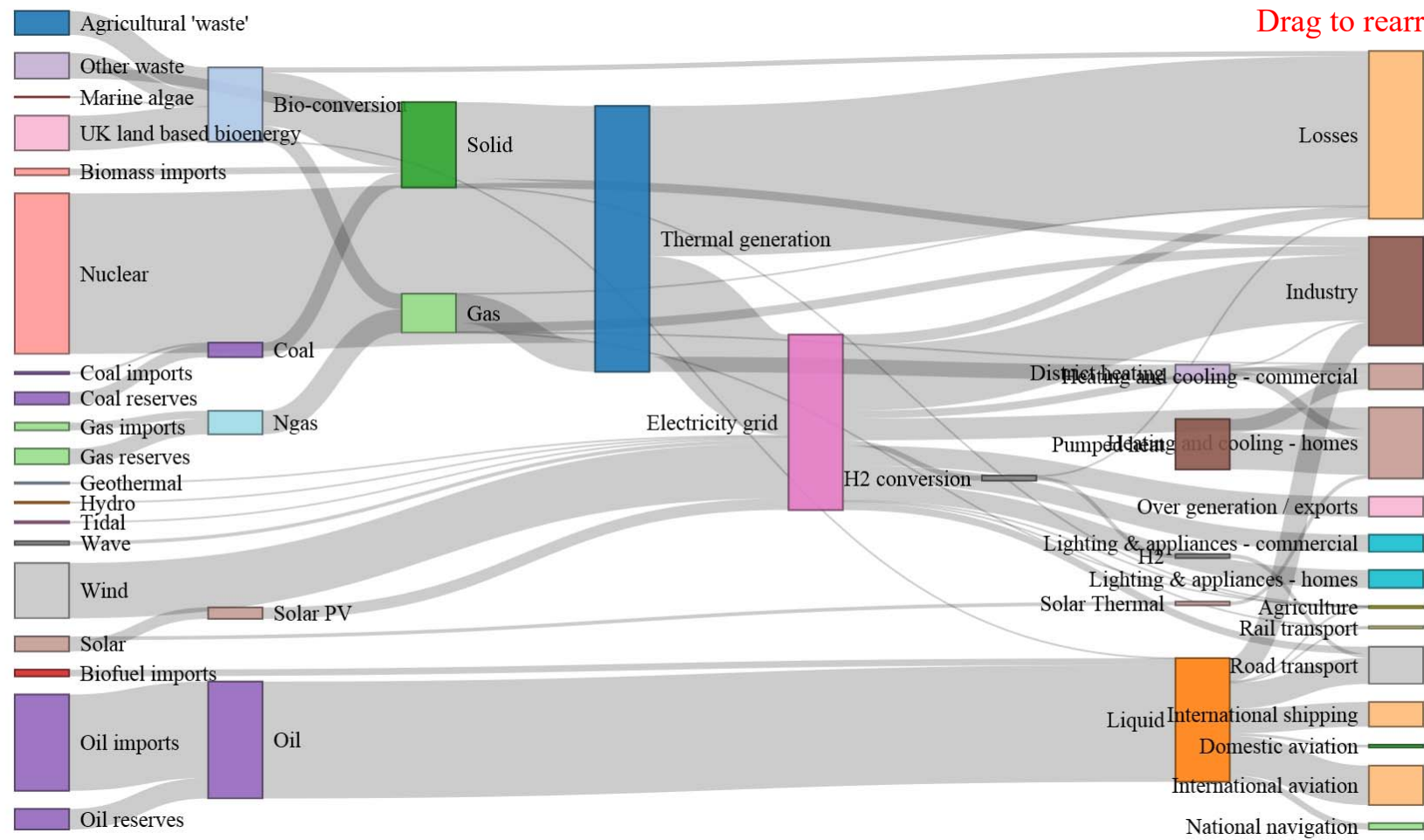
	source	target	value
1	0	1	124.729
2	1	2	0.597
3	1	3	26.862
4	1	4	280.322
5	1	5	81.144
6	6	2	35.000
7	7	4	35.000
...			
67	46	15	19.013
68	47	15	289.366



Sankey diagrams are closely related to alluvial diagrams (沖積圖), which show how network structure changes over time. https://en.wikipedia.org/wiki/Alluvial_diagram

sankeyNetwork

```
> sankeyNetwork(Links = Energy$links, Nodes = Energy$nodes,
+               Source = "source", Target = "target",
+               Value = "value", NodeID = "name",
+               fontSize = 12, nodeWidth = 30)
```



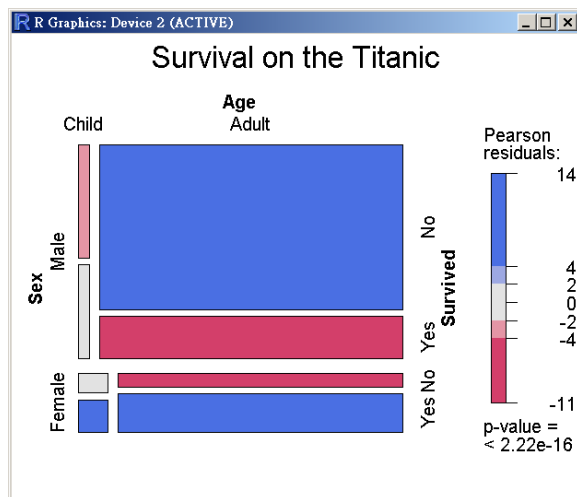
Drag to rearrange nodes.

- **vcd**: Visualizing Categorical Data
- Visualizing Categorical Data with SAS and R: <http://www.datavis.ca/courses/VCD/>
- Visualizing Categorical Data: <http://www.datavis.ca/books/vcd/>

```
> library(vcd)
vcd: Visualizing Categorical Data
http://cran.r-project.org/web/packages/vcd/index.html
```

- Fourfold Display for 2x2 Tables
- Association Plots
- Mosaic Display

Mosaic Displays



Fourfold Display

